# Analysis of some virulence factors of *Vibrio vulnificus* isolated from Rio de Janeiro, Brazil

# D. P. RODRIGUES, R. V. RIBEIRO AND E. HOFER

Dalia dos Prazeres Rodrigues, Fundação Oswaldo Cruz, Av. Brasil 4365, Pavilhão Rocha Lima, Dpto Bacteriologia Manguinhos, Rio de Janeiro, Brazil, CEP 21045

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#### SUMMARY

Twenty strains of *V. vulnificus* isolated from the environment were investigated for characteristics related to their infectivity such as colonial morphology, enzymatic activity and animal assays. The presence of DNase, chitinase, amylase, lecithinase and gelatinase was observed in 100% of the strains, haemolytic activity was absent, and variable results were obtained in elastase, collagenase and chondroitinase. In the animal assays, 70% of the strains were lethal to adult mice, while 45% caused fluid accumulation in suckling mice. Although all strains had opaque colonies, only 3 of the 20 had the three enzymes elastase, collagenase and gelatinase, and only one of these was virulent in animal assays.

## INTRODUCTION

Vibrio vulnificus is the most invasive species of the genus Vibrio, with three different clinical syndromes [1]. The first involves progressive infection with a few diarrhoeal symptoms and is characterized by a rapid outset of fulminating septicaemia followed by the appearance of cutaneous lesions with 50% mortality. Patients with hepatic dysfunctions or other syndromes involving iron metabolism appear to be most susceptible to this type of infection [2]. The second, represented by cellulitis, results from the direct contact of wounds or skin lesions with sea ecosystem constituents in both apparently healthy and debilitated patients, and sometimes progresses into septicaemia [1, 3]. The third form, seldom found, causes acute self-limited diarrhoea with a low mortality rate [4].

Studies on the pathogenicity of this microorganism demonstrate that, in strains isolated from clinical cases, there is a strict relationship between the presence of a polysaccharide capsule-like structure and virulence [5]. Other factors have also been reported including the presence of some siderophore-like proteins in the membrane [6], the production of enzymes that damage the surrounding tissues [7, 8], and various toxins acting like invasive factors in acute infections [1, 3]. An interesting point is that some factors were detected in environmental strains as well as in strains of human origin [9].

In this paper, biochemical characteristics as well as potential virulence factors have been investigated in strains of V. vulnificus isolated from sea water and oysters from the coast of the state of Rio de Janeiro.

### MATERIALS AND METHODS

# Samples

Two types of samples were used. The first (15 strains) was isolated from sea water from the coast of Rio de Janeiro. The second (5 strains) was from oysters from natural breeding areas found in the Sepetiba Bay (Rio de Janeiro State). The cultures were maintained at room temperature in buffered nutrient agar with 1 % NaCl [10].

# Biochemical characteristics

The biochemical tests were performed as previously described [11, 12]. Colonial morphology was examined on nutrient agar (Difco) and brain heart infusion agar (Difco), using the criteria of Simpson and co-workers [5].

# Enzymatic assays

Published techniques [12–14] were used in the investigation of amylase, gelatinase, elastase, collagenase, chondroitinase, lecithinase, DNase and haemolysin.

# $Animal\ assay$

Each strain was inoculated into five adult and five suckling mice. Adult animals, weighing approximately 20g each, were inoculated intraperitoneally with 0·1 ml of an 18 h culture which had been incubated at 37 °C (about 10<sup>8</sup> cells/ml) in BHI Broth (Merck), supplemented with additional 2·5 % NaCl, and mortality observed until 18 h after inoculation [15].

Suckling mice (3–5 days old) received 0·1 ml of supernatant from an 18 h growth in heart infusion broth (Difco), obtained after centrifugation at 10000 g at 4 °C for 30 min, with 0·01 g% of Evans Blue. Mice were inoculated intragastrically by catheter. After 4 h the toxin activity was determined by a radio between the weight of guts and the remaining body [15].

## RESULTS

The biochemical characteristics of *V. vulnificus* isolated from sea water and oysters were in accordance with those reported in the literature [12, 17].

When cultures were inoculated on to nutrient agar and BHI agar we observed either mixtures of opaque and translucent colonies, or opaque colonies into both media with an excellent contrast and sharp differentiation between the colony types (Table 1).

All 20 strains produced DNase, chitinase, amylase, lecithinase and gelatinase, but no haemolytic activity of sheep erythrocytes was seen. Variable results were obtained for elastase, collagenase and chondroitinase (Table 2).

The animal assays (Table 3) showed that 70% of the strains were lethal for adult mice, while 45% caused intestinal fluid accumulation in suckling mice.

Only one strain had enzyme profile D (with the production of collagenase, gelatinase and elastase), and was pathogenic to both groups of mice.

Table 1. Colony morphology in relation to the culture media

Culture	media	
Nutrient agar	Heart infusion agar	Number of strains
O* T-O T-O	0 T-O 0	7 6 5
O Total	T-O	$\frac{2}{20}$

<sup>\*</sup> O, opaque colonies; T-O, opaque and translucent colonies.

Table 2. Distribution of the strains according to enzymatic profile

		Enzymatic profile				
Fnzymos	$ \begin{array}{c} A \\ (n=7) \end{array} $	$ \begin{array}{c} B\\ (n=7) \end{array} $	$C \\ (n=3)$	$ \begin{array}{c} \text{D} \\ (n=3) \end{array} $		
Enzymes	(n-1)	(n-1)	(n-3)	(n-3)		
Amylase DNase	+	+	+	+		
Gelatinase	+	+	+	+		
Lecithinase	+	+	+	+		
Chitinase	+	+	+	+		
Elastase	_	+	_	+		
Collagenase	_	_	_	+		
Chondroitinase	_	_	+	_		
Haemolysin	_	_	_	_		

Table 3. Correlation between enzymatic profile and tests for biological activity

Enzymatie	Percentage distribution of animal assays				
profile*	$\overline{AM + SM + \dagger}$	AM + SM -	AM - SM +	AM-SM-	
A/C	25	15	5	5	
$\mathbf{B}$	10	15	_	10	
D	5	_	_	10	
Total	40	30	5	25	

<sup>\*</sup> Enzymatic profile, Table 3.

## DISCUSSION

Ecological studies show that V. vulnificus, like all the other species of the genus Vibrio, is a normal constituent of the marine environment [18]. Its occurrence is determined by environmental factors such as salinity and temperature [8, 19]. It is found in high frequency in sea water and sea food, which contrasts with the small number of clinical cases reported worldwide. This indicates a large variation in the pattern of virulence among the strains; this is similar to V. parahaemolyticus, in which only 1% of the strains found in the aquatic environment is pathogenic for man [3].

Patients with pre-existing liver or blood disorders, or a history of alcohol abuse,

<sup>†</sup> AM, lethal for adult mice; SM, fluid accumulation in suckling mice.

are more susceptible to infection with V. vulnificus, which results in elevated serum iron levels; the resistance of V. vulnificus to killing by human serum is mediated by the presence of excess iron [2].

The evaluations carried out by West [1] indicated that strains of V. vulnificus are more resistant to lysis by complement than are V. parahaemolyticus and V. cholerae, and that during invasion it can activate both the classic and alternate pathways [2].

Another characteristic that confers resistance to the bactericidal effects of human serum is the polysaccharide capsule, recognized by Simpson and coworkers [5] in strains isolated from clinical and environmental material. In the laboratory, the presence of capsule is recognized by colonial morphology as capsulated strains form opaque colonies; translucent colonies result when this structure has been lost. This phenomenon was observed in all strains in both culture media; moreover the preservation of the strains for periods from 1 to 4 years in buffered nutrient agar did not alter this characteristic. This result is in contrast to that observed by Simpson and colleagues [5], who reported the transformation of opaque colonies into translucent ones in more than 60% of samples kept in maintenance medium.

The invasive capacity and resulting tissue damage suggest the role of toxins or components of enzymatic nature (e.g. collagenase, gelatinase, elastase), or the action of compounds with cytolytic and proteolytic activities. In our strains we found four enzymatic profiles with variation in elastase, collagenase and chondroitinase. The presence of elastase was detected in  $50\,\%$  of strains, collagenase in  $15\,\%$  and chondroitinase in  $15\,\%$ ; in only  $15\,\%$  of strains were both collagenase and elastase detected.

However, there was an inverse relationship between pathogenicity for mice and production of elastase and collagenase, although every strain presented opaque colonies that suggested the presence of a polysaccharide capsule.

The mortality among our animals (70%) was greater than that reported by Kaysner and co-workers [8], of 60% mortality within 72 h, and less than that reported by Tilson and Kelly [9] in an analysis of 29 environmental strains, of which 25 were pathogenic (86%). This difference may be a reflection of the fact that their experiments were carried out immediately after isolation of the microorganism, and that observation of the animals was prolonged for 24 h instead of the 18 h period adopted by us.

Virulence has also been ascribed to the production of enterotoxin. types LT or ST, observed by the accumulation of intestinal fluid in suckling mice, usually detected in the O 1 and non-O 1 strains of *Vibrio cholerae*, *V. fluvialis* and *V. hollisae*. In *V. vulnificus* the activity of these substances depends on the presence of a bacterial envelope. We obtained extremely variable results when using this model (Table 3), but these are in agreement with those obtained by Bowdre and colleagues [15]. It may be that these findings are related to the existence of other toxic substances or enzymatic factors.

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### REFERENCES

- 1. West PA. The human pathogenic vibrio a public health update with environmental perspectives. Epidemiol Infect 1989; 103: 1–34.
- 2. Janda JM, Powers C, Bryant RG, Abbott SL. Current perspectives on the epidemiology and pathogenesis of clinically significant *Vibrio* spp. Clin Microbiol Rev 1988; 1: 245–67.
- 3. Morris JG. Black RE. Cholera and other vibriosis in the United States. New Engl J Med 1985; **315**: 343–50.
- 4. Klontz KC, Lieb S, Schreiber M, Janowski HT, Baldry LM, Gunn RA, Syndromes of *Vibrio vulnificus* infections: clinical and epidemiologic features in Florida cases, 1981–1987. Ann Int Medicine 1988: 109: 318-23.
- Simpson LM, White VK, Zane SF, Oliver JD. Correlation between virulence and colony morphology in V. vulnificus. Infect Immun 1987; 55: 269-72.
- 6. Gander RM. La Rocco M. Detection of piluslike structures on clinical and environmental isolates of *Vibrio vulnificus*. J Clin Microbiol 1989; 27: 1015-21.
- 7. Smith CG, Merkel RJ. Collagenolytic activity of *Vibrio vulnificus*. Potential contribution to its invasiveness. Infect Immun 1982; **35**: 1155-7.
- 8. Kaysner LA. Wekell MM. JR CA, Stott RF, Leitch JM. Virulent strains of *V. vulnificus* isolated from estuaries of the United States west coast. Appl Environ Microbiol 1987; **53**: 1349–51.
- Tison DL, Kelly MT. Virulence of V. vulnificus strains from marine environments. N Engl J Med 1986; 51: 1004-6.
- Hofer E. Métodos utilizados para o isolamento e identificação de Vibrio cholerae. Inf Pat Clínica 1975; 1: 5–18.
- Furniss AL, Lee JV, Donovan TJ. The vibrios. London: Public Health Laboratory Service, Monograph Series, 1979: 1–58.
- 12. West PA, Colwell RR. Identification and classification of *Vibrionaceae* an overview. In: Colwell R. R., ed., Vibrios in the environment. New York: John Wiley & Sons, 1984: 285–363.
- Thorpe P. Miller B. Extracellular enzymes of Legionella pneumophila. Infect Immun 1981;
   33: 632-5.
- Rodrigues DP, Hofer E. Vibrio species from the water-oyster ecosystem of Sepetiba Bay in Rio de Janeiro State, Brazil. Rev Microbiol (São Paulo) 1986; 17: 332-8.
- Bowdre JH. Poole MD. Oliver JD. Edema and hemoconcentration in mice experimentally infected with V. vulnificus. Infect Immun 1981; 32: 1193-9.
- Nishibuchi M, Seidler RJ. Medium-dependent production of extracellular enterotoxins by non O1 Vibrio cholerae. V. mimicus and V. fluvialis. Appl Environ Microbiol 1983; 45: 228-31
- Baumann P, Schubert RHW. Family II Vibrionaceae. In: Krieg NR, Holt JG, eds. Bergey's manual of systematic bacteriology, vol. 1. Baltimore: The Wilkins Co., 1984: 516–50.
- 18. Colwell RR. Vibrios in the environment. New York: John Wiley & Sons, 1984: 1-12.
- 19. Kelly MT. Effect of temperature and salinity on Vibrio (Beneckea) vulnificus occurrence in Gulf Coast estuaries. Appl Environ Microbiol 1982; 44: 820–4.
- 20. Kreger A. Lockwood D. Detection of extracellular toxin(s) produced by V. vulnificus. Infect Immun 1981; 33: 583–90.
- 21. Gray LD, Kreger AS. Purification and characterization of an extracellular cytolysin produced by *V. vulnificus*. Infect Immun 1985; **48**: 62-72.
- 22. Kothary MH. Kreger AS. Purification and characterization of an elastolytic protease of V. vulnificus. J Gen Microbiol 1987: 133: 1783–91.