# Does infusion time affect the retention of parenteral trace elements?

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# **Abstract**

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When readymade parenteral nutrition in multi-chamber bags is supplied without vitamins and minerals, these have to be added or given separately. Separate rapid infusion has logistical advantages but has been claimed to saturate tissue mechanisms, potentially increasing urine micronutrient losses. The present study compared urinary losses after fast (1 h) v. slow (10 h) trace elements infusion. The study enrolled thirty-nine consecutive patients who were starting parenteral nutrition postoperatively. One day's dose of a complete intravenous micronutrient product was infused over 1 h and over 10 h, in random order, with a washout day after each infusion day. Urinary Zn, Mn, Se, Cr, Cu and Fe losses were measured by 24-h urine collection, recorded for each infusion day and subsequent washout day. For Zn, a dose of 100 μmol was given, and total urinary loss over the next 2 d was mean 40.6 (sp 23.8) µmol after the fast (1 h) infusion v. 33.4 (sp 25.4) µmol after the slow (10 h) infusion, that is, 7% more of the 1-d dose was lost after fast infusion (P = 0.01). For Mn, after a dose of 1000 nmol, losses were 9.8 (sp 23.9) nmol after the fast infusion v. 22·1 (sp 47·2) nmol after the slow infusion, that is, 1% more of the 1-d dose was lost after slow infusion (P = 0.04). There were no other significant differences: after 1 μmol Se, the losses were 1.5 (sd 0.6) μmol fast v. 1.3 (sd 0.5) μmol slow; after 200 nmol Cr, 257 (sd 92) μmol fast v. 246 (sp 107) nmol slow; after 8 μmol Cu, 1·6 (sp 1·4) μmol fast v. 1·5 (sp 1·3) μmol slow; and after 20 μmol Fe, 0·6 (sp 1·1) μmol fast v. 0·8 (sp 1·6)  $\mu$ mol slow (P > 0.05 for all). Overall, trace element retention appears to be minimally affected by infusion time.

Key words: Parenteral nutrition: Trace elements: Renal elimination

Readymade parenteral nutrition (PN) products are typically presented without vitamins and trace elements. For patients receiving readymade PN, the trace elements and water-soluble and fat-soluble vitamins are either added to the PN bag or infused separately as recommended by the British Pharmaceutical Nutrition Group<sup>(1)</sup>. When given as a separate intravenous (IV) infusion in a small volume of fluid, there is significant variation in practice regarding infusion time. When both vitamins and trace elements are combined into a small volume in this way, there is potential for them to interact, leading to a loss of vitamin activity<sup>(2)</sup>; so if it is not possible to give vitamins separate from the trace elements<sup>(3)</sup>, then a fast infusion might be preferred. A contrasting view is that a fast infusion could lead to the saturation of tissue transport mechanisms, resulting in increased urinary losses, as has been seen in studies where very large doses of Cu, Se and Zn were used<sup>(4)</sup>. It is unclear whether this phenomenon occurs with the small doses used for meeting patients' daily requirements as part of PN.

Cu, given intravenously as copper chloride, is mostly excreted via the biliary system rather than the urine. IV Cu is rapidly bound to albumin for transport to the liver, where there are large stores, or it is incorporated into caeruloplasmin with daily turnover of about 0.5 mg. Urinary excretion does appear to increase with high doses, but the total urinary loss is still quite low. For example, one study gave 4–9 µg of Cu as a fast IV injection, and <1% of the dose appeared in the urine over the next 72 h<sup>(5)</sup>.

Similarly, Mn has mostly biliary excretion. Mn, given intravenously as manganese sulphate or manganese chloride, rapidly disappears from the plasma and is absorbed into the tissue. Mn turnover rates vary widely (from days to weeks) between individuals<sup>(6)</sup>. About 40 % of the dose is lost via the biliary system, but this varies with Mn status<sup>(7)</sup>. Only small amounts of Mn are excreted in the urine, and this does not appear to be affected by dose or infusion time.

In contrast, the main route by which Se is lost from the body is via the urine. Se is given intravenously as sodium selenite and has a biphasic action with brief pro-oxidant effects from free Se, transitioning to an antioxidant effect after incorporation into selenoenzymes such as glutathione peroxidase<sup>(8)</sup>; Se is stored in erythrocytes, liver, spleen and heart. Less than 1% of plasma Se is present as free Se; about 10% is carried by albumin<sup>(9)</sup> and the remainder is protein-bound such as in

Abbreviations: ICU, intensive care unit; IV, intravenous; PN, parenteral nutrition.

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selenoenzymes. In one animal study, urinary losses increased after a rapid IV bolus but not during continuous infusion<sup>(10)</sup>. Recent human studies using large pharmacological doses of Se in the critically ill did not measure urinary losses but found that large doses had little effect on blood Se levels(11).

Urinary losses of Zn may depend on the dose and on Zn status. Parenterally administered Zn, given as zinc chloride or zinc sulphate, is cleared rapidly (in hours) from the plasma with 400-500 mg turnover daily between plasma and tissues; <0.1 % of body Zn remains in the bloodstream<sup>(12)</sup>. The clearance rate appears to be determined by Zn status<sup>(13)</sup> with Zn deficiency causing an increase in turnover rate but a decrease in the amount of each turnover, which would then be expected to reduce urinary losses<sup>(14)</sup>. A rapid IV dose of 8 mg Zn (described as an 'overload dose') caused a 20% increase in 24-h urine Zn in one study(15). In diabetes, usual renal losses of Zn may be higher (e.g. double normal)(15).

Fe is not normally lost in the urine in significant amounts, but urinary losses of IV Fe depend on the form in which it is given, as the commonly used Fe-carbohydrate complexes vary in molecular weight. IV Fe-carbohydrate complex travels through the blood vessels of the liver, spleen and bone marrow where the phagocytes of the reticuloendothelial system remove the complex from the plasma in an apparently saturable process<sup>(16)</sup>. Within the phagocytes, Fe is removed from the core of the complex, and is taken up by ferritin for storage, or by transferrin. Larger-molecular-weight complexes (such as iron dextran) are more stable; the smaller the particle size, the less stable the preparation and the more rapid the release of Fe, which is undesirable as it can result in the saturation of transferrin, slower clearance from plasma and a sudden high dose of labile pro-oxidative Fe<sup>(17)</sup> with greater losses in the urine(16).

Similarly, Cr losses may depend on the form in which it is given. Cr turnover is slow, with prolonged tissue and bone retention(18). IV Cr easily binds to transferrin, taking up one of the Fe sites, and a large proportion of circulating Cr is found in this form. Stimulated by insulin, transferrin moves into cells via a transferrin receptor, where Cr becomes bound to a protein, chromodulin, which then returns to the bloodstream. Only chromodulin-bound Cr can be lost in the urine<sup>(19)</sup>. Perhaps 20% of an IV dose might be excreted in the urine, but the proportion lost in the biliary system depends on the form of Cr given<sup>(20)</sup>. Urinary Cr losses are greater in diabetes, insulin therapy and with any high carbohydrate load that stimulates extra insulin, since insulin then promotes Cr uptake to cells and binding to chromodulin<sup>(19)</sup>. Increasing transferrin saturation with Fe would also affect Cr status by competing with Cr for transport into the cell - thus, Cr losses might be affected by the form in which IV Fe is given.

Overall it appears that infusion time has a potential to affect urinary losses of Zn, Se and (in diabetes or insulin resistance) Cr, but this has not been studied in humans using the small daily dose used in everyday practice. The present study aimed to quantify urinary losses comparing a fast (1 h) v. a slow (10 h) infusion of a 1-d dose of six trace elements.

Table 1. Composition of the parenteral micronutrient additive used in the

Cernevit and CCADTEN in 100 ml 5 % dextrose (Baxter Australia)							
Cr Cu lodine Fe Mn Mo Se Zn	200 nmol (10 μg) 8 μmol (5·5 μg) 1 μmol (130 μg) 20 μmol (1·2 mg) 1000 nmol (55 μg) 0·2 μmol (19 μg) 1 μmol (80 μg) 100 μmol (6·5 mg)	Vitamin A Thiamine Riboflavin Niacin Dexpantothenic acid Pyridoxine Biotin Folic acid Vitamin B <sub>12</sub> Vitamin C Vitamin D <sub>3</sub> Vitamin E	1.05 mg 3.51 mg 4.14 mg 46 mg 17.25 mg 4.53 mg 69 µg 414 µg 6 µg 125 mg 5.5 µg 7.5 mg				

### Methods

#### Enrolment and consent

Patients starting PN in the intensive care unit (ICU) were screened for eligibility. Patients were excluded if they were malnourished (Subjective Global Assessment 'B' or 'C'), had impaired renal function (estimated glomerular filtration rate < 60), were under 18 years of age or pregnant or lactating. Each eligible patient (or a responsible relative) provided written consent to participate. The study was approved by the hospital's ethics review committee.

#### Intervention

A 1-d dose of parenteral vitamins and trace elements was used for each infusion following the Australasian Society for Parenteral and Enteral Nutrition recommendations for adults<sup>(21,22)</sup>. After the baseline day of enrolment, when no micronutrients were given, patients received a standard vitamin and trace elements bag (Cernevit and CCADTEN in 100 ml 5% dextrose; Baxter Australia; see Table 1 for composition) over the two infusion times (1 v. 10 h) in random order on two different days with a washout day after each dose; see Fig. 1 for study procedure timeline. A 24-h urine collection, in acid-washed preservativefree bottles, commenced on the day of enrolment and continued for 5 d. Blood samples were obtained each morning as part of routine ICU care. Urine and blood trace element levels were tested each day of the study in the hospital's central laboratory using inductively coupled plasma MS. For each patient, factors that can affect trace element absorption, metabolism or excretion were recorded such as diabetes, diarrhoea or a concurrent prescription of diuretics, insulin, phenytoin or antibiotics.

# **Statistics**

Regression analysis within the framework of generalised estimating equations, using SPSS version 21 (IBM Australia) was used to compare the daily urinary losses between the two different infusion times and to identify whether there was a significant influence from the order in which each patient received the two infusions. For each trace element, the adjusted regression equations controlled for the covariates of the subjects' baseline trace element loss, and for the order in which each subject received



58 S. Ferrie

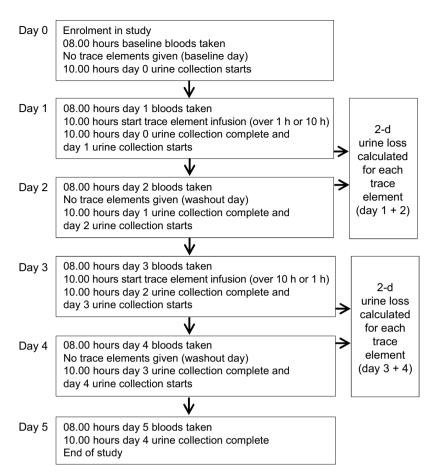


Fig. 1. Study procedure timeline.

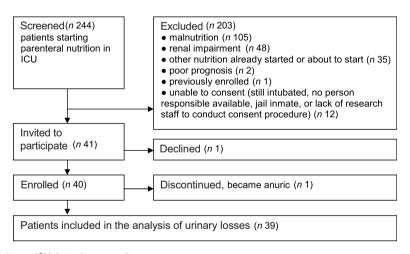


Fig. 2. Study screening and enrolment. ICU, intensive care unit.

the two infusions. A P value < 0.05 was considered significant for all tests. Data from a previous pilot study<sup>(23)</sup> indicated that for 90 % power, recruitment would require thirty-three subjects to detect a difference in daily Zn loss of 6 (sp 10) mg; twenty-four subjects to detect a difference for Se loss of 0.4 (sp 0.6)  $\mu$ g; and twenty-nine subjects to detect a difference for Cu loss of 0.2 (sp 0.4)  $\mu$ g. The study, therefore, aimed to

recruit forty people to allow for dropouts, missing values and sample losses.

## **Results**

Forty patients consented to participate in the study; one subsequently discontinued after becoming anuric (see Fig. 2).





Table 2. Study patient characteristics									
(Mean va	alues and standard deviations: numbers of	patients)							

	Received 1 h infusion first (n 19)		Received 10 h infusion first (n 20)		Total ( <i>n</i> 39)		
Patient characteristics	Mean	SD	Mean	SD	Mean	SD	
Age (years)	53.7	8.0	56.4	11.2	55.1	9.8	
Sex (n)							
Male	12	12		10		22	
Female	7		10		17		
BMI (kg/m <sup>2</sup> )	27.6	6.0	28.1	5.1	27.9	5.5	
Diabetes (n)	1		2		3		
Received insulin during study (n)	6		7		13		
Received diuretics during study (n)	0		0		0		
Liver disease (n)	0		0		0		
Current infection (n)	0		1		1		
Received antibiotics during study (n)	11		14		25		
Diarrhoea during study (n)	2		1		3		

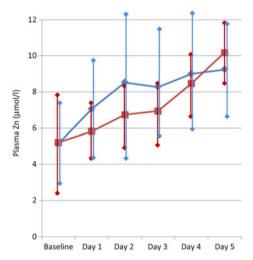


Fig. 3. Change in serum zinc over the 5-d study period, for the patients who received the 1 h infusion first (---) and for the patients who received the 10 h infusion first (---). Values are means, with standard deviations represented by vertical bars.

One further patient had incomplete urine collection, but this result was averaged over 24 h and included in the analysis. Table 2 shows the study patient characteristics.

There was a wide variation between individuals in trace element losses, as expected. For each individual patient, the baseline trace element losses were strongly predictive of the losses after each infusion. Blood levels of all trace elements increased during the 5 d of the study, and the increase was significant (P < 0.05) for Cu, Cr and Se, which all increased by 20-50% over the study period; and for Zn it increased about 80% over the study period; see Fig. 3. However, the order in which a patient received the two infusions had no statistically significant influence on the results (P > 0.05), suggesting that urinary losses were not strongly associated with the blood levels, and that one washout day was sufficient to show the total losses after each infusion.

Fig. 4 shows the baseline (1 d) and post-infusion (2 d) urinary losses of each trace element, compared with the dose given. For Zn, one day's requirement was provided as a dose of 100 µmol in each infusion, and urinary loss over the next 2 d was mean 40.6 (sp 23.8) µmol after the fast (1 h) infusion v. 33.4(sp 25.4)  $\mu$ mol after the slow (10 h) infusion (P = 0.01). Se, Cr and Cu also had trends towards greater losses with fast infusion, but these were not statistically significant (P > 0.05): for Se, one day's requirement of 1 µmol was given in each infusion, and urinary loss over the next 2 d was 1.5 (sp 0.6) µmol after fast infusion v. 1·3 (sd 0·5)  $\mu$ mol after slow infusion; for Cr, a dose of 200 nmol was given, and urinary losses were 257 (sp 92) nmol fast v. 246 (sd 107) nmol slow; for Cu, a dose of 8 µmol was given, and urinary losses were 1.6 (sD 1.4) fast v. 1.5 (sD 1.3)  $\mu$ mol slow.

For Mn, after a dose of 1000 nmol, losses were mean 9.8 (SD 23.9) nmol after fast infusion v. 22.1 (SD 47.2) nmol after slow infusion (P = 0.04). Fe also had a trend towards greater losses with slow infusion, but again this was not statistically significant (P > 0.05): a dose of 20  $\mu$ mol was given and urinary losses were 0.6 (sD 1.1) fast v. 0.8 (sD 1.6)  $\mu$ mol slow.

Notably, Zn and Cr urine losses appeared to be higher in patients with diabetes, although the small numbers mean this did not reach statistical significance. Compared with nondiabetic patients, urinary losses approximately doubled in diabetes at baseline and with fast infusion, but not with slow infusion.

# Discussion

After a dose of one day's requirement of trace elements, there was little difference in urinary trace element losses with a fast (1 h) infusion compared with a slow (10 h) infusion. Some trace elements had greater losses with fast infusion, while others had slightly greater losses with slow infusion. As a proportion of the 1-d dose, only Zn (additional 7% of the dose lost, P = 0.04) and Se (additional 12% of the dose lost, but not statistically significant) had losses of any potential clinical importance. However, in patients with diabetes who received the fast infusion, this appeared to be more marked for Zn (doubled, from 7 to 15%) and Cr (doubled, from 5 to 11%) although the study size was not adequate to show statistical significance for either one.



60 S. Ferrie

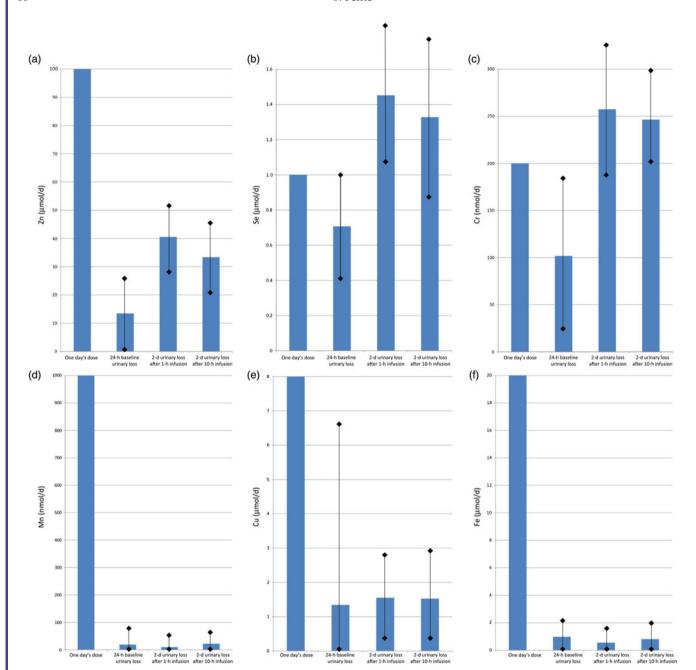


Fig. 4. Urinary trace element losses at baseline and after fast (1 h) v. slow (10 h) parenteral infusion, compared with one day's dose: (a) zinc, (b) selenium, (c) chromium, (d) manganese, (e) copper, (f) iron. Values are means, with standard deviations represented by vertical bars.

One cause of reduced statistical power was the significant variation in urinary loss between individual patients, which was not apparent during the pilot study on which sample size calculations were based. For example, in the case of Fe (which is not normally lost in the urine in significant amounts), twentynine patients had negligible urinary Fe losses throughout the study, losing <0.1 µmol each day, but nine patients lost >1–2 µmol/d on at least 1 d in the study, resulting in a 100-fold difference between individuals. Losses of the other five trace elements varied too, by more than ten-fold between different patients.

Other limitations of the study include the use of only one single washout day after each infusion. Studies using large trace element doses have found that it took longer than this for complete urinary clearance of the dose (5), so it might be preferable to have  $\geq 2$  washout days for accurate quantification of urine losses. This was not feasible in the setting of the present study as it would mean a study period of  $\geq 7$  d, leading to new confounders arising such as the introduction of significant oral intake, removal of urinary catheter (and increased risk of incomplete collection), and discharge from hospital. To identify whether the results were affected by this short washout period, the analysis looked at the





order in which each patient received the two infusions. With an insufficient washout period, it would be expected that the urinary losses after the second infusion would have been boosted by the ongoing residual losses from the first infusion, regardless of whether it was the fast or slow infusion. Reassuringly, the order in which a patient received the two infusions had no statistically significant influence on the results (P > 0.05), suggesting that one washout day was sufficient to establish separation of the total losses after each infusion.

Another limitation is the failure to measure baseline serum trace element levels prior to enrolment, in order to enrol only patients with normal baseline levels. Although malnourished patients were excluded, it was notable that many of the patients had below-normal levels on enrolment, and mean plasma concentrations of all the trace elements increased over the 5 d of the study. Ideally patients with abnormal baseline serum levels would have been excluded, because depleted trace elements status might be expected to be a significant confounder for a study of urinary losses. Independent of the infusion speed, urinary losses might decrease to compensate for deficiency, and might then increase as trace elements status improved, later in the study period. However, as the order in which a patient received the two infusions had no statistically significant influence on the results (P > 0.05), it appears that losses did not decrease significantly when blood levels were lower and thus this potential confounder did not significantly alter the results of the study.

Urinary losses may be unrelated to plasma concentrations in the critically ill for several reasons. Firstly, plasma concentrations of trace elements do not necessarily reflect whole-body status of those elements, particularly in inflammatory conditions where demand for particular elements may be altered, resulting in changes in the location and concentration of those elements within the body<sup>(24)</sup>. Transporters of trace elements include proteins such as albumin, which are present in lower concentrations during the acute phase response, leading to reduced trace elements content in the serum independent of whole-body trace elements status. Also, critical illness may increase the demand for those trace elements that participate in inflammatory processes; oxidative stress may also increase the demand. Reduced serum levels could also reflect the fact that surgery, trauma and burns, and prolonged renal replacement therapy cause increased losses of trace elements by routes other than urine. Gagnon et al. (25) found that plasma levels tended to be low in the first 7 d in ICU, but then mostly resolve. They suggested that short-lived deficits like this may not be meaningful, but equally it could be argued that it is not just a trivial fluctuation but actually indicates the need for increased provision of trace elements early in the ICU stay. Maintenance of normal plasma levels might be assumed to be important for a normal functioning and maintenance of inflammatory response and immune system, as trace elements form a part of key antioxidant enzymes. Berger et al. (26) found that keeping Se concentration in the normal range resulted in a normal plasma glutathione peroxidase activity and reduced infectious complications. Likewise, there are several studies supporting early replacement or supplementation of trace elements in conditions where these losses and demands are high, showing that such replacement improves outcomes<sup>(27-29)</sup>.

There are limited data to indicate precisely the micronutrient requirements of a critically ill patient. The reference values for micronutrient intake in oral and enteral nutrition are based on the maintenance of normal functioning in healthy people. Solid evidence from oral diets allows these reference values to be specified for different groups within the healthy population, such as infants, the elderly, or pregnant women, but requirements during illness are much less clear. Additionally, for PN, the recommended requirements for adults(21,22) are based on much more limited data, insufficient to make separate recommendations for different groups. Knowing the wide individual variations in how trace elements are handled by the body, and the varying effects of illness, it can be assumed that such a 'one size fits all' approach cannot meet the needs of every individual in the ICU. Requirements during critical illness are not adequately described by the limited data on which current parenteral daily dosing is based. Although it appears that parenteral trace element infusions can be given rapidly (e.g. over 1 h) without causing excessive urinary loss, in the ICU population it is essential to follow usual processes for monitoring the dosing and ensuring that individual patients' requirements are being met, to maintain optimal trace elements status.

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S. F. conceived the study, collected, analysed and interpreted all data, and prepared and approved the manuscript.

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# References

- 1. British Pharmaceutical Nutrition Group (2008) Position statement on the use of multichambered parenteral nutrition. http://www.bpng.co.uk/resources/bpng-publications (accessed January 2010).
- 2. Gibbons E, Allwood MC, Neal T, et al. (2001) Degradation of dehydroascorbic acid in parenteral nutrition mixtures. J Pharm Biomed Anal 25, 605-611.
- 3. Berger MM & Shenkin A (2006) Vitamins and trace elements: practical aspects of supplementation. Nutrition 22, 952-955.
- 4. Berger MM, Cavadini C, Chiolero R, et al. (1996) Copper, selenium, and zinc status and balances after major trauma. J Trauma 40, 103–109.
- 5. Tauxe WN, Goldstein NP, Randall RV, et al. (1966) Radiocopper studies in patients with Wilson's disease and their relatives. Am J Med **41**, 375–380.
- Davidsson L, Cederblad A, Lönnerdal B, et al. (1989) Manganese retention in man: a method for estimating manganese absorption in man. Am J Clin Nutr 49, 170-179.
- 7. Malecki EA, Huttner DL & Greger JL (1994) Manganese status, gut endogenous losses of manganese, and antioxidant enzyme activity in rats fed varying levels of manganese and fat. Biol *Trace Elem Res* **42**, 17–29.
- Vincent JL & Forceville X (2008) Critically elucidating the role of selenium. Curr Opin Anesthesiol 21, 148-154.





Manzanares W & Hardy G (2009) Selenium supplementation in the critically ill: posology and pharmacokinetics. Curr Opin Clin Nutr Metab Care 12, 273-280.

- Wang Z, Forceville X, Van Antwerpen P, et al. (2009) A largebolus injection, but not continuous infusion of sodium selenite improves outcome in peritonitis. Shock 32, 140-146.
- 11. Angstwurm MWA, Engelmann L, Zimmermann T, et al. (2007) Selenium in Intensive Care (SIC) study: results of a prospective, randomized, placebo-controlled, multiple-centre study in patients with severe systemic inflammatory response syndrome, sepsis and septic shock. Crit Care Med **35**, 118–126.
- King JC, Shames DM & Woodhouse LR (2000) Zinc homeostasis in humans. J Nutr 130, 1360S-1366S.
- Vale SH, Leite LD, Alves CX, et al. (2014) Zinc pharmacokinetic parameters in the determination of body zinc status in children. Eur J Clin Nutr 68, 203-208.
- 14. Prasad AS (2013) Discovery of human zinc deficiency: its impact on human health and disease. Adv Nutr 4, 176–190.
- Maldonado MA, Gil EB, Fernández SM, et al. (1991) Zinc levels after intravenous administration of zinc sulphate in insulindependent diabetes mellitus patients. J Klin Wochenschr 69, 640-644.
- 16. Danielson BG (2004) Structure, chemistry, and pharmacokinetics of intravenous iron agents. J Am Soc Nephrol 15, Suppl. 2, S93-S98.
- Geisser P & Burckhardt S (2011) The pharmacokinetics and pharmacodynamics of iron preparations. *Pharmaceutics* 3,
- O'Flaherty EJ (1993) A pharmacokinetic model for chromium. Toxicol Lett 68, 145-158.
- Clodfelder BJ, Upchurch RG & Vincent JB (2004) A comparison of the insulin-sensitive transport of chromium in healthy and model diabetic rats. J Inorg Biochem 98, 522-533.

- 20. Cikrt M & Bencko V (1979) Biliary excretion and distribution of 51Cr(III) and 51Cr(VI) in rats. J Hyg Epidemiol Microbial Immunol 23, 241-246.
- 21. Osland EJ, Ali A, Nguyen T, et al. (2016) Australasian Society for Parenteral and Enteral Nutrition (AuSPEN) adult vitamin guidelines for parenteral nutrition. Asia Pac J Clin Nutr 25, 636-650
- 22. Osland EJ, Ali A, Isenring E, et al. (2014) Australasian society for parenteral and enteral nutrition guidelines for supplementation of trace elements during parenteral nutrition. Asia Pac I Clin Nutr 23, 545-553.
- 23. Ferrie S (2010) Does infusion time affect the retention of parenteral micronutrients? Poster. AuSPEN 2010 Annual Scientific Meeting. Gold Coast Queensland. Date of meeting 20-23 October.
- 24. Forceville X, Vitoux D, Gauzit R, et al. (2007) Selenium, systemic immune response syndrome, sepsis, and outcome in critically ill patients. J Trace Elem Med Biol 21, Suppl. 1, 44-48.
- 25. Gagnon G, Voirol P, Soguel L, et al. (2015) Trace element monitoring in the ICU: quality and economic impact of a change in sampling practice. Clin Nutr 34, 422-427.
- 26. Berger MM, Baines M, Raffoul W, et al. (2007) Trace element supplements after major burns modulate antioxidant status and clinical course by way of increased tissue trace element concentration. Am J Clin Nutr 85, 1293-1300.
- 27. Heyland DK, Dhaliwal R, Suchner U, et al. (2005) Antioxidant nutrients: a systematic review of trace elements and vitamins in the critically ill. *Intensive Care Med* **31**, 321–337.
- Visser J, Labadarios D & Blaauw R (2011) Micronutrient supplementation for critically ill adults: a systematic review and meta-analysis. Nutrition 27, 745-758.
- Manzanares W, Dhaliwal R, Jiang X, et al. (2012) Antioxidant micronutrients in the critically ill: a systematic review and meta-analysis. Crit Care 16, R66.

