

## Brain-derived neurotrophic factor (BDNF) gene and rapid-cycling bipolar disorder

Family-based association study

DANIEL J. MÜLLER, VINCENZO DE LUCA, TRICIA SICARD, NICOLE KING, JOHN STRAUSS and JAMES L. KENNEDY

**Background** We have previously reported the Val66Met and GT(n) repeat polymorphisms of the brain-derived neurotrophic factor (BDNF) gene to be associated with bipolar disorder. However, these findings have not been replicated consistently.

**Aims** To dissect the association of the *BDNF* gene with bipolar disorder by examining additional markers at the DNA level and by testing the illness categories of bipolar disorder I and II and rapid cycling.

**Method** We performed a family-based association study and haplotype analyses with 312 nuclear families using four single nucleotide polymorphisms (SNPs) and the Val66Met and GT(n) repeat polymorphisms.

**Results** The SNPs hCVI1592756 and rs2049045, the Val66Met and GT(n) were significantly associated with bipolar disorder using transmission disequilibrium analyses ( $P=0.02, 0.009, 0.001$  and  $0.008$  respectively). The effect at these markers was mainly driven by the rapid-cycling patients.

**Conclusions** Within bipolar disorder, variation in the *BDNF* gene appears to predict risk for developing rapid cycling according to DSM–IV. Incorporating this clinical sub-phenotyping into other studies of the *BDNF* gene may help to resolve some of the inconsistencies reported thus far concerning BDNF and bipolar disorder.

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Polymorphic markers in the brain-derived neurotrophic factor (BDNF) gene have been found to be significantly associated with bipolar disorder in two relatively large samples (Neves-Pereira *et al*, 2002; Sklar *et al*, 2002). However, these findings have not consistently been replicated in other studies (Hong *et al*, 2003; Nakata *et al*, 2003; Kunugi *et al*, 2004; Oswald *et al*, 2004; Skibinska *et al*, 2004; Neves-Pereira *et al*, 2005). Of further interest, studies of the *BDNF* gene in three independent samples with child-onset mood disorder have found a significant association in each sample (Geller *et al*, 2004; Strauss, 2004; Strauss *et al*, 2005). The mixed findings across genetic studies of BDNF in mood disorders may be owing to the variable ascertainment of subtypes, small sizes of effect, genetic heterogeneity or other methodological variation among studies. As a first step in the process of disentangling genotype–phenotype correlations, we examined the role of *BDNF* gene polymorphisms in specific clinical subgroupings of bipolar disorder.

### METHOD

#### Sample

The families were recruited in Toronto and across central Canada through newspaper advertisements and hospital clinic referrals. The procedures were approved by the research ethics board of the Centre for Addiction and Mental Health. After complete description of the study, written informed consent was obtained from each proband and family member.

The sample investigated was mostly recruited from out-patient populations, and consisted of 312 nuclear families with at least one offspring (118 males and 194 females) who had experienced at least one hypomanic or manic episode throughout life diagnosed as bipolar disorder I or II, or schizoaffective disorder, bipolar type, according to DSM–IV criteria (American

Psychiatric Association, 1994). Whenever possible, siblings were included in the study. Thus, 26 siblings with bipolar disorder and 46 non-affected siblings were included. In addition, 45 first- or second-degree relatives (e.g. parents or grandparents) were included, and 12 of these had a lifetime history of bipolar disorder. Thus, the total sample comprised 1043 people, 350 with bipolar disorder (131 males and 219 females) and 693 non-affected relatives. The distribution of bipolar disorder I and II according to DSM–IV criteria was assigned as follows: I, 200; I with rapid cycling, 27; I with seasonal patterns, 4; I with mixed episodes, 3; II, 57; II with rapid cycling, 31; II with seasonal patterns, 8. One person had a diagnosis of bipolar disorder not otherwise specified and 19 schizoaffective disorder, bipolar type. DSM–IV defines rapid cycling as the occurrence of four or more (depressive and/or manic) mood episodes within 12 months. Participants' mean age was 35.4 years (s.d.=10.3), with a mean age at onset of 20.1 years (s.d.=7.5). Participants were mainly White and of European origin ( $n=332, 95%$ ), with 12 Asians (3.4%), 3 Native Americans (0.8%), and 3 African-Americans (0.8%).

#### Diagnostic assessment

Details of the diagnostic assessment procedures for this sample have been published previously (Carter *et al*, 2003). Briefly, DSM–IV diagnoses were based on a standardised best-estimate procedure. A semi-structured clinical interview (SCID–I; American Psychiatric Association, 1994) was performed by a trained research assistant who also interviewed relatives and collected information from medical records. Two experienced psychiatrists subsequently reviewed information in order to assign best-estimate consensus diagnoses. A third psychiatrist reviewed a preset percentage of all cases for quality assurance, and reviewed cases with diagnostic disagreement before a consensus diagnosis was assigned. All individuals with a diagnosis of bipolar disorder I and II were thoroughly assessed for the occurrence of rapid cycling. If a clinical subtype could not be assigned with certainty, these individuals were then excluded from our analyses.

**Table I** Examples of individual haplotype transmission analyses of the *BDNF* gene markers and bipolar disorder (I and II; TDTPHASE)

hCV11592756	Val66Met	GT(n) <sup>1</sup>	rs2049045	T	Freq T	U	Freq U	RR	$\chi^2$	P
A	G (Val)			77	0.59	38	0.29	1.54	11.09	0.0008
A	G (Val)	3		51	0.53	26	0.27	0.37	7.04	0.008
A	G (Val)	3	G	49	0.52	25	0.26	0.36	6.37	0.008
A			G	77	0.59	46	0.35	1.41	6.46	0.01
A	G (Val)		G	73	0.59	36	0.29	2.13	10.43	0.001
	G (Val)	3		65	0.56	30	0.26	2.29	11.2	0.0008

T, number of transmitted haplotypes; U, number of untransmitted haplotypes; Freq, frequency; RR, relative risk.

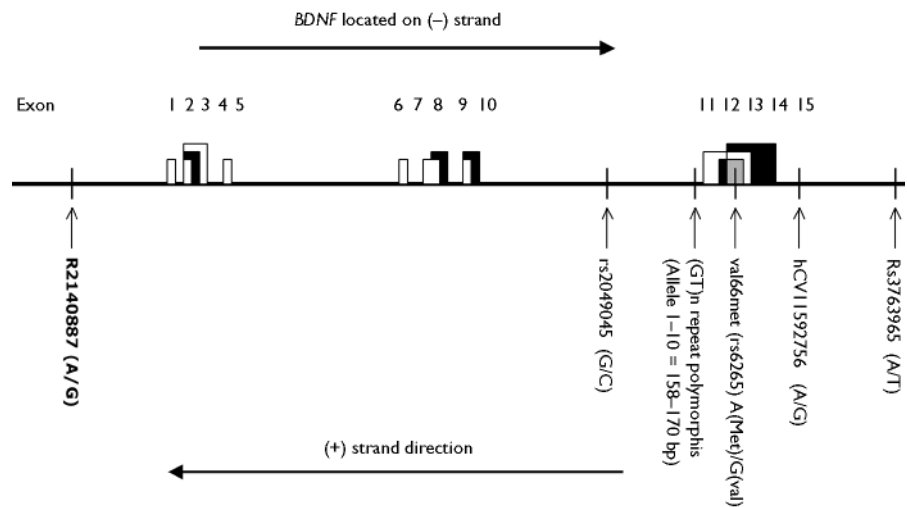
1. Alleles of the GT repeat polymorphism have been dichotomised into two groups (1, presence of allele 1; 2, absence of allele 3);

## Genotyping

The organisation of the *BDNF* gene is rather complex. There are at least two isoforms involving both coding and non-coding exons that are transcribed in both sense and antisense directions (Liu *et al*, 2005). The six markers that we have analysed (Fig. 1) cover a relatively broad region of the gene. Two of the markers, the Val66-Met polymorphism (NCBI SNP cluster ID: rs6265) and the GT dinucleotide repeat polymorphism (Proschel *et al*, 1992) have been previously analysed in our sample for association with bipolar disorder (Neves-Pereira *et al*, 2002). The GT(n) repeat polymorphism consists of up to ten different alleles in various populations. In our analyses, allele 1 is 174 base pairs, allele 2 is 172 base pairs, allele 3 is 170 base pairs, etc.

The four new markers chosen were two each in the upstream (5') and downstream (3') regions of the Val66Met and the GT repeat. Marker rs3763965 is located approximately 148 kb and marker hCV11592756 (Celera ID) about 7 kb upstream of the Val66Met polymorphism. Marker rs2049045 is located 13 kb downstream of the GT repeat and rs2140887 is located approximately 352 kb downstream. The reader is referred to <http://www.hapmap.org> for the latest haplotype interpretation of the *BDNF* gene.

Genotyping of the single nucleotide polymorphisms (SNPs) was performed using 5' nuclease Taqman allelic discrimination assay on the ABI 7000 Sequence Detection System (Applied Biosystems, CA, USA). Commercially available ABI Taqman assays were used, following the manufacturers' recommended protocol. Results were verified independently by two laboratory personnel masked to affection status.



**Fig. 1** Map of *BDNF* gene and markers used in analyses. White, grey or black coloured boxes indicate the alternate splicing. Vertical lines indicate the approximate location of *BDNF* gene polymorphisms. Note that the map represents a simplified model and is not to scale.

## Statistical analyses

Association tests and haplotype analyses between *BDNF* markers and bipolar disorder were performed using TDTPHASE (Dudbridge, 2003). Analyses were first conducted on the total sample with bipolar disorder then on the DSM-IV subtypes of bipolar disorder I and II. Finally, the total sample was divided according to rapid-cycling status: those with rapid cycling (56 nuclear families,  $n=180$ ) and those without (256 nuclear families,  $n=863$ ).

Linkage disequilibrium and identification of haplotype blocks within the *BDNF* gene were performed using HAPLOVIEW (Barrett *et al*, 2005). The standard Lewontin  $D'$  and correlation coefficient  $r^2$  were calculated using the expectation-maximisation algorithm implemented in

HAPLOVIEW (data not shown). The GT(n) repeat polymorphism was divided into allele 3 and other alleles for the linkage disequilibrium analyses.

## RESULTS

### *BDNF* gene polymorphisms and bipolar disorder

The genotype distributions in the overall sample did not deviate from Hardy-Weinberg equilibrium at any of the six *BDNF* polymorphisms.

Significant associations between the total sample and the *BDNF* gene were found for four markers: hCV11592756, Val66Met, GT(n) repeat polymorphism, and rs2049045. Over-transmissions were observed for the A allele of hCV11592756,

the Val (or G) allele of Val66Met, for allele 3 of the GT(n) repeat and for the G allele of rs2049045. Allele 3 of the GT(n) repeat was the most common, and thus was tested against the other alleles for association with bipolar disorder and bipolar disorder I and II, with and without rapid cycling, and for subsequent haplotype analyses. Findings related to the Val66Met and the GT(n) repeat polymorphism were published earlier (Neves-Pereira *et al*, 2002). Two of these *BDNF* markers (hCV11592756 and rs2049045) were not previously known to be associated with bipolar disorder. The remaining two markers (rs3763965 and rs2140887) did not show an association with bipolar disorder (Table 2).

When participants with bipolar disorder were divided into two groups (bipolar disorder I and II), a significant association was found for the Val66Met polymorphism of the *BDNF* gene but for none of the remaining five markers. The Val (or G) allele was over-transmitted (data not shown) compared with the Met (or A) allele (bipolar disorder I: 80 *v.* 52 transmissions,  $P=0.01$ ; bipolar disorder II: 34 *v.* 18 transmissions,  $P=0.02$  respectively).

When participants with bipolar disorder were split into rapid-cycling and non-rapid-cycling subgroups, only the

former showed significant associations with all four polymorphisms that had been found to be associated with the total sample (marker hCV11592756, Val66Met, GT(n), and rs2049045). Overall, the same pattern of over-transmitted alleles was observed as in the total sample. The ratio of transmitted *v.* untransmitted alleles was more pronounced in the rapid-cycling sampling compared with the total sample (Table 3). On the other hand, the non-rapid-cycling sample showed no significant associations with any of the six polymorphisms on the *BDNF* gene (Table 4).

#### Linkage disequilibrium and haplotype analyses

Analyses with HAPLOVIEW revealed that marker rs2049045 is in linkage disequilibrium with marker hCV11592756 and the Val66Met polymorphism ( $D'$  above 0.80). A reduced linkage disequilibrium was noted for allele 3 of the GT(n) repeat polymorphism. The four markers that were individually associated with bipolar disorder proved to be in linkage disequilibrium and are part of a block within the *BDNF* gene. Next, we performed haplotype analyses of these four markers *v.* the phenotype of bipolar disorder. The A-Val-3-G

haplotype proved to be significantly over-transmitted (49:25) in participants with bipolar disorder ( $P=0.008$ ) (Table 1). The strongest results, however, were obtained when the Val allele was combined with either marker hCV11592756 or allele 3 of the GT(n) repeat polymorphism ( $P=0.0008$ , Table 1). We then performed haplotype analyses in the rapid-cycling sample. Although the combination of the Val allele with either marker hCV11592756 or allele 3 of the GT(n) repeat polymorphism yielded significant results, the four-marker haplotype composed of marker hCV11592756 (A allele), Val66Met (Val allele), GT(n) repeat (allele 3) and rs2049045 (G allele) yielded a non-significant trend (Table 5).

## DISCUSSION

### Association of four linked markers of the *BDNF* gene with bipolar disorder

In addition to our previously reported association between the Val66Met and the GT(n) repeat polymorphisms of the *BDNF* gene and bipolar disorder (Neves-Pereira *et al*, 2002), we now report additional markers (hCV11592756

**Table 2** Association tests between *BDNF* markers and bipolar disorder (I and II; TDTPHASE)

Marker	Nucleotide change/ number of alleles	Number of transmitted alleles	Frequency of transmitted alleles	Number of untransmitted alleles	Frequency of untransmitted alleles	RR	LRS	d.f.	<i>P</i>
rs3763965	A	113	0.53	99	0.47	1	0.92	1	0.33
	T	99	0.47	113	0.53	0.88			
hCV11592756	G	59	0.40	87	0.60	1	5.40	1	0.02
	A	87	0.60	59	0.40	1.48			
Val66Met	A (Met)	70	0.38	114	0.62	1	10.62	1	0.001
	G (Val)	114	0.62	70	0.38	1.63			
GT(n)	Allele 1	44	0.28	59	0.37	1	11.66	8	0.17
	Allele 2	06	0.04	10	0.06	0.91			
	Allele 3	83	0.53	54	0.34	1.48			
	Allele 4	18	0.11	25	0.16	0.94			
	Others <sup>1</sup>	06	0.04	09	0.07	–			
	Allele 3	81	0.61	51	0.39	1	6.87	1	0.008
rs2049045	Others <sup>2</sup>	51	0.39	81	0.61	0.63			
	G	83	0.61	53	0.39	1	6.67	1	0.009
rs2140887	C	53	0.39	83	0.61	0.63			
	A	110	0.50	111	0.50	1	0.004	1	0.94
	G	111	0.50	110	0.50	1			

RR, relative risk; LRS, likelihood ratio statistics; allele 1, 174 bp; allele 2, 172 bp; allele 3, 170 bp etc. (bp, base pairs).

1. Alleles 5, 7, 8, 9, 10.

2. Alleles 1, 2, 4, 5, 7, 8, 9, 10.

**Table 3** Association tests between *BDNF* markers and bipolar disorder (I and II) with rapid cycling (TDTPHASE)

Marker	Nucleotide change/ number of alleles	Number of transmitted alleles	Frequency of transmitted alleles	Number of untransmitted alleles	Frequency of untransmitted alleles	RR	LRS	d.f.	P
rs3763965	A	22	0.47	25	0.53	1	0.19	1	0.66
	T	25	0.53	22	0.47	1.13			
hCVI1592756	G	8	0.29	20	0.71	1	5.31	1	0.02
	A	20	0.71	8	0.29	2.5			
Val66Met	A (Met)	8	0.24	25	0.76	1	9.19	1	0.002
	G (Val)	25	0.76	8	0.24	3.12			
GT(n)	Allele 1	5	0.15	18	0.53	1	12.67	5	0.03
	Allele 2	1	0.03	1	0.03	4.03			
	Allele 3	24	0.70	8	0.23	4.03			
	Allele 4	3	0.09	6	0.18	1.42			
	Others <sup>1</sup>	1	0.03	1	0.03	–			
	Allele 3	24	0.75	8	0.25	1	8.37	1	0.004
	Others <sup>2</sup>	8	0.25	24	0.75	0.33			
rs2049045	G	19	0.76	6	0.24	1	7.10	1	0.008
	C	6	0.24	19	0.76	0.31			
rs2140887	A	23	0.47	26	0.53	1	0.18	1	0.67
	G	26	0.53	23	0.47	1.13			

RR, relative risk; LRS, likelihood ratio statistics.

1. Alleles 5, 7, 8, 9, 10.

2. Alleles 1, 2, 4, 5, 7, 8, 9, 10.

**Table 4** Association tests between *BDNF* markers and bipolar disorder (I and II) without rapid cycling (TDTPHASE)

Marker	Nucleotide change/ number of alleles	Number of transmitted alleles	Frequency of transmitted alleles	Number of untransmitted alleles	Frequency of untransmitted alleles	RR	LRS	d.f.	P
rs3763965	A	86	0.53	75	0.47	1	0.75	1	0.38
	T	75	0.47	86	0.53	0.87			
hCVI1592756	G	49	0.45	59	0.55	1	0.92	1	0.33
	A	59	0.55	49	0.45	1.20			
Val66Met	A (Met)	60	0.43	78	0.57	1	2.35	1	0.12
	G (Val)	78	0.57	60	0.43	1.3			
GT(n)	Allele 1	37	0.32	37	0.32	1	10.98	8	0.20
	Allele 2	5	0.04	9	0.08	0.65			
	Allele 3	55	0.48	44	0.38	1.07			
	Allele 4	14	0.12	18	0.15	0.81			
	Others <sup>1</sup>	5	0.04	8	0.07	–			
	Allele 3	52	0.55	43	0.45	1	0.85	1	0.35
	Others <sup>2</sup>	43	0.45	52	0.55	0.83			
rs2049045	G	54	0.54	46	0.46	1	0.64	1	0.42
	C	46	0.46	54	0.54	0.83			
rs2140887	A	79	0.49	82	0.51	1	0.05	1	0.81
	G	82	0.51	79	0.49	1.03			

RR, relative risk; LRS, likelihood ratio statistics.

1. Alleles 5, 7, 8, 9, 10.

2. Alleles 1, 2, 4, 5, 7, 8, 9, 10.

and rs2049045) to be associated with bipolar disorder. Further analyses indicated that these four gene variants are in linkage disequilibrium with each other,

and are part of a haplotype block within the *BDNF* gene. In subsequent studies it may be valuable to sequence affected and unaffected individuals, to

determine whether there are variants not previously known that are highly represented in affected patients and not in controls.

**Table 5** Examples of individual haplotype transmission analyses of the *BDNF* gene marker with rapid cycling bipolar disorder (I and II; TDTPHASE)

hCV11592756	Val66Met	GT(n) <sup>1</sup>	rs2049045	T	Freq T	U	Freq U	RR	$\chi^2$	P
A	G (Val)			16	0.72	4	0.18	6.55	6.58	0.01
A	G (Val)	3		12	0.66	4	0.22	1.17e+008	3.76	0.05
A	G (Val)	3	G	9	0.6	4	0.26	1.06e+008	1.97	0.16
A			G	15	0.75	5	0.25	1	4.81	0.02
A	G (Val)		G	14	0.7	4	0.2	5.12	4.91	0.02
	G (Val)	3		18	0.72	4	0.16	1.70	8.36	0.003

T, number of transmitted haplotypes; U, number of untransmitted haplotypes; Freq, frequency; RR, relative risk.

1. Alleles of the GT repeat polymorphism have been dichotomised into two groups (1, presence of allele 1; 2, absence of allele 3).

### Potential biological impact of *BDNF* markers associated with bipolar disorder

The functional relevance of the GT(n) repeat polymorphism and the other SNPs (apart from Val66Met) remains unknown.

In the case of the GT(n) marker, we have categorised alleles into two groups: allele 3 *v.* alleles 1, 2 and 4–10. This approach, however, remains arbitrary and is different from one previous study that analysed for association between this *BDNF* GT(n) marker and age at onset and therapeutic response in schizophrenia, grouping alleles into longer (172–176 base pairs) *v.* shorter variants (166–174 base pairs) (Krebs *et al.*, 2000). Thus, allele grouping remains arbitrary until future studies elucidate the functional relevance of the GT(n) polymorphism.

In contrast to the other markers in the *BDNF* gene, relatively extensive *in vitro* and *in vivo* functional analyses have been performed for the Val66Met polymorphism. In cultured hippocampal neurons, the Val allele (vBDNF) has been shown to increase intraneuronal BDNF peptide secretion and distribution compared with the Met allele (mBDNF) (Egan *et al.*, 2003). Neurons expressing vBDNF were shown to express BDNF in the cell body and distal processes (dendrites). In contrast, mBDNF was mainly localised in cell bodies. In humans, the Met allele was associated with poorer episodic memory, and abnormal activation of the hippocampus, as measured by functional magnetic resonance imaging (Egan *et al.*, 2003; Pezawas *et al.*, 2004). Our results, and those of others that show the Val allele of Val66Met to be contributing to risk, suggest that relatively rapidly changing mood episodes might be associated with enhanced memory function (Egan *et al.*, 2003) and increased hippocampal–frontal connectivity (Pezawas

*et al.*, 2004). This speculation will require more investigation using cognitive testing and functional imaging techniques, preferably in individuals with rapid-cycling compared with those with other mood patterns. Alternatively, the enhanced memory may be a separate effect of the *BDNF* Val allele and have no biological connection to mood disorder.

In terms of the other markers across the *BDNF* gene, it may be that the GT(n) repeat polymorphism and *BDNF* variants at marker sites hCV11592756 and rs2049045 are not of functional relevance, but are associated with bipolar disorder because of relatively strong linkage disequilibrium with the functionally relevant Val66Met polymorphism. However, it is also plausible that the GT(n) and/or other markers may alter mRNA stability or processing, thus altering the amount of the BDNF peptide that is produced, increasing or decreasing the effect of the Val66Met change in the protein. The net effect on risk for bipolar disorder may thus be best captured by typing multiple markers across the gene.

### *BDNF*: possible association with mood disorders despite inconsistent findings

Our findings obviously need to be put in the context of previous findings. As mentioned above, a number of studies failed to detect a significant association between the *BDNF* gene and bipolar disorder (Hong *et al.*, 2003; Nakata *et al.* 2003; Kunugi *et al.*, 2004; Oswald *et al.*, 2004; Skibinska *et al.*, 2004; Neves-Pereira *et al.*, 2005). Further supportive evidence for association of the *BDNF* gene and mood disorder derives from a recent study that found an association between the Val66Met polymorphism and a prepubertal and early adolescent phenotype (Geller *et al.*, 2004). Other

interesting findings derive from preliminary analyses of a study that included large samples of German descent, and revealed most significant associations between haplotypes (including the Val/Met polymorphism) in two independent samples of patients with major depression ( $n=465$  and  $312$ ), as well as significant association in one sample with bipolar disorder ( $n=281$ ) and one with schizophrenia ( $n=533$ ) (Cichon *et al.*, 2004).

Inconsistent findings may be the result of general problems of molecular genetic association studies dealing with complex disorders (Schulze *et al.*, 2003). Some studies may be too small or under-powered to detect modest gene effects. There is also variation in terms of study design, such as ascertainment strategies and inclusion criteria. Thus far, negative findings have been observed in population-based case–control studies, whereas initial positive findings were based on family-based association studies (Neves-Pereira *et al.*, 2002; Sklar *et al.*, 2002). Indeed, it has been noted that different ascertainment strategies may introduce notable differences with respect to important clinical and demographic characteristics, particularly in samples ascertained for case–control studies *v.* family-based studies (Schulze *et al.*, 2001). Our sample included relatively young out-patients and patients with less severe forms of bipolar disorder (i.e. type II according to DSM–IV). In contrast, other studies have included in-patients who were likely to be older at time of inclusion and possibly affected with more chronic forms of bipolar disorder. Finally, other studies analysed samples of different ethnic backgrounds (including Chinese or Japanese), and thus the *BDNF* gene may represent a genetic risk factor which is more pronounced in Whites than in other ethnic groups, possibly because of local differences in and around the *BDNF* gene or differences in more general genetic background.

## BDNF and rapid cycling

According to the concept of people with bipolar disorder forming sub-populations that share a distinct genetic liability, we analysed the well recognised subgroups, including those with rapid cycling (Dunner & Fieve, 1974). Rapid cycling represents a widely accepted clinical category, and is reported to occur in 5–15% of persons with bipolar disorder (American Psychiatric Association, 1994). People experiencing rapid cycling may represent a distinct subtype with respect to pharmacological response (i.e. resistance to lithium therapy) or demographic factors (i.e. female preponderance) (Mackin & Young, 2004). We observed that our significant association between the *BDNF* gene and bipolar disorder is mainly driven by the inclusion of this particular group of participants. The impact of this effect is strong, as removing the rapid-cycling participants from our analyses eliminates any positive association. This result is mirrored by another study of White people representing the largest bipolar disorder case–control sample to date, that did not detect significant associations between the Val66Met polymorphism and bipolar disorder but did detect a significant association in their subset of rapid-cycling patients (Green *et al*, 2006).

## Implications and limitations

The concordance of our study and that of Green *et al* (2006) has important implications. First, inconsistent findings in previous studies might at least partly result from not examining patients with rapid cycling. Thus, our findings should stimulate re-analysis of previous samples, selecting for patients that experienced rapid cycling. Second, it is plausible that the *BDNF* gene is associated with rapid mood swings or with more general mood instability. Such symptoms are not exclusively encountered in bipolar disorder, but also present in a variety of other neuropsychiatric disorders. In accordance with that hypothesis, *BDNF* gene polymorphisms have been associated with disorders that are characterised by mood symptoms such as schizophrenia (Muglia *et al*, 2003; Neves-Pereira *et al*, 2005), obsessive–compulsive disorder (Hall *et al*, 2003) or adult attention-deficit hyperactivity disorder (Lanktree *et al*, 2004). Third, it is also plausible that *BDNF* is associated with depressive symptoms without mania. Although our findings in the bipolar disorder II sample did only reveal a significant

association with the Val66Met polymorphism, it remains possible that *BDNF* is particularly associated with bipolar disorder II. Rapid cycling is mostly encountered in patients with bipolar disorder II who have less severe manic symptoms, but who tend to have persistent treatment-resistant depression, with at least one full-blown episode of major depression. Also in line with this hypothesis are the significant findings with the *BDNF* gene and childhood-onset mood disorders in a sample of adults who were followed up over two decades from the childhood mood disorder, and only a minority of them became bipolar (Strauss, 2004). Fourth, *BDNF* may be associated with personality traits that may confer susceptibility to mood or depressive symptoms. One study found anxiety- and depression-related personality traits to be associated with the Val/Val genotype of the *BDNF* gene (Lang *et al*, 2005) and another study found an association between the Met allele and lower scores of neuroticism, a risk factor for depression (Sen *et al*, 2003). Thus, it remains unclear whether the association is a direct effect or a confounding factor through some indirect mechanism such as personality traits.

## Summary and outlook

In summary, we hope that our findings will shed some light onto the mixed results surrounding the relationship between the *BDNF* gene and bipolar disorder. According to our findings, the *BDNF* gene could be interpreted as a genetic risk factor for distinct symptoms found in bipolar disorder and other major neuropsychiatric disorders. We hypothesise that rapid cycling is an important feature in the phenotype associated with the *BDNF* gene. Furthermore, another study finds this same specific association (Green *et al*, 2006). None the less, our results should be regarded as preliminary and the specificity of the *BDNF* gene for rapid cycling in bipolar disorder requires replication in further studies. Finally, it is important to bear in mind that effect size of the *BDNF* gene polymorphism appears to be relatively small, with maximum relative risk factors at about 3 for single marker association tests. This, however, is consistent with other complex disorders, where a variety of genes, all bearing small-to-moderate impact on the genetic susceptibility, are observed. In mood disorders, it is probable that different sets of

genes predispose to overlapping phenotypes, some of which belong to the spectrum of bipolar disorder (Kelsoe, 2003). The relatively consistent association of the *BDNF* gene with mood disorders should, with more research, lead to improved understanding of their aetiology and may pave the way for novel and more efficient diagnostic and treatment strategies.

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DANIEL J. MÜLLER, MD, Department of Psychiatry, University of Toronto, Canada, and Charité University of Berlin, Germany; VINCENZO DE LUCA, MD, TRICIA SICARD, BSc, NICOLE KING, BSc, JOHN STRAUSS, MD, JAMES L. KENNEDY, MD, Department of Psychiatry, University of Toronto, Canada

Correspondence: Dr James L. Kennedy, Neurogenetics Section, Centre for Addiction and Mental Health Department of Psychiatry, University of Toronto, 250 College Street R30, Toronto, ON, M5T 1R8, Canada. Tel: +1 416 979 4987; fax +1 416 979 4666; email: James\_Kennedy@camh.net

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