Invited Commentary

Green tea catechins suppress NF-κB-mediated inflammatory responses: relevance to nutritional management of inflammation

(First published online 27 January 2011)

Blood loss after trauma induces several systemic inflammatory responses culminating in the dysfunction and failure of organs. In this issue of the British Journal of Nutrition, Relia et al. (1) have examined the inflammatory signals at the subcellular, cellular and tissue levels after haemorrhage-induced hepatic injury and resuscitation in rats. Hepatic injury and resuscitation induced the expression of intercellular adhesion molecule-1, neutrophil infiltration and necrosis in the liver, and augmented serum alanine transaminase and IL-6 levels. It also induced IκBα phosphorylation and the activation of NF-κB. Pre-treatment with green tea extract (GTE: catechins >80%, with >40% of epigallocatechin gallate (EGCG)) suppressed the inflammatory responses at all levels, including neutrophil infiltration, intercellular adhesion molecule-1 expression and the release of IL-6, and, importantly, suppressed the activation of NF-κB.

The inflammatory responses occurring in the liver after haemorrhage are parallel to the inflammatory events occurring after inducing ischaemia, and EGCG is also active in the latter setting⁽²⁻⁵⁾. The anti-inflammatory efficacy of EGCG demonstrated in all these studies generates a unifying hypothesis. Hepatic injury induced by ischaemia⁽²⁾ caused oxidative stress with enhanced production of reactive oxygen species and TNF-α; both mediated the expression of nuclear factors and kinases, activating the signal transduction pathways to trigger cell death. The liver that stained positive for NF-κB in the ischaemia group remained negative in the EGCG-pretreated group. Neutrophil infiltration that was enhanced in the ischaemia group was significantly reduced after EGCG. Ischaemia-induced myocardial injury(3) also caused significant neutrophil infiltration, an increase in plasma IL-6, and activation of IκB kinase and NF-κB in the tissues. EGCG pre-treatment significantly reduced myocardial damage, neutrophil infiltration and plasma IL-6, and also suppressed the NF-κB pathway. Intestinal injury induced by ischaemia (4) also resulted in an enhanced production of reactive oxygen species, neutrophil infiltration and activation of NF-kB. EGCG pre-treatment significantly deactivated NF-κB, decreased neutrophil infiltration and lowered reactive oxygen species production. All these studies support the conclusion derived by Relja et al. and collectively point out that induced inflammatory responses are mediated through NF-κB-dependent mechanisms, and EGCG per se or in combination with other catechins suppresses NF-κB activation and alleviates inflammation.

There are enumerable reports on the efficacy of EGCG perse or EGCG in combination with other catechins (epigallocatechin or epicatechin gallate or gallocatechin gallate) (5-9) on inflammatory responses induced by different exogenous and endogenous factors. The inflammatory inducers include polymicrobial sepsis⁽¹⁰⁾, lipopolysaccharide^(5-7,11), Staphylococcus aureus enterotoxin B⁽¹²⁾, Helicobacter pylori infection⁽¹³⁾, IL-1β alone $^{(8,14-16)}$ or in combination with β -amyloid $^{(17)}$ or oxygen tension⁽¹⁸⁾ or TNF- α ^(9,19) or TNF- α alone⁽²⁰⁻²²⁾, UV-B⁽²³⁻²⁵⁾ repetitive oxidative stress⁽²⁶⁾, cigarette smoke condensate⁽²⁷⁾, phorbol 12-myristate 13-acetate^(28–30), trinitrobenenesulphonic acid⁽³¹⁾- or acetic acid⁽³²⁾-induced colitis, receptor activator for the NF-κB ligand^(33,34) or high glucose⁽³⁵⁾. Most importantly, all these studies document that consequent to the downregulation of NF-κB pathways, EGCG or catechin combination suppressed the levels of several pro-inflammatory cytokines $(TNF-\alpha^{(11,12,28,32,35)} IL-6^{(17,24,28)}, IL-8^{(16,17,20,27,28)}, interferon \gamma^{(12,32)}$), chemokine (Fractalkine⁽²²⁾) and enzymes (matrix metaloproteinases-1, -3, -9⁽²⁷⁾, -13^(8,15,18), NO synthase^(6,10,14,25,27,32); cyclo-oxygenase-2^(17,19,30), glucosyl/lactosyl and Gb3 transferases⁽²¹⁾), growth factors (vascular endothelial growth factor⁽¹⁷⁾), cell adhesion molecules (intercellular adhesion molecule-1, vascular cell adhesion molecule-1 and E-selectin⁽⁹⁾) and monocyte chemotactic protein-1⁽²⁹⁾. In this regard, the study of Relja et al. is well justified in the use of GTE, since other tea catechins act synergistically with EGCG. It is important to use GTE with a greater percentage of EGCG to counteract inflammation. These preclinical studies on the induced inflammatory responses promote the hypothesis that green tea catechins have the potential to suppress the NF-kBmediated inflammatory pathway into a salient concept relevant to nutritional management of inflammation. The emerging concept is that EGCG or GTE has the potential to block the NF-κB pathway, which plays a critical role in inflammation induced by various factors and also in malignancy. These aforementioned studies pave the way for phase I and II clinical trials using GTE or EGCG to control trauma, haemorrhage or ischaemia-induced inflammation.

There is no conflict of interest.

Naren H. Ravindranath

Department of Biomedical Sciences Norris Dental Science Center Herman Ostrow School of Dentistry
University of Southern California
925 W 34th Street
Den 4110B
Los Angeles
CA 90089
USA
email nravindr@yahoo.com

Mepur H. Ravindranath

Terasaki Foundation Laboratory
11570 West Olympic Boulevard
Los Angeles
CA 90064
USA
emails mepurravi@yahoo.com;
glycoimmune@terasakilab.org

References

- Relja B, Töttel E, Breig L, et al. (2011) Effects of green tea catechins on the pro-inflammatory response after haemorrhage/resuscitation in rats. Br J Nutr 105, 1791–1797.
- Giakoustidis DE, Giakoustidis AE, Iliadis S, et al. (2010) Attenuation of liver ischemia/reperfusion induced apoptosis by epigallocatechin-3-gallate via down-regulation of NFkappaB and c-Jun expression. J Surg Res 159, 720–728.
- Aneja R, Hake PW, Burroughs TJ, et al. (2004) Epigallocatechin, a green tea polyphenol, attenuates myocardial ischemia reperfusion injury in rats. Mol Med 10, 55–62.
- Giakoustidis AE, Giakoustidis DE, Koliakou K, et al. (2008) Inhibition of intestinal ischemia/reperfusion induced apoptosis and necrosis via down-regulation of the NF-kB, c-Jun and caspace-3 expression by epigallocatechin-3-gallate administration. Free Radic Res 42, 180–148.
- Ichikawa D, Matsui A, Imai M, et al. (2004) Effect of various catechins on the IL-12p40 production by murine peritoneal macrophages and a macrophage cell line, J774.1. Biol Pharm Bull 27, 1353–1358.
- Lin YL & Lin JK (1997) Epigallocatechin-3-gallate blocks the induction of nitric oxide synthase by down-regulating lipopolysaccharide-induced activity of transcription factor nuclear factor-kappaB. *Mol Pharmacol* 52, 465–472.
- 7. Yang F, Oz HS, Barve S, *et al.* (2001) The green tea polyphenol (—)-epigallocatechin-3-gallate blocks nuclear factor-kappa B activation by inhibiting I kappa B kinase activity in the intestinal epithelial cell line IEC-6. *Mol Pharmacol* **60**, 528–533.
- 8. Corps AN, Curry VA, Buttle DJ, *et al.* (2004) Inhibition of interleukin-1beta-stimulated collagenase and stromelysin expression in human tendon fibroblasts by epigallocatechin gallate ester. *Matrix Biol* **23**, 163–169.
- Ludwig A, Lorenz M, Grimbo N, et al. (2004) The tea flavonoid epigallocatechin-3-gallate reduces cytokine-induced VCAM-1 expression and monocyte adhesion to endothelial cells. Biochem Biophys Res Commun 316, 659–665.
- Wheeler DS, Lahni PM, Hake PW, et al. (2007) The green tea polyphenol epigallocatechin-3-gallate improves systemic hemodynamics and survival in rodent models of polymicrobial sepsis. Shock 28, 353–359.
- Yang F, de Villiers WJ, McClain CJ, et al. (1998) Green tea polyphenols block endotoxin-induced tumor necrosis factor-production and lethality in a murine model. J Nutr 128, 2334–2340.

- 12. Watson JL, Vicario M, Wang A, *et al.* (2005) Immune cell activation and subsequent epithelial dysfunction by *Staphylococcus* enterotoxin B is attenuated by the green tea polyphenol (–)-epigallocatechin gallate. *Cell Immunol* **237**, 7–16.
- Lee KM, Yeo M, Choue JS, et al. (2004) Protective mechanism of epigallocatechin-3-gallate against Helicobacter pyloriinduced gastric epithelial cytotoxicity via the blockage of TLR-4 signaling. Helicobacter 9, 632–642.
- Singh R, Ahmed S, Islam N, et al. (2002) Epigallocatechin-3-gallate inhibits interleukin-1beta-induced expression of nitric oxide synthase and production of nitric oxide in human chondrocytes: suppression of nuclear factor kappaB activation by degradation of the inhibitor of nuclear factor kappaB. Artbritis Rheum 46, 2079–2086.
- Ahmed S, Wang N, Lalonde M, et al. (2004) Green tea polyphenol epigallocatechin-3-gallate (EGCG) differentially inhibits interleukin-1 beta-induced expression of matrix metalloproteinase-1 and -13 in human chondrocytes. J Pharmacol Exp Ther 308, 767–773.
- Wheeler DS, Catravas JD, Odoms K, et al. (2004) Epigallocatechin-3-gallate, a green tea-derived polyphenol, inhibits IL-1 beta-dependent proinflammatory signal transduction in cultured respiratory epithelial cells. J Nutr 134, 1039–1044.
- 17. Kim SJ, Jeong HJ, Lee KM, *et al.* (2007) Epigallocatechin-3-gallate suppresses NF-kappaB activation and phosphorylation of p38 MAPK and JNK in human astrocytoma U373MG cells. *J Nutr Biochem* **18**, 587–596.
- Andriamanalijaona R, Kypriotou M, Baugé C, et al. (2005) Comparative effects of 2 antioxidants, selenomethionine and epigallocatechin-gallate, on catabolic and anabolic gene expression of articular chondrocytes. J Rheumatol 32, 1958–1967.
- Heinecke LF, Grzanna MW, Au AY, et al. (2010) Inhibition of cyclooxygenase-2 expression and prostaglandin E₂ production in chondrocytes by avocado soybean unsaponifiables and epigallocatechin gallate. Osteoarthr Cartil 18, 220–227.
- Chen PC, Wheeler DS, Malhotra V, et al. (2002) A green tea-derived polyphenol, epigallocatechin-3-gallate, inhibits IkappaB kinase activation and IL-8 gene expression in respiratory epithelium. *Inflammation* 26, 233–241.
- Moon DO, Choi SR, Lee CM, et al. (2005) Epigallocatechin-3-gallate suppresses galactose-alpha1,4-galactose-1beta, 4-glucose ceramide expression in TNF-alpha stimulated human intestinal epithelial cells through inhibition of MAPKs and NF-kappaB. J Korean Med Sci 20, 548–554.
- Lee AS, Jung YJ, Kim DH, et al. (2009) Epigallocatechin-3-O-gallate decreases tumor necrosis factor-alpha-induced fractalkine expression in endothelial cells by suppressing NF-kappaB. Cell Physiol Biochem 24, 503–510.
- 23. Afaq F, Adhami VM, Ahmad N, *et al.* (2003) Inhibition of ultraviolet B-mediated activation of nuclear factor kappaB in normal human epidermal keratinocytes by green tea constituent (–)-epigallocatechin-3-gallate. *Oncogene* 22, 1035–1044.
- 24. Xia J, Song X, Bi Z, *et al.* (2005) UV-induced NF-kappaB activation and expression of IL-6 is attenuated by (–)-epigal-locatechin-3-gallate in cultured human keratinocytes *in vitro*. *Int J Mol Med* **16**, 943–950.
- Song XZ, Bi ZG & Xu AE (2006) Green tea polyphenol epigallocatechin-3-gallate inhibits the expression of nitric oxide synthase and generation of nitric oxide induced

https://doi.org/10.1017/S0007114510005611 Published online by Cambridge University Press

- by ultraviolet B in HaCaT cells. Chin Med J (Engl) 119, 282 - 287
- 26. Sen P, Chakraborty PK & Raha S (2006) Tea polyphenol epigallocatechin 3-gallate impedes the anti-apoptotic effects of low-grade repetitive stress through inhibition of Akt and NFkappaB survival pathways. FEBS Lett 580, 278-284.
- 27. Syed DN, Afaq F, Kweon MH, et al. (2007) Green tea polyphenol EGCG suppresses cigarette smoke condensateinduced NF-kappaB activation in normal human bronchial epithelial cells. Oncogene 26, 673-682.
- Shin HY, Kim SH, Jeong HJ, et al. (2007) Epigallocatechin-3gallate inhibits secretion of TNF-alpha, IL-6 and IL-8 through the attenuation of ERK and NF-kappaB in HMC-1 cells. Int Arch Allergy Immunol 142, 335-344.
- Hong MH, Kim MH, Chang HJ, et al. (2007) Epigallocatechin-3-gallate inhibits monocyte chemotactic protein-1 expression in endothelial cells via blocking NF-kappaB signaling. Life Sci 80, 1957-1965.
- Kundu JK & Surh YJ (2007) Epigallocatechin gallate inhibits phorbol ester-induced activation of NF-kappa B and CREB

- in mouse skin: role of p38 MAPK. Ann N Y Acad Sci 1095,
- 31. Abboud PA, Hake PW, Burroughs TJ, et al. (2008) Therapeutic effect of epigallocatechin-3-gallate in a mouse model of colitis. Eur J Pharmacol 579, 411-417.
- 32. Ran ZH, Chen C & Xiao SD (2008) Epigallocatechin-3-gallate ameliorates rats colitis induced by acetic acid. Biomed Pharmacother 62, 189-196.
- 33. Lin RW, Chen CH, Wang YH, et al. (2009) Epigallocatechin gallate inhibition of osteoclastic differentiation via NFkappaB. Biochem Biophys Res Commun 379, 1033-1037.
- 34. Lee JH, Jin H, Shim HE, et al. (2010) Epigallocatechin-3gallate inhibits osteoclastogenesis by down-regulating c-Fos expression and suppressing the nuclear factor-kappaB signal. Mol Pharmacol 77, 17-25.
- 35. Wu CH, Wu CF, Huang HW, et al. (2009) Naturally occurring flavonoids attenuate high glucose-induced expression of proinflammatory cytokines in human monocytic THP-1 cells. Mol Nutr Food Res 53, 984-995.