

Dynamic Nanoscale Imaging of Beam Sensitive Materials with *In Situ* Fluid Stage

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In effort to solve biological processes at the molecular level, the dynamic transmission electron microscope (DTEM) has been used in combination with an *in situ* fluid stage holder. The DTEM has proven its utility by imaging material processes such as phase transformations with spatial resolutions of about 10 nm and time resolutions of 15ns.[1] In this experiment the dynamic transmission electron microscopy techniques are applied to solution systems using a fluid stage holder. The holder consists of 50 nm thick electron transparent Si₃N₄ windows separated by 0.5 μm Au platforms, 4 nm spatial resolution has been demonstrated with a fluid stage in scanning transmission electron microscopy (STEM).[2] The primary advantage of the DTEM over STEM imaging is the capability of acquiring signals on ultrafast time scales. Under this condition the effects of Brownian motion are reduced, and do not contribute to image blur. This is especially useful for easily damaged biological specimens, as image is captured by the nanosecond pulse before damage occurs and the fluid flow inside the cell extracts the damaged structures created from preceding electron beam interactions. In particular, membrane protein imaging could benefit from this technique.

Gold nanoparticles were used for comparison of the dynamic imaging to conventional cryo-electron microscopy and negative stain techniques that image static biomaterials. Scattering of the electron beam through the fluid cell in combination with space charge effects from the high current electron beam of the DTEM have degraded the signal to noise ratio. General biological processes require higher spatial resolution in comparison to temporal resolution, providing a tradeoff to increase the resolvable characteristics within the sample. The signal to noise ratios achieved with DTEM are comparable to those in standard cryo-EM imaging, but with the added benefit of very high time resolution and the capability to image structures before damage can occur.

References

- [1] T.B. LaGrange et al., *Ultramicroscopy*. 108 (2008) 1441.
- [2] N. de Jonge et al., *PNAS*. 106, 7 (2009) 2159.
- [3] M.R. Armstrong, *Ultramicroscopy*. 107 (2007) 356.
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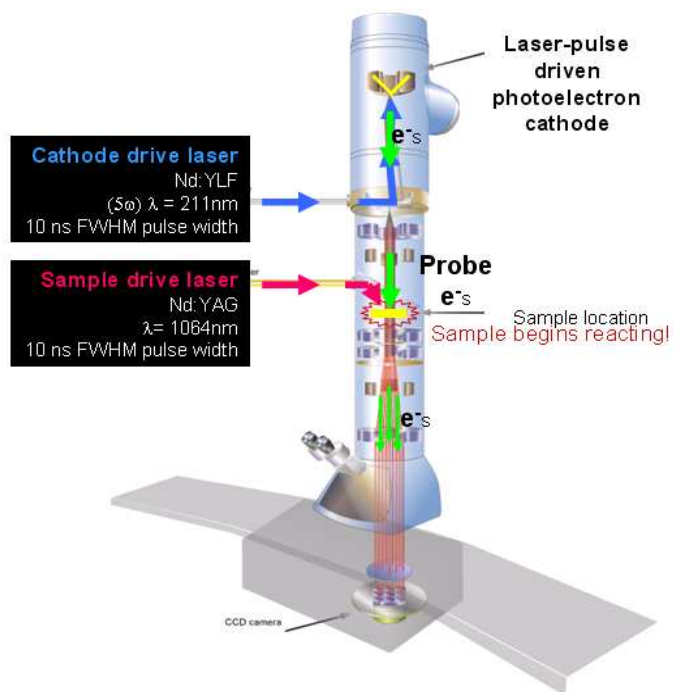


FIG. 1. Dynamic Transmission Electron Microscope: JEOL 2000FX column with two laser ports into the column.[1,3] The cathode drive laser stimulates a photoelectron cathode for emission pulse of 10^9 electrons; sample drive laser interacts directly with the sample for reaction initiation.

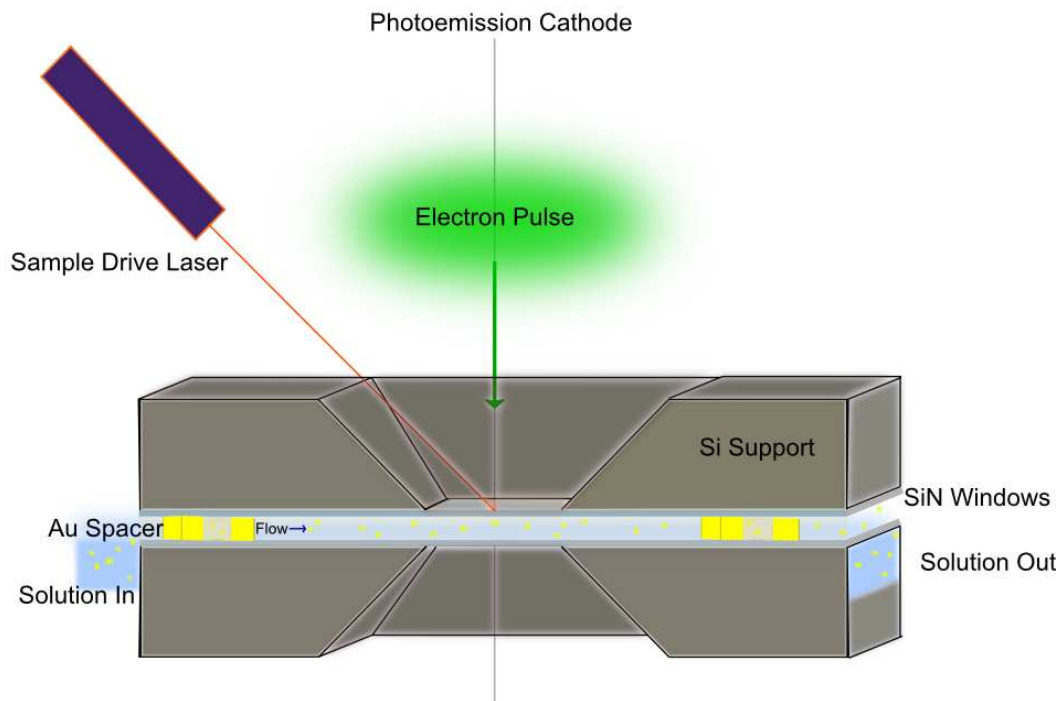


FIG. 2. Dynamic transmission electron microscope imaging, used in combination with the fluid stage holder. A cross-section view of a fluid cell flowing Au nanoparticles between two 50 nm thick SiN membrane windows spaced with 500 nm Au platforms.