

Mycobacterial bone marrow infections at a medical centre in Taiwan, 2001–2009

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SUMMARY

Mycobacterial bone marrow (BM) infection is the most common diagnosis established by BM examinations for fever of unknown origin. In this study, clinical features and outcomes of patients who fulfilled the criteria for BM infection due to Mycobacterium tuberculosis (MTB) and non-tuberculous mycobacteria (NTM) at a medical centre in Taiwan from 2001 to 2009 were investigated. The BM histopathological findings were also analysed. A total of 24 patients (16 men, eight women) with mycobacterial BM infections were found. Of these, nine (38%) were positive for human immunodeficiency virus (HIV) and six (25%) had no pre-existing immunocompromised conditions. MTB isolates were obtained from 11 (46%) patients and NTM species were isolated from 10 (42%) patients, including M. avium complex (MAC, n=7) and M. kansasii (n=3). Patients with MTB infections were significantly older than those with NTM infections (60.5 vs. 47.7 years, P = 0.043) and were less likely to have a positive BM culture (45% vs. 100%, P=0.012). The 90-day survival rates for MTB and NTM BM infections were 68% and 60%, respectively (P = 0.61). In addition, the presence of BM granulomas was significantly more common in patients with MTB BM infections than in those with NTM infections (82% vs. 30%, P=0.030). In Taiwan, the importance of NTM was not inferior to MTB and besides MAC, M. kansasii might be an important pathogen in non-HIV-infected patients. The presence of BM granulomas and caseation provides valuable information regarding early treatment pending culture results.

Key words: Bone marrow infection, granuloma, *Mycobacterium avium* complex, *Mycobacterium kansasii*, *Mycobacterium tuberculosis*.

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INTRODUCTION

Bone marrow (BM) examination is a useful method for establishing a diagnosis in patients with or without human immunodeficiency virus (HIV) infection who present with fever of unknown origin (FUO) and cytopenia [1, 2]. The diagnostic yield of BM examination varies from 18% to 47%, and the most common diagnoses made by BM examination are haematological malignancies and infections, especially those caused by mycobacteria [1-6]. Mycobacterial BM infection is a disseminated infection associated with a high mortality rate and prompt anti-mycobacterial treatment is crucial to achieve a good outcome [7, 8]. However, timely identification of mycobacterial species is not simple because mycobacterial culture methods are time consuming and a substantial proportion of mycobacterial infections are culturenegative [9–11]. Therefore, empirical therapy for tuberculosis (TB) is often instituted pending bacteriological confirmation of mycobacterial species [10, 11].

In countries with a high burden of TB, mycobacterial BM infections in HIV-positive patients are almost always caused by *Mycobacterium tuberculosis* (MTB) [3, 12]. Therefore, the empirical use of anti-TB treatment is a rational decision [10]. However, in countries with a low prevalence of TB, the case numbers of BM infections due to *M. avium* complex (MAC) and those due to MTB in HIV-positive patients are similar [2, 13]. Furthermore, in the USA, a country with a very low prevalence of TB, the majority of mycobacterial BM infections in HIV-infected patients are caused by MAC [5, 6, 14].

In Taiwan, where TB is endemic, the prevalence of TB decreased during 2000–2009, while the prevalence of non-tuberculous mycobacterial (NTM) infections increased [15, 16]; in addition, the prevalence of HIV infection is very low in Taiwan [17]. There is a lack of studies regarding mycobacterial BM infections in Taiwan. Moreover, previous studies have shown limited knowledge about mycobacterial BM infections and discrimination between MTB and NTM BM infections is difficult. Since a better understanding of mycobacterial BM infections could be vital to ensuring that these patients receive timely and accurate anti-mycobacterial treatment, we performed a retrospective cross-sectional study to investigate the profile of HIV-infected and non-HIV-infected patients with mycobacterial BM infections in which histopathological and microbiological BM studies were performed at a medical centre in northern Taiwan,

and to compare the clinical characteristics between patients with MTB and NTM BM infections.

PATIENTS AND METHODS

Patient population and definitions

This study was performed at a 2500-bed, universityaffiliated tertiary hospital in northern Taiwan. From 1 January 2001 to 31 December 2009, data of BM specimens sent for mycobacterial culture and histological examinations in the microbiology and pathology laboratories were retrieved from the computerized databases. During the 9-year study period, the mean annual incidences of TB, NTM disease and HIV infection in the study hospital were 391, 193, 106 cases per year, respectively. All patients with BM aspirates with a positive mycobacterial culture or histopathological specimens showing acid-fast bacilli (AFB) and/or granulomatous inflammation were analysed. BM is regarded as one of the involved sites in disseminated mycobacterial infections, and the gold standard for diagnosing mycobacterial BM infection is a positive BM mycobacterial culture [7, 18]. However, a substantial proportion of mycobacterial infections are culture-negative; therefore, a clinical diagnosis can be established based on histopathological features that are characteristic of mycobacterial infection combined with characteristic clinical presentations, such as miliary pattern, fibrocavitary, nodular or bronchiectatic lesions on the chest radiographs and presence of lymphocytosis in pleural fluid or cerebrospinal fluid (CSF) [9, 11].

In this study, both microbiological and histopathological criteria were adopted. Mycobacterial BM infection was diagnosed if the patient fulfilled any of the following criteria: (1) isolation of mycobacteria from BM culture; (2) histopathological demonstration of granulomatous changes and AFB from BM with negative stains for fungi; and (3) histopathological demonstration of granulomatous changes or AFB from BM plus a culture-proven diagnosis or a clinical diagnosis of mycobacterial infection in body sites other than BM and without concomitant fungal infections. Clinical diagnoses of MTB and NTM infections in other body sites were made as described above.

Mycobacteriology studies

All specimens sent for mycobacterial culture in the study hospital were processed according to previous description and guidelines [18–20]. NTM isolates were identified to the species level using conventional biochemical methods. Drug susceptibility testing to first-line anti-TB drugs including isoniazid, rifampicin and ethambutol was performed using the modified proportional disk elution method.

Histopathological examination of BM biopsy specimens

BM samples were obtained by posterior iliac crest trephine biopsy. Histopathological changes were assessed by pathologists and the routine stains for microorganisms included Gomori methenamine-silver stain, periodic acid-Schiff stain, and Kinyoun acid-fast stain [21].

Clinical characteristics and outcome

Information on age, sex, underlying immunocompromised conditions, haematological tests, extent of mycobacterial infection, types of specimens positive for mycobacteria, histopathological findings from BM, and in-hospital mortality were collected by chart review and further analysed. In addition to a positive mycobacterial culture, mycobacterial infection in other body sites was also recorded based on the presence of relevant laboratory, histopathological or radiological findings. Underlying immunocompromised conditions, including diabetes mellitus, HIV infection, solid-organ cancer, haematological disorders, liver cirrhosis, autoimmune disease, and end-stage renal disease (ESRD) requiring dialysis were recorded. Patients without HIV serological testing were presumed to be HIV negative if they had no past history to suggest HIV infection. Leukopenia was defined as leukocyte count of <4000 cells/mm³, thrombocytopenia was defined as platelet count of <150000 cells/mm³, and anaemia was defined as haemoglobin level of <13 g/dl for men or <12 g/dl for women. Pancytopenia was defined as the presence of leukopenia, thrombocytopenia, and anaemia. The 90-day survival rate was evaluated and that of patients discharged from the hospital within 90 days was investigated using medical records of subsequent outpatient department followup visits.

Statistical analysis

Continuous variables are expressed as mean±standard deviation (s.d.) when there was a parametric

distribution. Otherwise, data are expressed as median [interquartile range (IQR)]. Student's t test or Mann–Whitney U test were used to compare continuous variables. We used Fisher's exact test or χ^2 test to compare proportions; besides odds ratios (ORs) and their 95% confidence intervals (CIs) were calculated. 90-day survival curves of MTB, and NTM BM infections were compared using the Kaplan–Meier method and the log-rank test. P values <0.05 were considered to be statistically significant. All analyses were performed with SPSS, version 11.0 (SPSS Inc., USA).

RESULTS

Clinical characteristics

From January 2001 to December 2009, pathological examinations were performed in 1191 BM biopsy samples and a total of 693 BM specimens were sent for mycobacterial culture. Mycobacterial BM infection was diagnosed in 24 patients (Fig. 1), which was equivalent to 4.0 cases/100000 inpatients and their characteristics are given in Table 1. Of these, 15 (five TB, 10 NTM), two (one TB), and seven (five TB) were diagnosed as mycobacterial BM infections based on the first, second and third criteria mentioned above, respectively. Fever (83%) was the most common reason of BM examination, followed by cytopenia (38%). Sixteen (67%) patients were men and the mean age of all patients was 53.7 ± 16.9 years. BM was the sole infection site in four (17%) patients and the most frequent concomitant infection foci were lung (n=16, 67%), bloodstream (n=9, 38%), abdomen (n=9, 38%), and central nervous system (CNS) (n=4, 17%). Nine (38%) of the patients were HIV positive. Of the remaining 15 patients, 12 (80%) had a negative serological test for HIV infection and the remaining three (20%) had no past history to suggest HIV infection. Other underlying immunocompromised conditions included diabetes mellitus (n=4, 17%), liver cirrhosis (n=3, 13%), malignancy (n=3, 13%), ESRD (n=1, 4%), systemic lupus erythematosus (n=1, 4%) and myelodysplastic syndrome (n=1,4%); six (25%) patients had no immunosuppressed condition.

MTB isolates were obtained from 11 (46%) patients and NTM species were isolated from 10 (42%) patients, including MAC in seven and *M. kansasii* in three patients. Three (27%) patients with MTB infection and four (40%) with NTM infection were HIV

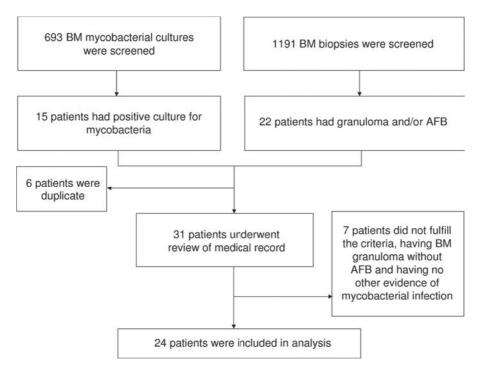


Fig. 1. Fluxogram of identification of patients. BM, bone marrow; AFB, acid-fast bacilli.

positive (OR 0.6, 95% CI 0.09–3.5, P = 0.66). None of the three patients with infection due to M. kansasii were HIV positive. At least one set of mycobacterial blood culture was performed for all patients. Bloodstream infection was found in four (37%) patients with MTB infection and in five (50%) with NTM infection (OR 0.6, 95% CI 0.1–3.3, P = 0.67); lung involvement was found in eight (73%) patients with MTB infection and in six (60%) with NTM infection (OR 1·8, 95% CI 0·3–11·1, P = 0.66); abdomen involvement was found in six (55%) patients with MTB infection and in three (30%) with NTM infection (OR 2·8, 95% CI 0·5–16·9, P = 0.39); and CNS involvement was found in two (18%) patients with MTB infection and in two (20%) with NTM infection (OR 0.9, 95% CI 0.1–7.9, P = 0.65). Patients with MTB BM infection were significantly older than those with NTM infection $(60.5 \pm 11.7 \text{ vs. } 47.7 \pm 15.3 \text{ ms. } 47.7 \pm 15.3 \text{ ms.})$ years, P = 0.043).

Outcome

All MTB isolates were susceptible to rifampicin and isoniazid. All patients with MTB or unspecified mycobacterial infection were initially treated with a four-drug combination anti-TB treatment (isoniazid, rifampicin, pyrazinamide, ethambutol), and patients with NTM infection received standard antibiotic

treatment [11]. MAC infections were treated with a macrolide (clarithromycin or azithromycin) in combination with ethambutol, or rifampicin, or rifabutin. M. kansasii infection was treated with an ethambutol-based combination regimen. The all-cause in-hospital mortality rate was 33%. No significant difference in clinical characteristics was noted between patients who died and those who survived (Table 2). The Kaplan–Meier survival curves for patients with MTB and NTM BM infections are shown in Figure 2, and the 90-day survival rates were 68% and 60%, respectively (log-rank P=0·61).

BM examinations

Granulomas were seen in 15 (63%) patients, and only three (13%) had caseation. BM mycobacterial culture was positive in 15 (63%) patients and AFB was seen in nine (38%). Of the nine patients with a negative BM mycobacterial culture, all had granulomas while AFB was seen in only two (22%) patients. Of the 15 patients with a negative blood culture for mycobacteria, eight (53%) had a positive BM culture for mycobacteria. Of the seven patients with negative BM and blood mycobacterial cultures, three had positive mycobacterial cultures from other body sites, two had a clinical diagnosis of miliary TB, and one had BM granuloma plus AFB. Of the nine patients

Table 1. Characteristics of 24 patients with mycobacterial BM infection, 2001–2009

Patient no.	Age (yr)	Sex	Underlying immunocompromised status	Fever/cytopenia	BM histological findings suggestive of mycobacterial infection	Probable site of concomitant involvement without positive culture for mycobacteria	Specimens with positive cultures for mycobacteria	Mycobacterial species
1	72	F	None	+/_	Granuloma	None	Sputum	M. tuberculosis
2	59	M	None	+/_	Granuloma, caseation	None	Blood, sputum	M. tuberculosis
3	58	F	ESRD, DM	-/-	Granuloma	CNS (CSF lymphocytosis)	Sputum	M. tuberculosis
4	45	M	Cancer of unknown origin	+/thrombocytopenia	None	None	BM	M. kansasii
5	53	M	HIV infection	+/pancytopenia	Granuloma, AFB	Neck LN (granuloma)	BM, blood	M. tuberculosis
6	46	M	None	+/pancytopenia	Granuloma, AFB	None	Sputum, ascites	M. tuberculosis
7	59	M	Lymphoma	+/leukopenia	None	None	BM	M. avium complex
8	86	M	None	+/pancytopenia	Granuloma, AFB	None	None	Negative
9	29	M	HIV infection	+/pancytopenia	Granuloma	Lung (miliary lesions)	None	Negative
10	55	M	HIV infection	–/pancytopenia	None	None	BM, sputum, CSF	M. tuberculosis
11	30	M	HIV infection	+/-	Granuloma	Lung (miliary lesions), neck LN (granuloma)	None	Negative
12	57	M	None	-/thrombocytopenia	Granuloma	None	Blood, ascites, liver	M. tuberculosis
13	44	M	HIV infection, liver cirrhosis	+/-	Granuloma, caseation, AFB	None	BM, blood, ascites	M. tuberculosis
14	49	M	HIV infection	+/_	None	None	BM, blood, sputum	M. avium complex
15	45	M	MDS	+/_	Granuloma	None	BM	M. avium complex
16	45	M	HIV infection	+/_	AFB	None	BM, blood, sputum	M. avium complex
17	60	F	None	+/pancytopenia	Granuloma	Skin (granuloma), mediastinal LN (granuloma)	BM	M. kansasii
18	38	F	Lymphoma	+/_	None	None	BM, sputum, CSF	M. kansasii
19	22	M	HIV infection	+/-	AFB	None	BM, blood, sputum, pleural fluid, ascites, stool	M. avium complex
20	82	F	DM, liver cirrhosis	+/pancytopenia	None	None	BM, sputum	M. tuberculosis
21	78	M	DM	+/_	Granuloma, AFB	None	BM, blood, sputum, CSF	M. avium complex
22	69	F	SLE	+/_	Granuloma	Lung (miliary lesions)	Urine	M. tuberculosis
23	36	F	HIV infection	+/-	AFB	None	BM, blood, sputum, pleural fluid, skin	M. avium complex
24	71	F	DM, liver cirrhosis	+/_	Granuloma, caseation, AFB	None	BM, sputum, urine, pleural fluid, ascites	M. tuberculosis

BM, Bone marrow; ESRD, end-stage renal disease; DM, diabetes mellitus; CNS, central nervous system; CSF, cerebrospinal fluid; HIV, human immunodeficiency virus; AFB, acid-fast bacilli; LN, lymph node; MDS, myelodysplastic syndrome; SLE, systemic lupus erythematosus.

Ta	bl	e 2	2. (Comparison	of c	haracteristics	between	survivors an	d non-survivors

Characteristics	Survivors $(n=16)$	Non-survivors $(n=8)$	OR (95% CI)*	P value	
Demographics					
Age (yr), mean \pm s.D.	50.6 ± 16.4	59.8 ± 17.3		0.22	
Female gender, n (%)	6 (38)	2 (25)	1.8 (0.3–12.0)	0.68	
Co-morbidity, <i>n</i> (%)					
HIV infection	7 (44)	2 (25)	2.3 (0.4–15.3)	0.66	
Diabetes mellitus	3 (19)	1 (13)	1.6 (0.1 - 18.6)	>0.99	
Liver cirrhosis	3 (19)	0 (0)	_	0.52	
Malignancy	1 (6)	2 (25)	0.2 (0.015 - 2.6)	0.25	
Any immunocompromised	13 (81)	5 (63)	2.6 (0.4–17.5)	0.36	
Site of involvement, n (%)					
Bloodstream	6 (38)	3 (38)	1.0 (0.2-5.8)	>0.99	
Lung	10 (63)	6 (75)	0.6 (0.08 - 3.7)	0.67	
Abdomen	7 (44)	2 (25)	2.3 (0.4–15.3)	0.66	
CNS	1 (6)	3 (38)	0.1 (0.009–1.3)	0.09	
Lymph node	2 (13)	1 (13)	1.0 (0.08–13.0)	>0.99	

OR, Odds ratio; CI, confidence interval; s.D., standard deviation; HIV, human immunodeficiency virus; CNS, central nervous system.

^{*}Odds ratio is not calculable due to zero frequencies in some cells.

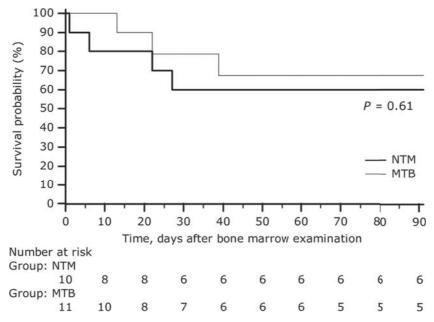


Fig. 2. The 90-day Kaplan–Meier survival curves of bone marrow infections due to non-tuberculous mycobacteria (NTM) and *M. tuberculosis* (MTB) during 2001–2009 (hazard ratio 0.68, 95% confidence interval 0.15–3.0, P=0.61, log-rank test).

with a positive blood culture for mycobacteria, seven (78%) had a positive BM culture for mycobacteria. A positive blood mycobacterial culture was not associated with a significantly higher positive rate of BM mycobacterial culture (OR $3\cdot1$, 95% CI $0\cdot5-19\cdot9$, $P=0\cdot39$). BM was the only body site in which mycobacteria were isolated in four (17%) patients (MAC, n=2; M. kansasii, n=2).

A comparison of haemograms and BM examinations between patients with MTB and those with NTM BM infections is shown in Table 3. There were no significant differences in haemograms between the two groups of patients. However, the proportion of patients with granulomas was higher in the group of patients with MTB infections than in patients with NTM infections (82% vs. 30%,

Table 3. Comparison of patients with BM infections due to MTB and those due to NTM by haemogram and BM findings

Finding		MTB $(n = 11)$		(n=10)	OR (95% CI)*	P value			
Haemogram									
Leukocyte count (10 ³ cells/mm ³), median (IQR)	4.3	(2.9-5.5)	4.7	(1.0-17.6)		0.89			
Platelet count (10 ³ cells/mm ³), median (IQR)	66.0	(44.3 - 83.3)	44.0	(20.0-99.0)		0.40			
Haematocrit (%), mean ± s.D.	28.6	±6.5	25.2	±4·1		0.18			
BM examination, n (%)									
Haemopoietic cellularity ≤30%,	6	(55)	5	(50)	1.2 (0.2–6.7)	>0.99			
Decreased megakaryocyte	4	(36)	4	(40)	0.9 (0.1-5.0)	>0.99			
Granuloma	9	(82)	3	(30)	10.5 (1.4-81.1)	0.030			
Caseation	3	(27)	0	(0)	_	0.21			
Positivity of BM acid-fast stain	4	(36)	4	(40)	0.9 (0.1-5.0)	>0.99			
Positivity of BM mycobacterial culture	5	(46)	10	(100)		0.012			

BM, Bone marrow; MTB, *M. tuberculosis*; NTM, non-tuberculous mycobacteria; OR, odds ratio; CI, confidence interval; IQR, interquartile range; s.d., standard deviation.

P = 0.030). Furthermore, caseation was observed exclusively in patients with MTB infections. All patients with NTM infections had BM cultures positive for mycobacteria and 46% of patients with MTB infections had BM cultures positive for mycobacteria (P = 0.012).

DISCUSSION

In this hospital-based study in Taiwan we found that a substantial proportion of patients with mycobacterial BM infections were HIV negative or without any preexisting immunocompromised condition. We also found that the number of NTM BM infections was similar to that of MTB infections and that in addition to MAC, *M. kansasii* was an important pathogen in non-HIV-infected patients. NTM and MTB BM infections were associated with similar outcomes and most deaths occurred within 30 days after diagnosis. Further, patient's age and histological features of BM biopsy specimens, such as granulomas and caseation, might be helpful in distinguishing between NTM and MTB BM infections before final identification of mycobacterial species.

Hafner *et al.* showed that in patients with disseminated MAC infection, BM bacterial load was 1000-fold higher than bacterial load in blood and was correlated with poor outcome [22]. In our study, culture-positive rates of BM and blood were 63% and 38%, respectively; the combination of both had a positive rate of 71%. Since the combination of BM and blood cultures provide the maximal diagnostic

yield for disseminated mycobacterial infections, the superiority of BM culture to blood culture was not always found in previous studies [13, 21, 23]. In the present study, mycobacterial species were isolated from BM in 8/15 patients with negative blood cultures. The additional diagnostic value of BM culture might be partially explained by the lack of association between BM and blood mycobacterial load [22]. Therefore, the importance of BM cultures should not be overlooked in patients with suspected disseminated mycobacterial infections.

Although previous studies have shown that the diagnostic value of BM culture is superior to that of BM histopathology for the detection of mycobacterial BM infections, BM histopathology nonetheless remains a valuable diagnostic tool [2, 5, 14, 24, 25]. In patients with disseminated mycobacterial infection, early treatment has been shown to be associated with better survival [7, 8]; however, it is difficult to distinguish between NTM and MTB BM infections before culture results are available. Nichols et al. found that caseous necrotic granulomas were observed exclusively in patients with MTB BM infections and that BM granulomas in patients with BM infections due to MTB tended to be larger and more tightly cohesive than those in patients with BM infections caused by MAC [24]. Benito et al. also showed that caseous necrosis was observed only in BM infections due to MTB and that BM granulomas were observed in 83.3% of patients with BM infections caused by MTB and in 64.3% of patients with BM infections due to MAC [2]. In the present study, caseation was

^{*} Odds ratio is not calculable due to zero frequencies in some cells.

noted in 3/11 patients with MTB BM infections and was not observed in any of the patients with BM infections due to NTM. In addition, the prevalence of BM granulomas was significantly higher in patients with MTB BM infections than in patients with BM infections caused by NTM. Further investigations utilizing polymerase chain reaction or interferon- γ assays to aid in the early differentiation of MTB from NTM BM infections are warranted [26, 27].

Studies investigating the utility of BM examinations in HIV-positive patients have demonstrated a positive relationship between the proportion of MTB in mycobacterial BM infections and the regional TB incidence density [2, 3, 5, 6, 12–14]. Unlike NTM infections, a clinical diagnosis of TB is generally accepted if there is good radiological or histopathological evidence of TB in patients who respond well to treatment [10, 11]. In countries with a high prevalence of TB, patients with suspected disseminated mycobacterial disease are usually assumed to have TB and receive early anti-TB treatment [28]. Although TB incidence has declined in Taiwan, the rate of NTM infections has increased [15, 16]. We found that in HIV-positive patients, the potential for BM infections caused by MAC might be similar to that caused by MTB. Therefore, caution is warranted when initiating empirical anti-TB therapy based on results of BM histopathological examinations before a culture-proven diagnosis is established.

Mycobacterial BM infection is relatively uncommon in HIV-negative patients. A study in the USA found that mycobacterial species were isolated from only 2% of BM specimens from HIV-negative patients with FUO compared to 20% from HIV-positive patients [6]. Similarly, a study in France found that only 1.5% of HIV-negative patients with FUO had BM infections caused by MTB [1]. Furthermore, studies have shown that mycobacterial BM infections in HIV-negative patients are almost always caused by MTB [1, 6, 13, 28]. In our study, the numbers of mycobacterial BM infections in non-HIV-infected and HIV-infected patients were 15 and nine, respectively. Moreover, 40% of mycobacterial BM infections in non-HIV-infected patients were caused by NTM. These findings may most likely be attributed to the relatively low HIV prevalence in Taiwan [17]. Additionally, our findings were in agreement with the increasing secular trend of NTM infections and the emergence of disseminated NTM disease in Taiwan [8, 16]. In our study, six patients were

considered to be immunocompetent and in addition to MAC, *M. kansasii* was another important pathogen of BM infections in HIV-negative patients. These findings imply that mycobacterial BM infections should be considered in HIV-negative patients as well as in immunocompetent hosts.

Our study has several limitations. First, the retrospective nature of the study might underestimate the incidence of mycobacterial BM infections because the decision to perform BM examinations was at the physicians' discretion. Second, in Taiwan a consent form must be obtained before HIV testing and in our study three patients with a low index of suspicion did not receive HIV tests. Third, this study was conducted in a tertiary referral hospital in northern Taiwan. Therefore, the data are not fully representative of the general population. Forth, the relatively small number of patients in this study limited the statistical power. Fifth, there are miscellaneous causes of BM granuloma, such as infections, sarcoidosis, malignancies, and drugs [29]. Therefore, a diagnosis of mycobacterial BM infection without a positive mycobacterial BM culture result would be erroneous. Given the probability of culture-negative mycobacterial infections [9–11], this bias would be minimized by the coexistence of BM AFB or mycobacterial infections at other body sites. Finally, a clinical diagnosis of TB, rather than NTM infection, would be made without positive cultures and the comparison between MTB and NTM BM infections might be further biased by the conception that more MTB BM infections were diagnosed according to the second and third criteria.

In summary, MTB and NTM BM infections had similar frequencies in patients hospitalized in the tertiary-care hospital in Taiwan. In addition to MAC, *M. kansasii* was another important agent of NTM in HIV-negative patients. A comprehensive evaluation including regional TB epidemiology, patients' characteristics and immune status, and BM histopathological findings are imperative in the management of mycobacterial BM infections.

DECLARATION OF INTEREST

None.

REFERENCES

Hot A, et al. Yield of bone marrow examination in diagnosing the source of fever of unknown origin. Archives of Internal Medicine 2009; 169: 2018–2023.

- Benito N, et al. Bone marrow biopsy in the diagnosis of fever of unknown origin in patients with acquired immunodeficiency syndrome. Archives of Internal Medicine 1997; 157: 1577–1580.
- 3. van Schalkwyk WA, Opie J, Novitzky N. The diagnostic utility of bone marrow biopsies performed for the investigation of fever and/or cytopenias in HIV-infected adults at Groote Schuur Hospital, Western Cape, South Africa. *International Journal of Laboratory Hematology* 2011; 33: 258–266.
- 4. Santos ES, et al. The utility of a bone marrow biopsy in diagnosing the source of fever of unknown origin in patients with AIDS. Journal of Acquired Immune Deficiency Syndromes 2004; 37: 1599–1603.
- Luther JM, et al. Utility of bone marrow biopsy for rapid diagnosis of febrile illnesses in patients with human immunodeficiency virus infection. Southern Medical Journal 2000; 93: 692–697.
- Riley UB, et al. Detection of mycobacteria in bone marrow biopsy specimens taken to investigate pyrexia of unknown origin. Journal of Clinical Pathology 1995; 48: 706–709.
- Wang JY, et al. Disseminated tuberculosis: a 10-year experience in a medical center. Medicine 2007; 86: 39–46.
- Chou CH, et al. Clinical features and outcomes of disseminated infections caused by non-tuberculous mycobacteria in a university hospital in Taiwan, 2004–2008. Scandinavian Journal of Infectious Diseases 2011; 43: 8–14.
- The American Thoracic Society and the Centers for Disease Control and Prevention. Diagnostic standards and classification of tuberculosis in adults and children. American Journal of Respiratory and Critical Care Medicine 2000; 161: 1376–1395.
- Blumberg HM, et al. American Thoracic Society/ Centers for Disease Control and Prevention/Infectious Diseases Society of America: treatment of tuberculosis. American Journal of Respiratory and Critical Care Medicine 2003; 167: 603–662.
- Griffith DE, et al. An official ATS/IDSA statement: diagnosis, treatment, and prevention of nontuberculous mycobacterial diseases. American Journal of Respiratory and Critical Care Medicine 2007; 175: 367–416.
- 12. **Khandekar MM**, *et al*. Profile of bone marrow examination in HIV/AIDS patients to detect opportunistic infections, especially tuberculosis. *Indian Journal of Pathology and Microbiology* 2005; **48**: 7–12.
- Pacios E, et al. Evaluation of bone marrow and blood cultures for the recovery of mycobacteria in the diagnosis of disseminated mycobacterial infections. Clinical Microbiology and Infection 2004; 10: 734–737.
- Northfelt DW, et al. The usefulness of diagnostic bone marrow examination in patients with human immunodeficiency virus (HIV) infection. Journal of Acquired Immune Deficiency Syndromes 1991; 4: 659–666.
- 15. **Lo HY**, *et al.* Trends in tuberculosis in Taiwan, 2002–2008. *Journal of the Formosan Medical Association* 2011; **110**: 501–510.

- Lai CC, et al. Increasing incidence of nontuberculous mycobacteria, Taiwan, 2000–2008. Emerging Infectious Diseases 2010; 16: 294–296.
- 17. Yang CH, et al. Trends of mortality and causes of deaths among HIV-infected patients in Taiwan, 1984–2005. HIV Medicine 2008; 9: 535–543.
- 18. Lai CC, et al. Emergence of disseminated infections due to nontuberculous mycobacteria in non-HIV-infected patients, including immunocompetent and immunocompromised patients in a university hospital in Taiwan. Journal of Infection 2006; 53: 77–84.
- Roberts GD, Koneman EW, Kim YK. Mycobacterium.
 In: Balows A, Hausler Jr. WJ, Herrmann KL, Isenberg HD, Shadomy K, eds. Manual of Clinical Microbiology, 5th edn. Washington: American Society for Microbiology, 1991, pp. 304–339.
- Tan CK, et al. Mycobacterial bacteraemia in patients infected and not infected with human immunodeficiency virus, Taiwan. Clinical Microbiology and Infection 2010; 16: 627–630.
- Ker CC, et al. Comparison of bone marrow studies with blood culture for etiological diagnosis of disseminated mycobacterial and fungal infection in patients with acquired immunodeficiency syndrome. *Journal* of Microbiology, Immunology and Infection 2002; 35: 89–93.
- Hafner R, et al. Correlation of quantitative bone marrow and blood cultures in AIDS patients with disseminated Mycobacterium avium complex infection. Journal of Infectious Diseases 1999; 180: 438–447.
- 23. Kilby JM, et al. The yield of bone marrow biopsy and culture compared with blood culture in the evaluation of HIV-infected patients for mycobacterial and fungal infections. American Journal of Medicine 1998; 104: 123–128.
- 24. Nichols L, et al. Bone marrow examination for the diagnosis of mycobacterial and fungal infections in the acquired immunodeficiency syndrome. Archives of Pathology & Laboratory Medicine 1991; 115: 1125–1132.
- Akpek G, et al. Bone marrow aspiration, biopsy, and culture in the evaluation of HIV-infected patients for invasive mycobacteria and histoplasma infections. American Journal of Hematology 2001; 67: 100–106.
- 26. **Escobedo-Jaimes L, et al.** Evaluation of the polymerase chain reaction in the diagnosis of miliary tuberculosis in bone marrow smear. *International Journal of Tuberculosis and Lung Disease* 2003; 7: 580–586.
- Lai CC, et al. Diagnostic performance of whole-blood interferon-gamma assay and enzyme-linked immunospot assay for active tuberculosis. *Diagnostic Micro*biology and Infectious Disease 2011; 71: 139–143.
- 28. **Rose PC**, *et al*. Value of bone marrow biopsy in children with suspected disseminated mycobacterial disease. *International Journal of Tuberculosis and Lung Disease* 2011; **15**: 200–204.
- Brackers de Hugo L, et al. Granulomatous lesions in bone marrow: clinicopathologic findings and significance in a study of 48 cases. European Journal of Internal Medicine 2013; 24: 468–473.