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Energy intake and resting energy expenditure in adult male rats after early postnatal food restriction

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Both in man and in animal models, changes in food intake and body composition in later life have been reported after alterations in perinatal nutrition. Therefore, we hypothesised that early postnatal undernutrition in the rat induces permanent changes in energy balance. Food restriction (FR) during lactation was achieved by enlarging litter size to twenty pups, whereas control animals were raised in litters containing ten pups. Energy intake and resting energy expenditure were determined in adult males. Early postnatal FR resulted in acute growth restriction followed by incomplete catch-up in body weight, body length and BMI. At the age of 12 months, middle-aged FR males had significantly lower absolute resting energy expenditure (200 v. 216 kJ/24 h, P=0·009), absolute energy intake (281 v. 310 kJ/24 h, P=0·001) and energy intake adjusted for BMI (284 v. 305 kJ/24 h, P=0·016) than controls, whereas resting energy expenditure adjusted for BMI did not differ significantly between the groups (204 v. 211 kJ/24 h, P=0·156). The amount of energy remaining for other functions was lower in FR males (80 v. 94 kJ/24 h, P=0·044). Comparable data were obtained at the age of 6 months. These results indicate that in rats energy balance can be programmed by early nutrition. A low early postnatal food intake appears to programme these animals for a low energy intake and to remain slender in adult life.

Developmental programming: Energy balance: Litter size: Rat

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Epidemiological studies that linked low birth weight with later disease¹ have led to the theory of developmental programming². The early environment, encountered during a sensitive period, is believed to influence the development and hence the function of the organism permanently³. Humans that are born small for gestational age represent an example of developmental programming. These individuals are thought to be adapted to a poor environment, and when confronted with a rich, western, environment they have an increased risk of insulin resistance, hypertension, obesity and CVD (collectively called the metabolic syndrome) in adult life^{1,2}. Other examples of developmental programming are various animal models that manipulate the perinatal nutritional environment⁴⁻⁶. Studies using these models have shown that, depending on the exact nature and timing of the manipulation, programming can act in different directions. For instance, different effects on energy balance have been reported after perinatal malnutrition⁷.

In different rat models, adult food intake⁸⁻¹¹ and body fat^{9,11-14} were either increased, decreased or unchanged. In man, programming of energy balance has also been reported. Although obesity rates have been reported to be lower after perinatal malnutrition¹⁵, there are now several

studies that associate low birth weight with a more central distribution of fat^{16,17} and a lower lean body mass¹⁸.

An important part of the regulation of energy balance takes place in the hypothalamus. Whereas in man, a substantial part of the development of the hypothalamus and the brain is completed *in utero*, in rats much of this development occurs postnatally ^{19–22}. Therefore, we have used early postnatal food restriction (FR) in rats to study developmental programming of energy balance. We have previously shown that raising rats in large litters reduced body weight into adulthood ^{12,23} and decreased the fat percentage in adult males ¹². These animals also showed disruptions in several processes that are regulated by the hypothalamus; a delayed onset of puberty²⁴, impaired testicular function²⁵ and changes in the growth hormone axis²⁶.

If the hypothalamus is affected in these animals, then its regulation of energy homeostasis may also be affected, which could ultimately lead to permanently altered energy balance. Changes in energy balance might contribute to the phenotype of these animals. Therefore, the aim of the present study was to elucidate whether early postnatal FR alters energy intake and resting energy expenditure (REE) in adult and middle-aged male rats.

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Methods

Experimental animals

Primiparous timed-pregnant Wistar rats (Harlan, Horst, The Netherlands) arrived on day 14 or 15 of gestation and were housed individually under controlled lighting (12h light, 12 h dark) and temperature (21.5 (SD 0.5) °C). Animals had unlimited access to tap water and standard rat chow (Ssniff R/M-H; Bio Services, Uden, The Netherlands; 12.8 kJ/g metabolisable energy, 19.0% protein, 3.3% fat, 36.5% starch, 4.7 % sugar and 4.9 % crude fibre), unless mentioned otherwise. Pups were born spontaneously on day 22 or 23 of gestation. From day 20 of gestation, the presence of pups was checked daily in the morning and the first day of life was designated postnatal day 1. On day 2, male and female pups were allocated to either a control litter of ten pups or a FR litter of twenty pups using computer-generated random numbers. Male-to-female ratio was 1:1 in all fostered litters. In large FR litters, less milk has been shown to be available per pup than in control litters, resulting in undernutrition²⁷. On day 25, the pups were weaned and males were housed two per cage, paired with another animal of the same experimental group. Subsets of animals were killed at different ages for another study. A subset of thirty-nine of the male animals in the experiment survived until the age of 1 year and were used in the present study (sixteen controls and twentythree FR animals). All procedures were approved by the Animal Experimentation Ethics Committee of the Vrije Universiteit and the VU University Medical Center in Amsterdam, The Netherlands.

Body dimensions

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Body weight was measured regularly throughout life. At the age of 12 months (day 380), body length was measured from the tip of the nose to the anus under pentobarbital or O_2/CO_2 anaesthesia before the animals were killed for further study. BMI was calculated as the ratio of body weight (g) to body length (cm) squared. In a previous study, we showed that at the age of 6 months control and FR males had a fatfree mass (FFM) of, respectively, 76 and 81% of their body weight 12. Therefore, an estimate of FFM at 6 months was calculated as $0.76 \times$ body weight in controls and $0.81 \times$ body weight in FR males.

Food intake

Individual food intake was determined in sixteen adult and middle-aged control animals and twenty-three FR animals at the ages of 6 and 12 months. The animals were housed individually at least 1 d before the measurement to become accustomed to the testing cage. The food provided was weighed at the beginning and the end of a 24 h period. Individual energy intake was calculated from 24 h food intake and the energy density of the diet (12·8 kJ/g).

Indirect calorimetry

REE was determined by means of indirect calorimetry in the same sixteen control and twenty-three FR animals at the ages of 6 and 12 months. The animals were housed individually at least 1 d before the measurement to become accustomed to the testing cage. All measurements were carried out during the light, inactive, phase of the day. During the measurements, no food and water were available. A metabolic monitor (Deltatrac II MBM-200; Datex-Ohmeda, Helsinki, Finland), adapted to fit the animal cages, was used to measure resting $V_{\rm O2}$ and carbon dioxide production rate ($V_{\rm CO2}$) every minute. The lower limit for reliable measurements was 5 ml/min for both $V_{\rm O2}$ and $V_{\rm CO2}$, restricting us to the measurement of adult males; neither females nor younger animals reached this limit of reliability. Before each measurement, the metabolic monitor was calibrated with a gas mixture of 95% O₂ and 5% CO₂. Mean V_{O2} and V_{CO2} values from stable measurements with a duration of at least 20 min and a CV \leq 5 % were used for calculations. REE was calculated using the modified Weir formula²⁸: REE (kJ/24 h) = $4\cdot184 \times (5\cdot50 \times V_{O2} \text{ (ml/min)} + 1\cdot76 \times V_{CO2} \text{ (ml/min)})$, without adjustment for urinary nitrogen excretion²⁹. To avoid possible effects of circadian rhythm on energy expenditure interfering with the group effects, control and FR animals were measured in an alternating manner. After the two energy balance measurements were completed, the animals were socially housed with the same individual as before.

Data analysis

The results were analysed using Statistical Product and Service Solutions software for Windows, version 12 (SPSS Inc., Chicago, IL, USA). All data were checked for normality and are expressed as means with their standard errors (except in Figs. 1 and 2, where standard deviations are shown for better visibility). After exclusion of animals with missing values or a CV > 5% in the indirect calorimetry, data were analysed for fourteen control and twenty-two adult FR males at 6 months and for sixteen control and twenty-one middle-aged FR males at 12 months. All outcome measures were initially analysed by means of one-way ANOVA. To confirm that the FR in the FR litters was distributed evenly over the pups within a litter, differences in variance of preweaning body weight between the groups were tested using Levene's test for homogeneity of variances. Potential confounding effects of biological and foster dams were tested in univariate ANOVA. Foster dam nested within group and the interaction between biological dam and group had no long-lasting significant effect and were omitted in further analyses. Energy utilisation is known to correlate with body size, and more specifically with FFM, and it has been recommended to adjust for FFM in an ANOVA when comparing energy utilisation between subjects with different body compositions^{30,31}. In a previous study we have shown that at the age of 6 months FR males indeed have a different body composition than controls¹², confirming the need for adjustment. At 6 months, we estimated FFM by means of the values found in this previous study. At 12 months, BMI was available as another estimate of body composition. Therefore, energy balance data were tested in a univariate ANOVA with estimated FFM (eFFM) as a covariate at 6 months and BMI at 12 months, as recommended 30,31. If these covariates did not have a significant effect, they were omitted from the analysis.

Results

The thirty-nine animals used in the present study were born from twenty-one of the thirty-three dams in the complete experiment and on day 2 were fostered to six different dams for each group. The original litter size of the foster dams nurturing FR pups (12·3 (SEM 1·0) pups) was not different from that of the foster dams that nurtured control pups (11·7 (SEM 0·8) pups, P>0.600). Nor did the original litter size of FR pups (12·0 (SEM 0·5) pups) differ from that of control pups (12·2 (SEM 0·5) pups, P>0.800). Of the thirty-nine pups in this study, 85% were cross-fostered, whereas 15% (three control and three FR animals) remained with the same dam after the random redistribution on day 2.

Early postnatal FR resulted in a persistent reduction in body weight, body length and BMI. Mean body weights of control and FR rats are shown in Fig. 1. Body weight on day 2 (before the redistribution into control and FR litters) was 7.7 (SEM 0·13) g. Body weight was lower in FR rats from day 4 until day 380 (P < 0.001). Relative to control values, body weight of FR animals decreased during lactation to 60% at weaning. After weaning, relative body weight of FR rats increased to 86 % on day 70 and then stabilised so that on day 380 FR animals weighed 89% of control weight (Fig. 2). During the lactation period, the variance in body weight did not differ significantly between the groups (P > 0.200), although on day 21 there was a trend towards larger variance in the FR group (P=0.093). Body dimensions of FR and control rats at 6 and 12 months are shown in Table 1. At both 6 and 12 months, body weight was lower in FR males (P < 0.001). At 6 months, eFFM, which was computed as 76 % of body weight in controls and 81 % of body weight in FR rats, was lower in FR animals (P=0.029). At 12 months, body length (P<0.001) and BMI (P=0.024) were lower in FR animals than in controls.

At 6 months, we could not obtain measurements with a CV below 5% for three animals (two controls and one FR rat), despite repeated attempts. These animals were excluded from all analyses at this time-point. At 12 months, two FR animals had to be excluded; one had to be killed prematurely, one had missing body length data at the time of killing.

Energy intake

At both 6 and 12 months, FR animals consumed a significantly smaller absolute amount of food than control animals (Table 2). Energy intake correlated with estimated body composition at both 6 months (eFFM, R 0-699, P<0-001) and 12 months (BMI, R 0-540, P=0-001). Energy intake was adjusted for eFFM at 6 months and for BMI at 12 months to account for differences in body composition between the groups. Adjusted energy intake at 6 months (Fig. 3 (A)) was lower in FR males (285-0 (sem 4-2) kJ/24 h) than in control males (303-4 (sem 5-3) kJ/24 h, P=0-012). At 12 months, adjusted energy intake (Fig. 3 (B)) was also lower in FR rats (284-4 (sem 5-1) kJ/24 h) than in controls (304-9 (sem 5-9) kJ/24 h, P=0-016).

Resting energy expenditure

Mean values for $V_{\rm O2}$ and $V_{\rm CO2}$ were 6.5 (SEM 0.1) and 6.0 (SEM 0.1) ml/min at 6 months and 6.9 (SEM 0.1) and 6.5 (SEM 0.1) ml/min at 12 months, respectively.

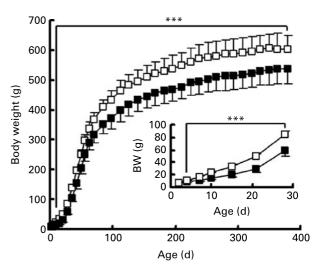


Fig. 1. Body weight (BW) throughout the experiment and during the first month of life (inset) of male food restriction (FR) rats (\blacksquare , n 23), which were food restricted during lactation, and male control rats (\square , n 16). Values are means with their standard deviations depicted by vertical bars (where the error bars are not visible, they are within the symbol). Mean values of the FR group were significantly different from those of the control group: ***P< 0.001 from day 4 until 380.

At both 6 and 12 months, FR animals had a significantly lower absolute REE than control animals (Table 2). REE correlated with estimated body composition at both 6 months (eFFM, R 0-870, P<0-001) and 12 months (BMI, R 0-680, P<0-001). At 6 months, energy expenditure adjusted for estimated body composition (Fig. 4(A)) was not significantly different between FR males (192-0 (sem 2-2) kJ/24 h) and controls (198-1 (sem 2-8) kJ/24 h, P=0-099), nor did adjusted energy expenditure at 12 months (Fig. 4(B)) differ significantly between FR rats (204-1 (sem 3-2) kJ/24 h) and controls (211-3 (sem 3-6) kJ/24 h, P=0-156).

Energy intake minus resting energy expenditure

When energy balance is neutral, energy intake equals total energy expenditure, so the difference between energy intake and REE represents the amount of energy available for other functions such as locomotor activity. This parameter did not correlate with BMI or eFFM (P>0.480). Energy intake

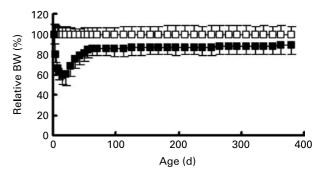


Fig. 2. Body weight (BW) expressed as a percentage of control body weight for food restriction animals (\blacksquare , n 23) and control animals (\square , n 16). Values are means with their standard deviations depicted by vertical bars.

Table 1. Body dimensions of control and food restriction (FR) males at the ages of 6 and 12 months (Mean values with their standard errors)

	6 months				12 months				
	Control (n 14)		FR (n 22)		Control (n 16)		FR (n 21)		
	Mean	SEM	Mean	SEM	Mean	SEM	Mean	SEM	
BW (g) eFFM (g) BL (cm) BMI (g/cm ²)	526-4 400-0 ND ND	12·5 9·5	461·1*** 373·5* ND ND	8·7 7·1	604·2 ND 27·1 0·82	11·9 0·2 0·01	538·4*** ND 26·1*** 0·79*	11.4 0.1 0.01	

BL, body length; BW, body weight; eFFM, estimated fat-free mass; ND, no data.

Mean values were significantly different from those of the control group at the same age: *P<0.05, ***P<0.001.

minus REE was lower in FR rats than in control rats at both 6 (P=0.038) and 12 months (P=0.044; Table 2).

Discussion

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In the present study, early postnatal FR of male rats resulted in an acute reduction in growth, followed by incomplete catch-up growth, and permanently altered energy balance. At both 6 and 12 months, FR rats consumed and expended less energy than controls. After subtraction of REE from energy intake, FR animals had less energy available for other functions. When estimated adult body composition was taken into account, energy intake was lower after FR, whereas energy expended in rest was similar to that of controls.

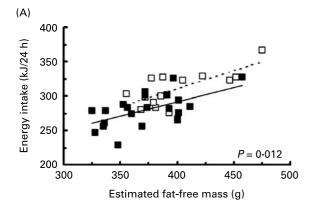
Programming of energy balance. In the present study, male FR rats remained lighter than control males until the end of the experiment at the age of 12 months. This suggests that early postnatal FR can programme later size. This is in contrast to FR later in life, which has been shown to induce reversible growth restriction with complete catch-up^{32,33}. BMI was also reduced in FR animals. Although BMI is not a direct measure for fat mass, it is strongly correlated with the percentage of body fat in both man³⁴ and rats³⁵. Therefore, the present results suggest that at least in rats early postnatal FR can programme a low level of adult adiposity. This may be through a reduced energy intake, but from the present data it is not possible to discern cause and effect in the relationship between BMI and food intake. Although REE was reduced in the FR animals, it seemed appropriate for the altered body composition. Therefore, programming of REE does not seem to have taken place. In the case of neutral energy balance, energy intake equals total energy expenditure. Since REE includes BMR, the thermic effect of food and energy expended for growth, the difference between total energy expenditure and REE represents activity-related energy expenditure. Energy intake minus REE, or activity-related energy expenditure, was reduced in FR males. Therefore, these animals may either be less active or expend less energy during their activity. In adult rats, the energy expended for growth is negligible. If during the development of these animals, energy intake was also reduced without a change in BMR, there may have been less energy available for growth. This may explain, at least in part, the permanent reduction in body weight, body length and BMI in the animals in the present study.

Early and late effects. When analysing data on late effects of early insults, it is important to separate the effects of the early insult from those of events later in life³⁷. Therefore, in the present study both unadjusted data and data adjusted for estimated adult body composition were presented. Energy intake and REE were both reduced in FR animals when early size (i.e. control or FR) was the sole independent variable. Adding estimated adult body composition as a covariate removed the effect on REE, but not that on energy intake. This suggests that later events may have been more important in determining REE than early postnatal FR, but that the FR was the most important determinant of energy intake in these animals. Here it should be noted that the adjustment for FFM as advised^{30,31} is essential for this result. When adjusted for the less recommended crude body weight instead of the metabolically active FFM, energy intake was not significantly different between the groups (data not shown). This emphasises the importance

Table 2. Energy intake (EI), resting energy expenditure (REE) and EI minus REE of control and food restriction (FR) males at the ages of 6 and 12 months (Mean values with their standard errors)

		6 m	onths		12 months			
	Control (n 14)		FR (n 22)		Control (n 16)		FR (n 21)	
	Mean	SEM	Mean	SEM	Mean	SEM	Mean	SEM
EI (kJ/24 h) REE (kJ/24 h) EI – REE (kJ/24 h)	310·9 205·6 105·2	6·7 5·5 4·1	280·3** 187·2** 93·1*	5·1 3·6 3·6	309·8 216·2 93·6	23·8 4·1 4·5	280·7** 200·3** 80·4*	25·4 4·0 4·3

Mean values were significantly different from those of the control group at the same age: $^*P < 0.05, ^{**}P < 0.01$.



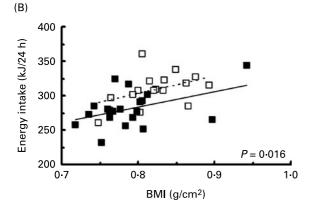
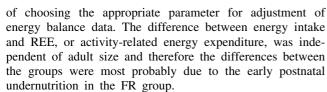
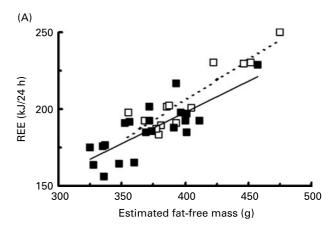


Fig. 3. Relationship between estimated body composition and energy intake at 6 months (A) and 12 months (B) in control (\square , - - -, n 14 in (A), n 16 in (B)) and food restriction rats (\blacksquare , —, n 22 in (A), n 21 in (B)). The given P values indicate a significant effect of experimental group on energy intake when the estimated fat-free mass was included as a covariate.



If the differences between the groups are to be attributed to true programming, the effects must be permanent. Therefore, the animals were tested in adulthood. Animals were retested when middle-aged at the age of 1 year to verify whether the effects were truly permanent. Since similar results were obtained at both ages studied, we are rather confident that permanent programming really occurred.

Energy balance in other models. Postnatal manipulations of litter size appear to yield consistent results. Other studies using large litters have also found a permanently reduced body weight^{8,10,12,13,33} and fat mass^{12,13}, a lower food intake in young adulthood⁸ and a far lower cumulative absolute food intake from weaning until over a year of age¹⁰. Studies using overfeeding in small litters have found opposite results: animals were permanently heavier than control animals^{8,38,39}, had an increased fat mass or BMI^{38,39} and had a larger absolute food intake in young adulthood^{8,38,39}. New in the present study is that food intake of male FR rats was not only significantly lower in absolute terms, but it was even reduced when their altered body composition was taken into account.



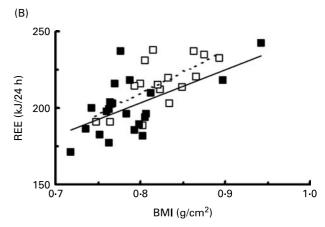


Fig. 4. Relationship between estimated body composition and resting energy expenditure (REE) at 6 months (A) and 12 months (B) in control (\square , - - -, n 14 in (A), n 16 in (B)) and food restriction rats (\blacksquare , —, n 22 in (A), n 21 in (B)).

In comparison with the present observation of an appropriately reduced REE, a previous study using early postnatally overfed small litter male rats showed increased total energy expenditure at the age of 5 weeks, but not in older animals⁴⁰.

Comparing the results of the present study with those of others that used different models of perinatal undernutrition is more complicated, however, as the direction of the changes observed appears to be highly dependent on the exact timing, type and severity of malnutrition⁴⁻⁶. Maternal 'caloric' and protein restriction during gestation or lactation have produced an increased, reduced or normal body weight and fat mass in adulthood^{5,7,9,11,14}, depending on the timing and severity of malnutrition. Moreover, 50 % FR of the dam during gestation increased adult food intake, but the same insult during lactation did not⁹, whereas a low-protein diet during lactation reduced adult food intake¹¹. In general, a lower food intake has been found in models with incomplete catch-up growth, whereas a higher food intake was found after postnatal overnutrition or prenatal undernutrition followed by overcomplete catch-up. Studies using prenatal maternal malnutrition have found reductions in total energy expenditure with unaltered REE (suggesting reduced activity-related energy expenditure)⁴¹ and an actual reduction in activity levels^{42,43} in adult males, a consequence also suggested by the results of the present study. In contrast, activity was not reduced in our previous study

in young adult males that were prenatally growth restricted by bilateral uterine artery ligation⁴⁴.

Unlike the early postnatally food-restricted rats, most of the humans that are born small for gestational age or after intra-uterine growth restriction catch up during infancy⁴⁵. However, it was shown that prepubertal children born small for gestational age that did not catch up had a food intake below the recommended energy intake for their age⁴⁶. These data seem to be in accord with the reduced food intake in the early postnatally food-restricted rats with incomplete catch-up growth in the present study. Studies in neonates have suggested that infants that are born small for gestational age have a higher energy expenditure per kg body weight or FFM than weight-matched controls^{47,48}. Although these data on REE relate to acute instead of long-term effects, they do indicate that perinatal malnutrition can also affect energy expenditure in man.

The differences outlined above warn us to exert extreme caution when attempting to extrapolate outcomes of perinatal malnutrition, not only between rats and man, but also between different animal models. Seemingly comparable manipulations of pre- or early postnatal nutrition can yield widely differing results⁴⁻⁶.

Because of the different timing of birth relative to development, early postnatal FR in rats is probably somewhat similar to undernutrition in human fetuses during the third trimester, although the potential for catch-up growth is evidently different between the two. It could be speculated that the window of plasticity for body dimensions, adiposity and food intake may close before the end of the lactation period in rats, whereas in man it may extend into the postnatal period.

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Technical considerations. A concern when using large litters to reduce early postnatal food intake is the lack of control over the distribution of the available milk within litters. There may be competition between the pups over the milk supply and as a consequence the pups in a litter may be food restricted to different degrees⁴⁹. The fact that the variance in body weight during the lactation period was similar between control and FR males suggested that in the present study all FR pups were food restricted roughly to the same degree.

The energy balance measurements in the present study were restricted to adult male rats. Investigating the possible effects of FR on energy expenditure in females would be interesting. Unfortunately, we were unable to investigate this, because of the limitations of the metabolic monitor.

In the present study, actual measurements of FFM were not available. The variables eFFM and BMI were chosen as estimates for FFM. By extrapolating the percentage of FFM from one population to another, we introduced an uncertainty. Especially because at 6 months the population of the present study was heavier than that used in the other study 12, most probably because of the different diets the animals received. Therefore, energy intake and REE were also determined at another age, when BMI was available as a parameter of body composition. Although it is usually employed for its correlation with fat mass, BMI describes body weight relative to length, and hence does not discriminate between fat mass and FFM. It therefore also increases with increasing FFM⁵⁰. The fact that the analyses using these different covariates

produced comparable results at both ages suggests that the estimates BMI and eFFM were equally suitable approximations for FFM.

Implications. In the present study, we showed that male rats that were food restricted early postnatally remained lean with a reduced food intake in adult life. This fits in with the relatively recent idea that promoting catch-up growth in low birth weight infants may not be beneficial for their long-term outcome. Several studies in man as well as in animals have suggested that fast and early catch-up, sometimes through super-nutritious food, can be detrimental ^{14,43,51,52}. On the other hand, rats with this modest phenotype may not have sufficient supplies for normal growth ^{23,26} and possibly other matters such as reproduction ^{24,25} and locomotor activity. In summary, the present study demonstrates that in rats early postnatal FR can programme energy balance in later life. The present study provides additional support for the hypothesis that early nutritional insults may have long-term metabolic consequences.

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References

- Hales CN & Barker DJ (2001) The thrifty phenotype hypothesis. Br Med Bull 60, 5–20.
- Gluckman PD & Hanson MA (2004) The developmental origins of the metabolic syndrome. Trends Endocrinol Metab 15, 183–187.
- 3. Harding JE (2001) The nutritional basis of the fetal origins of adult disease. *Int J Epidemiol* **30**, 15–23.
- Bertram CE & Hanson MA (2001) Animal models and programming of the metabolic syndrome. Br Med Bull 60, 103-121.
- Holemans K, Aerts L & Van Assche FA (2003) Fetal growth restriction and consequences for the offspring in animal models. J Soc Gynecol Investig 10, 392–399.
- 6. Ozanne SE (2001) Metabolic programming in animals. *Br Med Bull* **60**, 143–152.
- McMillen IC, Adam CL & Muhlhausler BS (2005) Early origins of obesity: programming the appetite regulatory system. *J Physiol* 565, 9–17.
- Bassett DR & Craig BW (1988) Influence of early nutrition on growth and adipose tissue characteristics in male and female rats. J Appl Physiol 64, 1249–1256.
- Desai M, Gayle D, Babu J & Ross MG (2005) Programmed obesity in intrauterine growth-restricted newborns: modulation by newborn nutrition. *Am J Physiol Regul Integr Comp Physiol* 288, R91–R96.
- Oscai LB & McGarr JA (1978) Evidence that the amount of food consumed in early life fixes appetite in the rat. Am J Physiol 235. R141–R144.
- 11. Zambrano E, Bautista CJ, Deas M, Martinez-Samayoa PM, Gonzalez-Zamorano M, Ledesma H, Morales J, Larrea F & Nathanielsz PW (2006) A low maternal protein diet during pregnancy and lactation has sex- and window of exposure-specific effects on offspring growth and food intake, glucose

1155

- metabolism and serum leptin in the rat. J Physiol 571, 221 - 230
- 12. Engelbregt MJ, Van Weissenbruch MM, Lips P, Van Lingen A, Roos JC & Delemarre-van de Waal HA (2004) Body composition and bone measurements in intra-uterine growth retarded and early postnatally undernourished male and female rats at the age of 6 months: comparison with puberty. Bone 34, 180 - 186.
- Faust IM, Johnson PR & Hirsch J (1980) Long-term effects of early nutritional experience on the development of obesity in the rat. J Nutr 110, 2027-2034.
- Vickers MH, Breier BH, Cutfield WS, Hofman PL & Gluckman PD (2000) Fetal origins of hyperphagia, obesity, and hypertension and postnatal amplification by hypercaloric nutrition. Am J Physiol Endocrinol Metab 279, E83-E87.
- Ravelli GP, Stein ZA & Susser MW (1976) Obesity in young men after famine exposure in utero and early infancy. N Engl J Med 295, 349-353.
- Fall CH, Osmond C, Barker DJ, Clark PM, Hales CN, Stirling Y & Meade TW (1995) Fetal and infant growth and cardiovascular risk factors in women. Br Med J 310, 428-432.
- Valdez R, Athens MA, Thompson GH, Bradshaw BS & Stern MP (1994) Birthweight and adult health outcomes in a biethnic population in the USA. Diabetologia 37, 624-631.
- Gale CR, Martyn CN, Kellingray S, Eastell R & Cooper C (2001) Intrauterine programming of adult body composition. J Clin Endocrinol Metab 86, 267-272.
- Dobbing J & Sands J (1979) Comparative aspects of the brain growth spurt. Early Hum Dev 3, 79-83.
- Bouret SG, Draper SJ & Simerly RB (2004) Formation of projection pathways from the arcuate nucleus of the hypothalamus to hypothalamic regions implicated in the neural control of feeding behavior in mice. J Neurosci 24, 2797 - 2805.
- Grove KL, Allen S, Grayson BE & Smith MS (2003) Postnatal development of the hypothalamic neuropeptide Y system. Neuroscience 116, 393-406.
- Koutcherov Y, Mai JK, Ashwell KWS & Paxinos G (2002) Organization of human hypothalamus in fetal development. J Comp Neurol 446, 301-324.
- Huizinga CT, Engelbregt MJ, Rekers-Mombarg LT, Vaessen SF, Delemarre-van de Waal HA & Fodor M (2004) Ligation of the uterine artery and early postnatal food restriction animal models for growth retardation. Horm Res 62, 233-240.
- Engelbregt MJ, Houdijk ME, Popp-Snijders C & Delemarre-van de Waal HA (2000) The effects of intra-uterine growth retardation and postnatal undernutrition on onset of puberty in male and female rats. Pediatr Res 48, 803-807.
- Van Weissenbruch MM, Engelbregt MJ, Veening MA & Delemarre-van de Waal HA (2005) Fetal nutrition and timing of puberty. Endocr Dev 8, 15-33.
- Houdijk ME, Engelbregt MT, Popp-Snijders C & Delemarrevan de Waal HA (2003) Long-term effects of early postnatal food restriction on growth hormone secretion in rats. J Parenter Enteral Nutr 27, 260-267.
- 27. Fiorotto ML, Burrin DG, Perez M & Reeds PJ (1991) Intake and use of milk nutrients by rat pups suckled in small, medium, or large litters. Am J Physiol 260, R1104-R1113.
- Weir JB de V (1949) New methods for calculating metabolic rate with special reference to protein metabolism. J Physiol **109**. 1-9.
- Even PC, Mokhtarian A & Pele A (1994) Practical aspects of indirect calorimetry in laboratory animals. Neurosci Biobehav Rev 18, 435-447.
- Arch JR, Hislop D, Wang SJ & Speakman JR (2006) Some mathematical and technical issues in the measurement

- and interpretation of open-circuit indirect calorimetry in small animals. Int J Obes (Lond) 30, 1322-1331.
- 31. Toth MJ (2001) Comparing energy expenditure data among individuals differing in body size and composition: statistical and physiological considerations. Curr Opin Clin Nutr Metab Care 4, 391-397.
- Hughes PC (1982) Morphometric studies of catch-up growth in the rat. Prog Clin Biol Res 101, 433-446.
- Widdowson EM & McCance RA (1963) The effect of finite periods of undernutrition at different ages on the composition and subsequent development of the rat. Proc R Soc Lond B Biol Sci 158, 329-342.
- 34. Kopelman PG (2000) Obesity as a medical problem. Nature 404, 635-643.
- Novelli EL, Diniz YS, Galhardi CM, Ebaid GM, Rodrigues HG, Mani F, Fernandes AA, Cicogna AC & Novelli Filho JL (2007) Anthropometrical parameters and markers of obesity in rats. Lab Anim 41, 111-119.
- Wenk C, Colombani PC, Van Milgen J & Lemme A (2001) Glossary: Terminology in animal and human energy metabolism. In Energy Metabolism in Animals: Proceedings of the 15th Symposium on Energy Metabolism in Animals, pp. 409-421 [A Chwalibog and K Jakobsen, editors]. Wageningen: Wageningen Pers.
- Lucas A, Fewtrell MS & Cole TJ (1999) Fetal origins of adult disease - the hypothesis revisited. Br Med J 319, 245-249.
- Boullu-Ciocca S, Dutour A, Guillaume V, Achard V, Oliver C & Grino M (2005) Postnatal diet-induced obesity in rats upregulates systemic and adipose tissue glucocorticoid metabolism during development and in adulthood: its relationship with the metabolic syndrome. Diabetes 54, 197-203.
- 39. Plagemann A, Harder T, Rake A, Voits M, Fink H, Rohde W & Dorner G (1999) Perinatal elevation of hypothalamic insulin, acquired malformation of hypothalamic galaninergic neurons, and syndrome X-like alterations in adulthood of neonatally overfed rats. Brain Res 836, 146-155.
- 40. Wiedmer P, Klaus S & Ortmann S (2002) Energy metabolism of young rats after early postnatal overnutrition. Br J Nutr 88, 301 - 306
- 41. Daenzer M, Ortmann S, Klaus S & Metges CC (2002) Prenatal high protein exposure decreases energy expenditure and increases adiposity in young rats. J Nutr 132, 142-144.
- Bellinger L, Sculley DV & Langley-Evans SC (2006) Exposure to undernutrition in fetal life determines fat distribution, locomotor activity and food intake in ageing rats. Int J Obes (Lond) 30, 729-738.
- Vickers MH, Breier BH, McCarthy D & Gluckman PD (2003) Sedentary behavior during postnatal life is determined by the prenatal environment and exacerbated by postnatal hypercaloric nutrition. Am J Physiol Regul Integr Comp Physiol 285, R271-R273.
- Schreuder MF, Fodor M, Van Wijk JA & Delemarre-van de Waal HA (2006) Association of birth weight with cardiovascular parameters in adult rats during baseline and stressed conditions. Pediatr Res 59, 126-130.
- 45. Karlberg J & Albertsson-Wikland K (1995) Growth in full-term small-for-gestational-age infants: from birth to final height. Pediatr Res 38, 733-739.
- 46. Boonstra VH, Arends NJ, Stijnen T, Blum WF, Akkerman O & Hokken-Koelega AC (2005) Food intake of children with short stature born small for gestational age before and during a randomized GH trial. Horm Res 65, 23-30.
- 47. Cauderay M, Schutz Y, Micheli JL, Calame A & Jequier E (1988) Energy-nitrogen balances and protein turnover in small and appropriate for gestational age low birthweight infants. Eur J Clin Nutr 42, 125-136.

48. Davies PS, Clough H, Bishop NJ, Lucas A, Cole JJ & Cole TJ (1996) Total energy expenditure in small for gestational age infants. *Arch Dis Child Fetal Neonatal Ed* **75**, F46–F48.

- 49. Galler JR & Turkewitz G (1975) Variability of the effects of rearing in a large litter on the development of the rat. *Dev Psychobiol* **8**, 325–331.
- Kyle UG, Schutz Y, Dupertuis YM & Pichard C (2003) Body composition interpretation. Contributions of the fat-free mass index and the body fat mass index. *Nutrition* 19, 597–604.
- 51. Ong KK, Ahmed ML, Emmett PM, Preece MA & Dunger DB (2000) Association between postnatal catch-up growth and obesity in childhood: prospective cohort study. *Br Med J* **320**, 967–971.
- Stettler N, Stallings VA, Troxel AB, Zhao J, Schinnar R, Nelson SE, Ziegler EE & Strom BL (2005) Weight gain in the first week of life and overweight in adulthood: a cohort study of European American subjects fed infant formula. *Circulation* 111, 1897–1903.

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