Effect of glycomacropeptide fractions on cholecystokinin and food intake

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Glycomacropeptide (GMP) is the hydrophilic 64-amino acid C-terminal glycopeptide released into cheese whey when κ-casein is cleaved by chymosin. GMP exists as a mixture of different glycoforms due to the carbohydrates sialic acid (*N*-acetylneuraminic acid, Neu/Nac), galactose (Gal), galactosamine and glucosamine attached by *O*-glycosidic linkages. GMP reportedly stimulates the release of cholecystokinin (CCK), which may promote satiety. The objectives of the present study were to manufacture three glycoforms of GMP, minimally glycosylated GMP (3·5 (sp 0·1)% Neu/Nac and 1·5 (sp 0·1)% Gal), glycosylated GMP (12·0 (sp 0·3)% Neu/Nac and 4·2 (sp 0·2)% Gal) and a GMP-depleted whey protein concentrate, and to assess the effects of these fractions relative to glucose on CCK, subjective measures of satiety and food intake. In a randomised double-blind acute study, twenty overweight/obese males (56·9 (sp 7·2) years, 97·4 (sp 8·1) kg, 31·5 (sp 3·0) kg/m²) were recruited to consume four 50 g preloads (two GMP preparations, GMP-depleted whey and glucose) containing 895 kJ. Blood samples and subjective measures of satiety were collected before and at 15, 30, 60, 90, 120 and 180 min after the consumption of preload, and CCK levels were measured. A lunchtime meal of hot food was provided from which subjects ate *ad libitum* until satisfied. Energy and nutrient intakes from the food consumed were calculated. There was no significant difference in CCK levels, subjective measures of satiety or food intake between treatments at the given preload level. These results suggest that the protein fractions at the dose employed do not influence satiety, CCK levels or energy intake at a subsequent meal.

Glycomacropeptide: Cholecystokinin: Food intake: Appetite: Satiety

Glycomacropeptide (GMP) is a C-terminal fragment of κ-casein (residues 106–169) released by the endopeptidase chymosin (rennin). During commercial cheese making, this peptide is released into whey at a concentration of approximately $600 \,\mathrm{mg/l^{(1,2)}}$. Oral GMP stimulates cholecystokinin (CCK), a candidate satiety hormone, which may make this protein a useful component of a weight loss diet because CCK slows gastric emptying, which may in turn promote satiety⁽³⁻⁵⁾. GMP has also been detected in the blood of volunteers after milk or yogurt ingestion, suggesting that GMP can be formed in the gut and can be absorbed intact into intestinal cells⁽⁶⁾. We have shown previously that GMP was associated with reduced fat mass in Wistar rats fed ad libitum for 7 weeks with diets differing in protein-type amount⁽⁷⁾. In addition, it has been found that GMP (33 kJ) had no effect on satiety or on food intake 75 min after consumption, but it did reduce daily food intake⁽⁸⁾. However, Gustafson et al. (9) reported no effect of GMP on subjective satiety or food intake at a test meal after the consumption of 0.4 and 2.0 g GMP in a 33 kJ preload; however, CCK was not measured, and various study design procedures, such as delivery with a low energy load, may have influenced the negative findings.

GMP naturally exists as a mixture of different glycoforms due to the carbohydrates sialic acid (*N*-acetylneuraminic acid, galactose, galactosamine and glucosamine attached to the peptide by *O*-glycosidic linkages. There have been no reports showing the effect of these natural GMP glycoforms on CCK, subjective measures of satiety and food intake. The objective of the present study was therefore to examine the effect of different GMP glycoforms on CCK, subjective measures of satiety and food intake.

Methods

GMP fractions from bovine cheese whey were manufactured using simulated moving bed chromatography on a Continuous Separations (Calgon Carbon Corporation, Pittsburgh, PA, USA) platform. This platform consisted of thirty columns of 4-6 cm diameter and 10 cm bed height packed with Pharmacia Q Sepharose Big Beads ion exchange resin (GE Healthcare, Uppsala, Sweden). The columns were equilibrated with reverse-osmosis water (pH 4-0) in the adsorption wash zone, followed by counter-current loading of the whey (pH 4-0; Bega Cheese Co., Bega, NSW, Australia). The columns were then rinsed with reverse-osmosis water at a neutral pH,

and the glycosylated and minimally glycosylated GMP fractions were desorbed from the resin with 0·3 M-NaCl at pH 7·7 and was ultrafiltered using a spiral wound membrane (10 000 Da; Koch Membrane Systems, Inc., Wilmington, MA, USA). The flow through whey was ultrafiltered as described above to produce GMP-depleted whey protein concentrate. All samples were spray dried, and the *N*-acetylneuraminic acid, galactosamine and galactose contents of the glycosylated and minimally glycosylated GMP fractions were determined following a method that was similar to that reported previously for glycoprotein carbohydrates^(10,11).

The minimally glycosylated GMP contained 3.5 (sD 0.1) % N-acetylneuraminic acid and 1.5 (sD 0.1) % galactose, and the glycosylated GMP contained 12.0 (sD 0.3) % N-acetylneuraminic acid and 4.2 (sD 0.2) % galactose.

Subjects and methods

Twenty-two overweight or obese male volunteers (56·3 (SD 8·0) years) were recruited from the Commonwealth Scientific and Industrial Research Organisation (CSIRO) volunteer database.

Volunteers in this database are members of the public who have volunteered for studies at CSIRO and have provided consent to be contacted regarding a variety of studies. The present study was conducted according to the guidelines laid down in the Declaration of Helsinki, and all procedures involving human subjects were approved by the CSIRO Human Nutrition Human Research Ethics Committee. Written informed consent was obtained from all subjects.

Inclusion criteria were male sex, age 20-65 years and BMI >25 kg/m², with no recent history (past 3 months) of weight loss or changes to diet or physical activity routine and an unrestrained eater (i.e. restrained eater questionnaire score > 10)⁽¹²⁾. Exclusion criteria were type 1 or type 2 diabetes or active liver and kidney disease as noted from medical questionnaire, current gastrointestinal disease, past history of gastrointestinal surgery which may affect study outcomes, hypersensitivity to study foods (casein, whey or wheat), medications which affect gastrointestinal motility or hunger/appetite (e.g. metoclopramide, domperidone and cisapride, anticholinergic drugs (e.g. atropine) and erythromycin) or inability to comprehend study protocol. Twenty male volunteers completed the study $(56.9 \text{ (SD } 7.2) \text{ years}, 97.4 \text{ (SD } 8.1) \text{ kg}, BMI 31.5 \text{ (SD } 3.0) \text{ kg/m}^2).$ The volunteers in the study had a sedentary lifestyle estimated by a prestudy questionnaire. Two participants withdrew for personal reasons unrelated to the study.

The study was a randomised double-blind design of four different preload compositions with a 3d interval between study treatments. The preload compositions were controlled for colour, taste and texture, and this was achieved by adding a non-nutritive artificially sweetened chocolate syrup. The nutrient composition of the chocolate syrup (per 70 g) was 95 kJ, 0.4 g protein, 9.7 g carbohydrate and 0.2 g fat. The palatability of the preloads was tested by CSIRO staff before the study. The preloads were served in an opaque container, and were consumed through a straw to limit any effect of appearance or smell on response and palatability. Preload compositions were prepared on the day before the study. The amounts of protein in each of the test products used in the preloads were 41.3 g for the minimally

glycosylated GMP fraction, 42·3 g for the glycosylated GMP fraction and 44·4 g for GMP-depleted whey protein concentrate fraction, and total energy intake of the preload compositions was 895 kJ. Volunteers were asked to consume the preload within 5 min of commencing to drink.

A controlled meal was provided to each participant for consumption on the evening before trial commencement. This provided 3020 kJ, 58 g protein, 25 g fat, 56 g carbohydrate and 21 g fibre. Approximately, 25-30% of the estimated energy requirements were provided by the evening meal. The study participants were asked to abstain from alcohol consumption and to avoid excessive exercise on the evening before visiting the clinic. Volunteers were also asked to record any food eaten on the evening before each clinic visit. A hot lunchtime meal was provided for each participant 3 h after the preload treatment, where they could eat ad libitum until satisfied. Volunteers were given a choice of three options for this lunchtime meal: pasta with bolognaise sauce, veal casserole with rice or chicken curry with rice. In order to control for energy density, volunteers consumed the same meal for all the four study treatments. The amount of food given to each participant, the amount of food consumed and the amount remaining after eating until satisfied were weighed to calculate energy and nutrient intakes. This protocol has been described previously⁽¹³⁾.

Subjects rated their appetite using validated⁽¹⁴⁾ visual analogue scale before the preload and after every blood sample collection. The visual analogue scale was adapted to a sliding scale, computerised format (Northeast Data Corp. Slider ActiveX Custom Control (1.0) Charlotte, NC, USA)⁽¹³⁾. The questions were related to hunger, satisfaction, fullness and prospective food intake. A 100-mm horizontal red line was shown below each question, and opposite extremes of the feeling were described at either end of the line. Subjects moved the cursor along the line using the mouse to indicate how they felt at that moment.

A cannula was inserted into a forearm vein, and a fasting blood sample was collected for each participant. The preload was consumed at approximately 09.00 hours, and subsequent blood samples were collected at 15, 30, 60, 90, 120 and 180 min after the consumption of the preload for the measurement of CCK. The cannula was removed after the final blood sample collection. Biochemical analyses for the satiety hormone CCK were performed after study completion on all blood samples that had been taken for each participant. We have described this protocol previously⁽¹³⁾.

The blood was collected in EDTA (1 g/l) tubes containing aprotinin (5000 KIU (Kallikrein inhibitor units) per 8 ml blood; Mayne Pharma, Melbourne, Australia) and DPP-IV inhibitor (80 μ l per 8 ml blood; Australian Laboratory Services) and stored on ice until processed. The plasma was isolated by centrifugation for 10 min at 2000 g, (5°C; Allegra XR-12 Centrifuge) and stored at -80° C until analysis.

Statistical analysis

Statistical analyses were performed using SPSS 14.0 for WINDOWS (SPSS, Inc., Chicago, IL, USA). Repeated-measures ANOVA was used to assess the effect of time and the effect of treatment on the outcome variables measured, energy intake, CCK and subjective assessment of satiety.

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Table 1. Energy and macronutrient intakes at a hot buffet lunch 3 h after the consumption of preloads containing glycomacropeptide (GMP)-depleted whey protein concentrate (WPC) fraction, glycosylated GMP fraction, minimally glycosylated GMP fraction and glucose*

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(Mean values and standard deviations)

	GMP-depleted WPC fraction		Glycosylated GMP fraction		Minimally glycosy- lated GMP fraction		Glucose	
	Mean	SD	Mean	SD	Mean	SD	Mean	SD
Energy (kJ)	3985	917	3785	1037	3797	1023	3870	999
Protein (g)	63-8	18.3	61.3	19⋅6	61.0	16.6	63.4	18.7
Fat (g)	21.5	6.9	20.6	7⋅1	20.6	6.6	21.5	6.8
Carbohydrate (g)	119-8	26-6	112.5	31.4	113-6	35.7	113.4	30.5

^{*} Data were analysed using repeated-measures ANOVA.

Statistical significance was set at $P \le 0.05$. All data are presented as mean values and standard deviations.

Results

Food intake

Energy intake at the hot lunchtime meal was not different between treatments (P=0·72): 3985 (sD 917)kJ GMP-depleted whey protein concentrate fraction, 3785 (sD 1037)kJ glycosylated GMP fraction, 3797 (sD 1023)kJ minimally glycosylated GMP fraction and 3870 (sD 999)kJ glucose. In addition, there were no differences between treatments for protein (P=0·71), fat (P=0·70) or carbohydrate (P=0·59) intake at the hot lunchtime meal (Table 1).

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There were no significant differences in CCK concentrations between the preload treatments (P=0.45; Fig. 1).

Visual analogue scale

There was a time-by-treatment effect (P=0.043) for the visual analogue scale question 'How satisfied do you feel?'. There were no differences in the visual analogue scale questions 'How hungry do you feel?' (P=1.0), 'How full do you feel?' (P=0.952), 'How much do you think you can eat?' (P=0.531), 'Would you like to eat something sweet?' (P=0.239), 'Would you like to eat something salty?' (P=0.4), 'Would you like to eat something savoury?' (P=0.295) and 'Would you like to eat something fatty?' (P=0.728).

Discussion

The main findings of the present study were that none of the naturally occurring GMP glycoforms had an effect on CCK concentrations, subjective measures of satiety, and energy or macronutrient intake when compared with glucose.

These findings are in contrast to previous studies in which a glucose-containing preload compared with preloads containing a similar amount of protein from whey or casein was consumed^(15,16). In a study by Bowen *et al.* ⁽¹⁵⁾, nineteen overweight men consumed liquid preloads containing 52 g protein from whey or casein and 56 g lactose or glucose. In contrast to the present study, energy intake was 10% higher

after the consumption of glucose preload compared with lactose and protein preloads (P < 0.05). Also in contrast to the present study, CCK was 71% higher 90 min after the consumption of protein preloads compared with glucose and lactose (P < 0.05). In a second study by Bowen et al. (16), liquid preloads containing 50 g protein from whey, soya and gluten, or 63 g glucose and 1200 kJ were consumed. Once again in contrast to the present study, energy intake was 10% lower after the consumption of all protein preloads compared with glucose (P < 0.05), and CCK concentrations were elevated after the consumption of protein preloads. It is not clear why differences between the protein and glucose preloads were not observed in the present study as the participants were similar in age and BMI. One reason may be that the protein content of the preloads was 10 g less and the energy was 170 J less in the present study, and differences may have been observed if the lunchtime meal had been offered earlier. The inter-meal interval of 3 h may have been too long to detect subtle differences. Evidence to support this is provided by Bowen et al. (16), in which twenty-eight obese men consumed preloads containing 50 g whey, fructose, glucose or 25 g whey + 25 g fructose and 1100 kJ. While CCK concentrations were increased after the consumption of protein preload, energy intake was not different between preloads. The inter-meal interval was 4h in the present study, suggesting that this may have been too long to detect small

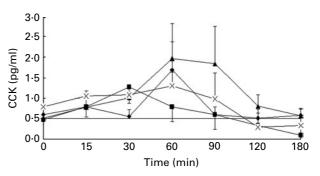


Fig. 1. The effect of four preloads containing either glycomacropeptide (GMP)-depleted whey protein concentrate (WPC) fraction, minimally glycosylated GMP fraction, glycosylated GMP fraction or glucose on plasma cholecystokinin (CCK) levels. Data are mean values with their standard errors. Data were analysed using repeated measures ANOVA. There was no time by treatment interaction (P=0.45). $- \spadesuit -$, GMP-depleted WPC fraction; $- \blacksquare -$, glycosylated GMP fraction; $- \blacktriangle -$, minimally glycosylated GMP fraction; $- \star -$, glucose.

differences in energy intake. Veldhorst *et al.* found that energy intake at a lunchtime meal 3 h later was lower after a breakfast meal containing whey than after a breakfast containing whey without GMP, suggesting that GMP may have been responsible for the effect⁽¹⁷⁾. However, the energy content of the test breakfast used was 2520 (SD 70)kJ much greater than that in the present study, in which it was 895kJ. It may be that hunger from the lower energy load may have overwhelmed any subtle effect of either protein or GMP. The volunteers were also young and lean in the Veldhorst studies. Energy and nutrient intakes from the controlled meal the evening before in the present study may have been less than usual, further contributing to hunger. However, as we did not collect food records before the study, we do not know whether usual food intake was greater than that provided.

In a separate study, Veldhorst *et al.* ⁽¹⁸⁾ found that whey was more satiating than either soya or casein only at the 10% of energy level and not at the 25% of energy level. But there were no differences in energy intake at either protein level. In a later study, Veldhorst *et al.* ⁽¹⁹⁾ found that food intake at lunch was 20% lower 180 min after a breakfast containing α -lactalbumin, gelatin or gelatin plus tryptophan compared with breakfast containing casein, soya, whey and whey with GMP, which were not different from each other.

Similar to the present results, Lam et al. (20) observed that energy and macronutrient intakes were not different following preloads given 30 min before lunch containing maltodextrin, whey protein isolate with no GMP, whey protein isolate with 21 % naturally present GMP or whey protein isolate with 21 % naturally present GMP plus 20 g added GMP. The energy content of the preloads, 1300 kJ, was greater than that in the present study with similar total protein (43-46g) in the whey-containing preloads. Burton-Freeman⁽⁸⁾ also found that while satiety scores were greater after the consumption of protein preloads, energy and macronutrient intakes at a test lunch, 1 h after the consumption of preload, were not different between treatments. Preloads contained 1000 kJ energy and whey protein isolate, whey protein without GMP, GMP isolate (0.8 g) or a high carbohydrate control⁽⁸⁾. Thus, the majority of studies have not shown that whey with GMP or isolated GMP is superior to whey without GMP or indeed any other protein source. The level of GMP fed in the present study was also higher than that fed in any other study, positive or negative. Inconsistent effects of different proteins and peptides have been shown in other studies. Diepvens et al. (21) observed lower hunger, desire to eat and thirst after consumption of pea compared with milk protein or combined pea and whey proteins⁽²¹⁾. Pea and whey protein separately led to greater satiety and fullness compared with milk protein or the combined pea and whey proteins in overweight subjects given 1024 kJ and 15 g protein. However, despite these differences in subjective measures, there was no effect on energy intake.

It has been postulated that that CCK responses may be abnormal in obesity. Zwirska-Korczala *et al.* ⁽²²⁾ observed that fasting CCK concentrations were lower in morbidly obese individuals compared with lean controls, and that the postprandial CCK response was reduced in morbidly obese individuals ⁽²²⁾. However, in a study from our group by Bowen *et al.* ⁽¹⁶⁾, there was no difference between lean and obese subjects in fasting CCK concentrations. Additionally, there was no effect of body weight on postprandial

CCK. The effect of body weight status on postprandial CCK responses remains to be clarified.

Conclusions

These results suggest that these protein fractions in the dose used were not effective in reducing food intake at a subsequent meal, and that they did not affect postprandial CCK concentrations. The extent of GMP glycosylation had no influence on the results.

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