



The Nutrition Society Irish Section Conference 2021 was held virtually on 22–24 June 2021

Conference on Nutrition, health and ageing – translating science into practice Symposium two: Anthropometric, lifestyle and dietary concerns in ageing – diagnostic and interventional considerations

Fish, *n*-3 fatty acids, cognition and dementia risk: not just a fishy tale

Rasha N. M. Saleh¹ and Anne Marie Minihaue^{1,2*}

¹Nutrition and Preventive Medicine, Norwich Medical School, BCRC, Rosalind Franklin Road, University of East Anglia (UEA), Norwich NR4 7UQ, UK

²Norwich Institute of Healthy Ageing, UEA, Norwich NR4 7UQ, UK

With growing and ageing populations, the incidence of dementia is expected to triple globally by 2050. In the absence of effective drugs to treat or reverse the syndrome, dietary approaches which prevent or delay disease onset have considerable population health potential. Prospective epidemiological studies and mechanistic insight from experimental models strongly support a positive effect of a high fish and long chain *n*-3 fatty acid (EPA and DHA) intake on a range of cognitive outcomes and dementia risk, with effect sizes equivalent to several years of ageing between the highest and lowest consumers. As reviewed here, an effect of EPA and DHA on neuroinflammation and oxylipin production is likely to in part mediate the neurophysiological benefits. However, randomised controlled trials (RCTs) with EPA and DHA supplementation have produced mixed findings. Insight into the likely modulators of response to intervention and factors which should be considered for future RCTs are given. Furthermore, the impact of *APOE* genotype on disease risk and response to EPA and DHA supplementation is summarised. The prevalence of dementia is several-fold higher in *APOE4* females (about 13% Caucasian populations) relative to the general population, who are emerging as a subgroup who may particularly benefit from DHA intervention, prior to the development of significant pathology.

EPA: DHA: Alzheimer's disease: Neuroinflammation and oxylipins

Cognition refers to the mental process of acquiring knowledge and processing information. It includes functions such as attention, memory, problem solving, decision making, planning, inhibition, judgement and evaluation, reasoning, comprehension and production of language and orientation/visuospatial skills. Dementia is a general term for a loss of one or more of these functions that is severe enough to interfere with daily life. There are now over 100 recognised forms of dementia, with Alzheimer's disease (AD) being the most prevalent, and responsible for about two-thirds of dementia cases.

Globally, there are 50 million living with dementia⁽¹⁾, occurring in 5–8% of those aged over 60 years, with the prevalence increasing exponentially with age^(2, 3) (Fig. 1).

With growing and ageing populations, and more widespread diagnostic services, diagnosed dementia rates are predicted to triple by 2050⁽¹⁾. Dementia is the second leading cause of death globally after IHD⁽²⁾ and in England and Wales dementia is now the single greatest cause of death in women, responsible for 16.5% of total mortality (v. 8.7% in men)⁽⁴⁾. This sex difference

Abbreviations: AD, Alzheimer's disease; BBB, blood–brain barrier; HR, hazard ratio; LC *n*-3 PUFA, long chain *n*-3 fatty acid; LPC, lysophosphatidylcholine; RCT, randomised controlled trial; RR, relative risk; SPM, pro-resolving mediator.

*Corresponding author: Anne Marie Minihaue, email a.minihane@uea.ac.uk

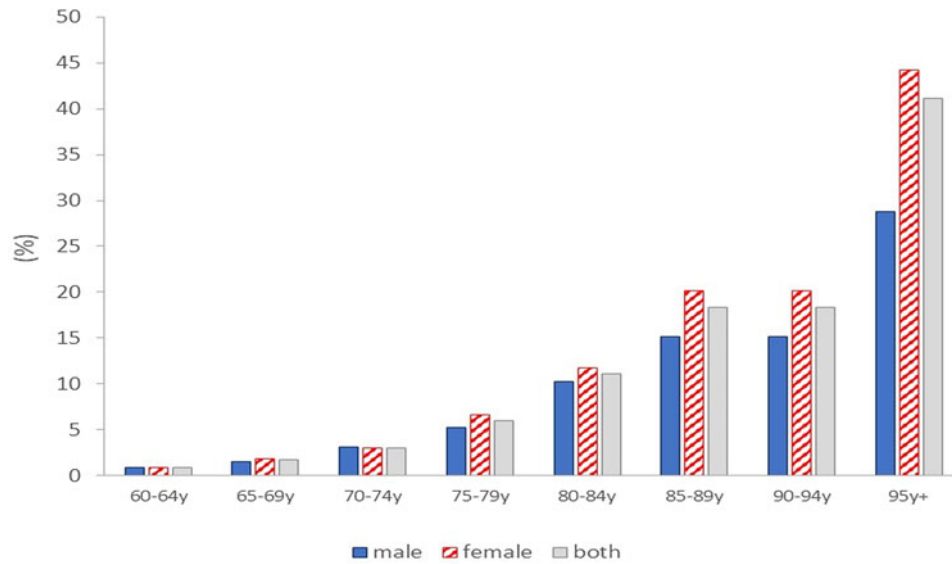


Fig. 1. Dementia prevalence (% of the population) by age group in the UK⁽³⁾.

in dementia-associated death rates is reflective of the fact that two-thirds of dementia patients are females⁽²⁾, the physiological and molecular basis of which is only partially elucidated⁽⁵⁻⁷⁾. Accelerated neuropathology and cognitive decline, evident during the menopausal transition in females, and the higher penetrance of the at-risk *APOE4* allele in female carriers are likely to be major contributing factors⁽⁸⁻¹¹⁾.

However, encouragingly age-standardised rates are decreasing in many high-income countries. Between 1990 and 2016, a 6.8, 10.3 and 8.4% reduction in dementia-associated death, prevalence and disability-adjusted life years rates, respectively, was observed in the UK⁽²⁾. These reductions have been attributed to greater education attainment (creating cognitive reserve), better cardiovascular health and improved nutrition.

Overview of interventions for dementia treatment and prevention

There are currently few effective drugs to prevent or treat dementias. In the UK, there are four licensed drugs available (donepezil, rivastigmine, galantamine and memantine) which temporarily treat symptoms by targeting synaptic function and neurotransmission. In 2021, the Food and Drug Administration granted accelerated approval to aducanumab, the first drug in 18 years for AD⁽¹²⁾. It is a monoclonal antibody which targets amyloid clearance and is currently undergoing regulatory review in Europe. Its purported efficacy is controversial with the benefits thought to be marginal in most patients⁽¹²⁾.

In the absence of effective pharmaceutical options to prevent, reverse or treat dementia, there is a widespread interest in lifestyle behaviour approaches including nutrition to prevent or delay neurophysiological and cognitive decline. In the 2020 *Dementia prevention, intervention,*

and care: 2020 report of the Lancet Commission, it was estimated 'that 12 modifiable risk factors account for around 40% of worldwide dementias, which consequently could theoretically be prevented or delayed'⁽¹³⁾. Many of these are nutrition-dependent (hypertension, obesity, diabetes, depression and recovery from traumatic brain injury) and likely mediate the emerging role of nutrition in brain health.

Research into the role of nutrition in age-related cognitive decline is in its relative infancy compared with other chronic conditions such as CVD and osteoporosis, with research evidence largely derived from prospective cohort studies or experimental models.

Although not fully consistent, a growing body of prospective cohort evidence shows that plant-based dietary patterns and individual dietary bioactives such as selenium, vitamin D, B-vitamins, polyphenols and long chain *n*-3 fatty acids (LC *n*-3 PUFA) improve cognition and reduce dementia risk, conversion of mild cognitive impairment to AD and brain atrophy⁽¹⁴⁻²⁰⁾. A Mediterranean dietary pattern and the Mediterranean-Dietary Approaches to Stop Hypertension Diet Intervention for Neurodegenerative Delay diet have emerged as particularly effective with high *v.* low adherence associated with up to a 40% reduced dementia rate^(17,21-23). The potential of the protective role of a Mediterranean dietary pattern was highlighted in the 2017 *Lancet Commission Dementia prevention, intervention, and care report*⁽²⁴⁾, with a Mediterranean dietary pattern being the only specific dietary approach for which the WHO 2019 *Risk Reduction of Cognitive Decline and Dementia* guidelines⁽²⁵⁾ recommended to adults with normal cognition to reduce the risk of cognitive decline and dementia. A defining component of a Mediterranean dietary pattern is a high fish and *n*-3 fatty acid intake. Here, we focus on the role of the LC *n*-3 PUFA DHA in brain function and on evidence for a protective role of higher fish and DHA and EPA intake and status in cognitive health. We critique the apparent

inconsistencies between the protective associations observed in prospective cohort and biological effects in experimental models v. the inconsistent and often null or marginal effects seen in randomised controlled trials (RCTs).

Fatty acid uptake into the brain

DHA is a 22-carbon *n*-3 fatty acid, with multifaceted structural and functional roles in the central nervous system. Although DHA can be synthesised in the liver from the plant precursor *n*-3 fatty acid α -linolenic acid, bioconversion is less than 0.2%⁽²⁶⁾. Within the brain, the synthesis of DHA from α -linolenic acid is negligible. Therefore, DHA uptake via the highly selective blood-brain barrier (BBB) is required to replace the DHA consumed in metabolic reactions. A dietary supply of DHA as either oily fish or supplements is recommended to meaningfully enrich brain levels. DHA is the predominant PUFA in the brain, accounting for 15% of total fatty acids which is several fold higher than most other tissues such as the heart and the liver where it constitutes about 2%⁽²⁷⁾. Grey matter, including synaptic membranes, synaptic vesicles and mitochondria, are particularly enriched⁽²⁸⁾.

The BBB is formed of tightly connected endothelial cells, embedded within a network of pericytes and astrocytes foot processes that support its function⁽²⁹⁾. Fatty acids cross the BBB by two known mechanisms, either facilitated transport by several transmembrane proteins or by passive diffusion⁽³⁰⁾. BBB uptake of plasma fatty acids was historically thought to be only from NEFAs, which originate from lipoproteins or are bound to plasma proteins, mainly albumin⁽³¹⁾. More than 99% of non-esterified-DHA is protein bound. NEFAs are transported through the endothelial cell membranes and cytoplasm via a group of fatty acid transport proteins (FATP) and fatty acid binding proteins. FATP1 and FATP4 are highly expressed in both the vascular and the parenchymal regions of the brain⁽³²⁾. Recent studies show that FATP1 participates in 60% of DHA uptake⁽³³⁾. Interestingly, in a cell culture model, amyloid- β (A β), the hallmark of AD pathology, induced a 96% reduction in FATP1 protein expression and an associated 45% reduction in DHA efflux⁽³⁴⁾. More recently, acyl-CoA synthetase 6 (*Acs6*) has been identified as an essential transporter for enriching the brain with DHA^(35,36).

Besides the NEFA form, DHA is also taken up into the brain in the form of lysophosphatidylcholine (LPC-DHA)⁽³⁷⁾. The major facilitator superfamily domain-containing protein 2a (*MFSD2A*) is considered the major route of LPC-DHA uptake^(38–40). Indeed, *Mfsd2a* knockout mice showed 50% lower brain DHA levels compared to wild-type animals, with consequent cognitive deficits, anxiety and microcephaly⁽³⁸⁾. Along with its emerging role in LPC fatty acid transport and the regulation of BBB permeability⁽⁴¹⁾, *MFSD2A* is emerging as having more far-reaching functions in neuroinflammation and other physiological and pathophysiological brain processes⁽⁴²⁾. Overall, although the

uptake and partitioning coefficient is higher for LPC-DHA relative to non-esterified-DHA following intravenous injections, non-esterified-DHA is thought to be the main source of DHA for the brain due to its higher circulating concentrations^(31,43).

Although currently largely unknown, it is emerging that variables such as age, menopause, neuropathology and *APOE* genotype status⁽⁴⁴⁾, may impact the brain DHA uptake processes, and has implications for the recommended DHA dose in population subgroups, and the optimal intervention ‘window of opportunity’ when DHA supplementation is most likely to bring about cognitive benefits. Furthermore, defective brain DHA uptake could underpin the lack of benefit of DHA observed in several RCTs, particularly in *APOE4* carriers (see section ‘Fish and DHA intake and status and cognition: randomised controlled trial evidence’).

APOE genotype: impact on dementia/Alzheimer’s disease risk and age of onset

ApoE, produced mainly in the brain by glial cells, is the principal lipid transporter within the brain and cerebrospinal fluid, but also has numerous other roles in neuroinflammation and neuronal function. Two missense mutations in *APOE* gene (rs429358 and rs7412) produce three allele variants: ϵ 2, ϵ 3 and ϵ 4. These alleles have different amino acids (cysteine or arginine) in positions 112 and 158, resulting in ApoE2 (Cys112 and Cys158), ApoE3 (Cys112 and Arg158) and ApoE4 (Arg112 and Arg158)⁽⁴⁵⁾. These amino acid differences lead to conformational changes in ApoE structure which affects binding to lipoprotein receptors and also the stability and tissue concentrations of the protein⁽⁴⁶⁾. The global frequencies of the ϵ 2, ϵ 3 and ϵ 4 alleles are approximately 8.4, 77.9 and 13.7%, respectively⁽⁴⁷⁾. *APOE* genotype is the most important common genetic determinant of cognitive decline and AD risk, with a 3-fold increased prevalence of the ϵ 4 allele in AD v. the general population and the *APOE3/E4* and *APOE4/E4* genotypes having a 2–3- and 12–15-fold increased risk of AD compared to the wild-type *APOE3/E3* genotype^(48,49). In addition, *APOE4* is associated with an average lower age of AD onset⁽⁴⁹⁾. It falls from 84 years in *APOE4* non-carriers to 76 years in *APOE3/E4* to 68 years in *APOE4/E4*^(49,50). The aetiology of the increased risk in *APOE4* carriers is multi-faceted and can be attributed to defective A β clearance, a loss of neuronal synaptic plasticity and dendrite outgrowth, neuroinflammation, cerebrovascular and BBB dysfunction and lower brain DHA status⁽⁵¹⁾.

In a transgenic rodent model, the uptake of [¹⁴C]-DHA using *in situ* cerebral perfusion was significantly lower in *APOE4* v. *APOE2* animals, which was exacerbated by age⁽⁵²⁾. This observation of a greater effect of age on brain DHA is consistent with our more recent rodent studies, where the effect of age was more evident in females⁽⁵³⁾ and following induction of menopause⁽¹⁰⁾. In human subjects, DHA supplementation resulted in lower circulating DHA levels^(54,55), higher systemic β -oxidation⁽⁵⁵⁾ and lower cerebrospinal fluid DHA

following 18 months of supplementation⁽⁵⁶⁾. Defective BBB transfer, brain lipid transport and increased oxidation of DHA following upregulated release by PhospholipaseA2, are all likely contributors to a lower DHA brain status in *APOE4*⁽⁴⁴⁾. As will be discussed later, *APOE* genotype has also emerged as an important mediator of the effect of DHA status and intervention on incident dementia and cognitive outcomes, but the effect is inconsistent and likely to be dependent on age, sex and brain health stage.

The role of DHA in the brain

Since first being identified in the brain by Klenk and Bongard in 1952⁽⁵⁷⁾, many neurophysiological roles have been identified for DHA in experimental models, including membrane structural roles (fluidity and modulation of membrane protein function) and the modulation of neurogenesis and neuronal cell growth and cell survival, A β clearance, vascular function and brain perfusion, BBB permeability, oxidative status, neuroinflammation, synaptic function and neurotransmission^(9,31,58–66). Synaptosomal membranes are particularly enriched in DHA, where it constitutes up to 40% of PUFA in select lipid species and modulates neurotransmitter levels and membrane dynamics⁽⁶⁷⁾. Loss of synaptic plasticity is a major contributor to the pathogenesis of cognitive decline, mediated in part through reduced levels of brain-derived neurotrophic factor and its related signalling pathways⁽⁶⁸⁾. DHA is known to increase the level of brain-derived neurotrophic factor and consequently activates protein kinase B (Akt) and extracellular signal-regulated kinase signalling pathways leading to improved synaptic plasticity⁽⁶⁹⁾. Reduced recognition memory was evident in menopausal *APOE4* mice models fed with a high-fat diet⁽¹⁰⁾. This memory deficit was associated with a 13% reduction in cortical DHA, reduced brain-derived neurotrophic factor expression and compromised Akt, mammalian target of rapamycin and extracellular signal-regulated kinase signalling pathways, highlighting the mechanistic role of DHA, interacting with menopause and *APOE4*, in cognitive decline via modulation of synaptic plasticity-related pathways⁽¹⁰⁾.

A systematic review on the effects of relatively long-term *n*-3 intervention in animal AD models included data from 15 studies and reported significant reductions in amyloid levels, plaque burden and neuronal loss and improved cognition following DHA only or EPA + DHA supplementation⁽⁷⁰⁾.

Once released from membrane phospholipids via phospholipase A2, DHA regulates inflammation through the modulation of cytokine production and as a precursor for a host of bioactive oxylipins^(30,71–75) (see the next section).

Neuroinflammation, oxylipins and brain health

Amyloid plaque deposition is one of the hall marks of AD pathology. Risk factors such as *APOE4* carrier status, vascular pathologies and neuroinflammation play

interactive roles in the cascade of synthesis of A β and the progression of cognitive decline⁽⁷⁶⁾. Indeed, pro-inflammatory cytokines such as IL6 and TNF α are increased in the blood and brain of patients with AD^(77,78). Brain microglia, the brain-resident immune cells, are the major regulator of brain inflammatory status via the release of inflammatory cytokines such as IL1 β , TNF α and inducible nitric oxide synthase⁽⁷⁹⁾. Activated microglia surround amyloid plaques in the cerebral cortex of AD patients, which suggests that A β deposition can trigger microglial activation and subsequent release of inflammatory cytokines^(80,81).

However, recent studies suggest that neuroinflammation also plays an A β -independent role in the pathogenesis of cognitive decline^(82,83). Imaging studies have observed microglial activation in patients with mild cognitive impairment even before the appearance of amyloid deposits^(84,85) which increases with disease progression^(86,87).

PUFAs have been extensively studied as a modulator of systemic inflammation in chronic diseases such as atherosclerosis, diabetes and rheumatoid arthritis. These conditions are consistently associated with higher C-reactive proteins (CRPs), TNFs, IL6, thromboxane A2 (TXA2) and leucotrienes B4 (LTB4)^(88,89), which are affected by tissue PUFA status⁽⁹⁰⁾ (Fig. 2). Similar to its systemic anti-inflammatory role, *n*-3 PUFAs are considered effective modulators of the brain inflammatory status^(91,92). Higher DHA intake was associated with lower IL6 in the mouse hippocampus⁽⁹³⁾ and inhibition of the NF- κ B inflammatory pathway⁽⁹⁴⁾. DHA reduced A β deposition in an AD mouse model⁽⁶²⁾ through the reduction of the IL12/IL23 signalling pathway⁽⁹⁵⁾.

The biological actions of PUFAs in controlling neuroinflammation are in part mediated through their enzymatically and non-enzymatically oxidised metabolites, called oxylipins⁽⁹⁶⁾ (Fig. 2). These lipid-derived oxygenated metabolites of PUFA are synthesised by three groups of enzymes: cyclooxygenases, lipoxygenases and cytochrome P450 enzymes⁽⁹⁷⁾, which produce hydroxy-, dihydroxy- or epoxy-fatty acids. Due to their highly unsaturated nature, PUFAs are also non-enzymatically oxidised (i.e. autooxidation) by reactive oxygen and nitrogen species⁽⁹⁸⁾. Oxylipins are precursors for specialised pro-resolving mediators (SPMs; resolvins, protectins, maresins and lipoxins) which have anti-inflammatory and pro-resolving roles⁽⁹⁹⁾. *n*-6 PUFA-derived oxylipins are generally pro-inflammatory relative to EPA/DHA-derived species. The enzymatic action of cyclooxygenases on AA produces the pro-inflammatory PGH2, TXA2 and 5-, 12- and 15-hydroxy-eicosatetraenoic acid (HETEs)⁽¹⁰⁰⁾. It is worth mentioning that AA is also a precursor of the anti-inflammatory and pro-resolving lipoxins A4 and B4⁽⁹⁶⁾.

n-3 PUFA-derived oxylipins generally have anti-inflammatory and pro-resolving properties. EPA produces less inflammatory PGs, TXAs (3-series) and LTs (5-series)⁽¹⁰¹⁾ relative to AA-derived oxylipins. Epoxy-EPA oxylipins produce anti-inflammatory responses⁽¹⁰²⁾ partly through the inhibition of NF- κ B pathway and through antagonising inflammation induced by PGE₂^(103,104). RvE1 reduces proinflammatory

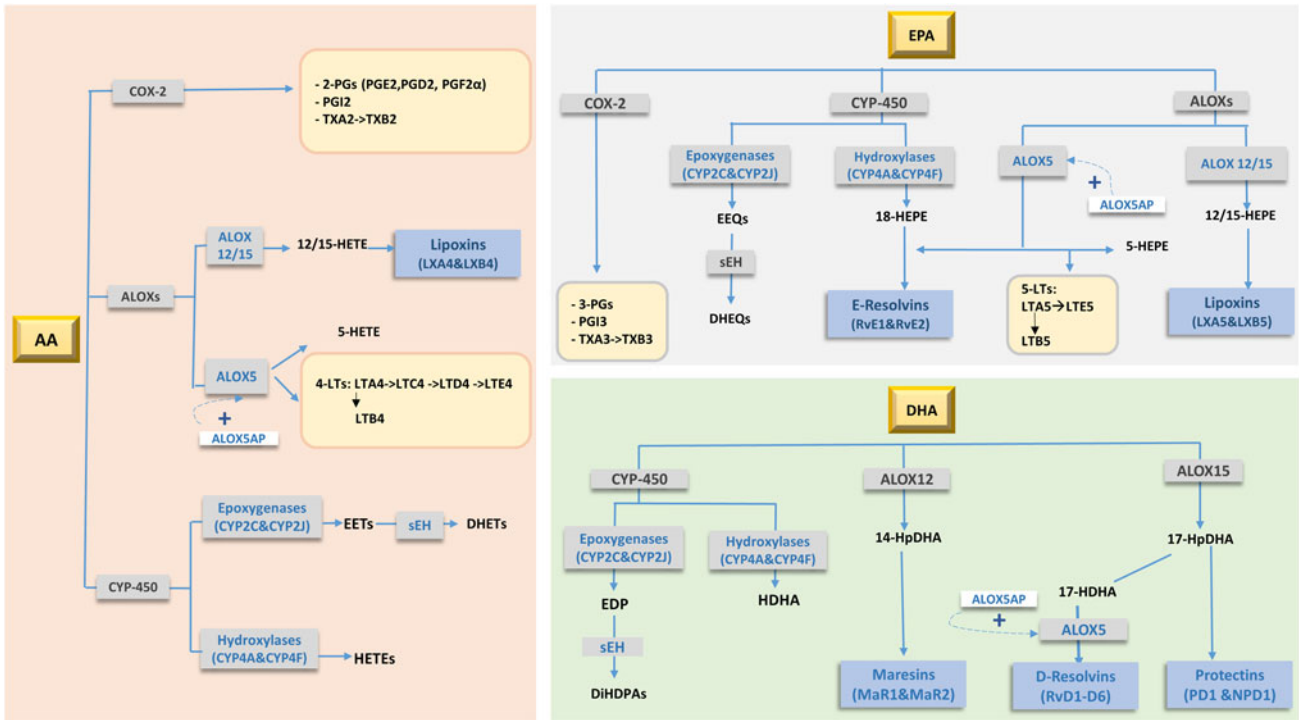


Fig. 2. Main oxidative products (oxylipins) of arachidonic acid (AA), EPA and DHA metabolism. Adapted from Schulze *et al.*⁽⁹⁷⁾. 2-PGs, 2 series prostaglandins; 3-PGs, 3 series prostaglandins; 4-LTs, 4 series leucotrienes; 5-LTs, 5 series leucotrienes; ALOXs, arachidonate lipoxygenases; ALOX5AP, 5-lipoxygenase activating protein; COX-2, cyclooxygenase-2; CYP-450, cytochrome-P450; DHEQs, dihydroxyeicosatetraenoic acids; DHET, dihydroxyeicosatrienoic acid; DiHDPA, dihydroxydocosapentaenoic acid; EDP, epoxydocosapentaenoic acid; EEQ, epoxyeicosatetraenoic acid; EETs, epoxyeicosatrienoic acid; HDHA, hydroxydocosahexaenoic acid; HEPE, hydroxyeicosapentaenoic acid; HETE, hydroxy-eicosatetraenoic acid; HpDHA, hydroperoxide intermediate of DHA; sEH, serum epoxide hydrolase enzyme; TX, thromboxanes.

cytokine production⁽¹⁰⁵⁾, neutrophil infiltration and reduces proinflammatory gene expression in peripheral blood mononuclear cells and microglia through binding to ChemR23 receptor^(106,107).

The role of oxylipins and SPMs in the protection against neuroinflammation and the development of AD is gaining research attention^(73,76,108). The resolution of the inflammatory process is disrupted with ageing and cognitive decline⁽¹⁰⁹⁾. In murine models, increased *n*-6 PUFA-derived oxylipins and decreased *n*-3-derived oxylipins and SPMs are generally observed in neuroinflammatory brain disorders⁽⁷⁶⁾. A reduction in cortical 14- and 17-hydroxydocosahexaenoic acid and hippocampal neuroprotectin D1 with age was observed⁽⁵³⁾. Brain LXA4 was also found to decline with age, with the reduction more pronounced in an AD-mouse model⁽¹¹⁰⁾. Interestingly, administration of LXA4 in this AD-mouse model⁽¹¹⁰⁾ and RvD1 in a post-operative cognitive impairment model⁽¹¹¹⁾ reduced cognitive decline under both conditions. Protectin D1 was first detected in murine blood and neuroprotectin D1 is present in the brain⁽¹¹²⁾. Neuroprotectin D1 levels greatly increased in the hippocampus after lipopolysaccharide stimulation⁽⁷⁴⁾. It binds to the GPR37 receptor⁽¹¹³⁾ to inhibit NF- κ B and pro-inflammatory gene expression⁽¹¹⁴⁾. Neuroprotectin D1 showed protective function in neurodegeneration through modulating synaptic plasticity and

microglial activity⁽¹¹⁵⁾. In human studies, LXA4 was lower in patients with AD compared to mild cognitive impairment or subjective cognitive impairment patients. Similarly, LXA4 and MaR1 were reduced in postmortem hippocampi of AD patients compared to controls, whereas the *n*-6 PUFA oxylipins 5-HETE, 15-HETE, TXB2 and PGs increased⁽¹¹⁶⁾.

Being precursors to oxylipins and SPMs, several studies have explored the potential benefits of *n*-3 PUFA in preventing cognitive decline via modulating the levels of brain oxylipins and SPMs. In aged rats, EPA and DHA supplementation increased cortical 5-HEPE, 7-, 10-, and 17-hydroxydocosahexaenoic acid, PD1, RvD1 and RvD2⁽¹¹⁷⁾. AA-derived PGE₂, PGD₂ and PGF_{2 α} significantly decreased with an associated improvement in reference memory. In response to lipopolysaccharide stimulation, *n*-3 PUFA supplemented mice showed an increase in hippocampal *n*-3 oxylipins compared to non-supplemented mice who showed an increase in the *n*-6 pro-inflammatory oxylipins⁽¹¹⁸⁾. In AD patients, EPA and DHA supplementation increased peripheral blood mononuclear cell RvD1 levels compared to controls⁽¹¹⁶⁾.

Significant inter-individual variability in the response of oxylipins to *n*-3 PUFA supplementation has been reported⁽¹¹⁹⁾. In mice, select EPA- and DHA-derived brain oxylipins and SPMs were lower in *APOE4* compared to *APOE3*⁽⁵³⁾. In addition, the plasma oxylipins

response to EPA + DHA supplementation was influenced by *APOE* genotype in healthy individuals with a greater production of a number of EPA- and DHA-derived species in *APOE4* carriers⁽¹²⁰⁾. Genetic variation in enzymes involved in PUFA metabolism has been implicated as a possible modulator of oxylipin production from PUFAs. Genetic variation in *LTA₄H*, an enzyme in the pathway of leucotriene synthesis, significantly interacted with dietary *n*-3 and *n*-6 fatty acid intake to determine intima-media thickness⁽¹²¹⁾. Variants in *ALOX5* gene were associated with a differential oxylipin response to fish oil supplementation in healthy African American adults⁽¹²²⁾.

Given the central role of neuroinflammation in cognitive decline, the modulation of cytokine, oxylipin and SPM production is a tractable target to prevent and delay neuropathology by increasing EPA and DHA status and intake.

Fish and DHA intake and status and cognition: prospective cohort evidence

There is a substantial and a relatively consistent body of research from prospective cohorts, of an inverse association between fish and EPA and DHA intake and status (measured in a number of blood lipid fractions), and dementia and AD risk, brain atrophy and cognitive decline. In the earliest report from the Rotterdam Cohort study (*n* 5386), with an incident case rate of 1.1% (*n* 58) over 2.1 years, total fat, saturated fat and fish intake were inversely related to incident dementia⁽¹²³⁾. However, in the 9.6 years follow-up of this cohort, with 465 dementia cases, total fish, EPA or DHA intake was not associated with either total dementia or AD risk⁽¹²⁴⁾. This lack of association is in contrast to the findings of the largest prospective analysis conducted to date on fish, *n*-3 fatty acids and dementia, namely the NIH-AARP study in 421 309 adults followed up for 16 years, with 85 112 deaths⁽¹²⁵⁾. Quintile 5 (Q5) *v.* Q1 of total fish intake was associated with a hazard ratio (HR) of AD death of 0.76 (95% CI: 0.61, 0.95) with an even stronger association evident when fried fish was removed. Considering LC *n*-3 PUFA intake, an HR of AD death of 0.70 (95% CI: 0.54, 0.89) was observed in Q5 *v.* Q1 in males, with an even greater benefit in females (HR: 0.59 (95% CI: 0.43, 0.80))⁽¹²⁵⁾. Q5 represented a mean intake of >180 mg and 160 mg daily in males and females of LC *n*-3 PUFAs, mainly EPA + DHA. This intake is modest compared to the typical UK and global recommended intakes of 450–500 mg daily EPA + DHA minimum recommended intake^(126,127), which is mainly targeted towards cardiovascular health.

In an analysis of post-mortem brains, seafood consumption (≥ 1 meal(s)/week), measured on average 4.5 years before death was correlated with less AD pathology including lower neuritic plaques, less severe and widespread neurofibrillary tangles, and lower neuropathologically defined AD but only among *APOE* $\epsilon 4$ carriers⁽¹²⁸⁾.

A number of analyses have reported positive associations between DHA or EPA + DHA status in blood

lipids fractions and cognitive outcomes^(129–131). In the Framingham Cohort, high *v.* low (Q4 *v.* Q1) phosphatidylcholine-DHA was associated with a 47% reduction in all cause dementia⁽¹²⁹⁾. In the Women's Health Initiative Memory Study, the HR of probable dementia in the 9.8 years follow-up was 0.92 (95%: 0.84, 1.00) per SD of erythrocyte EPA + DHA (*n*-3 index) with a similar HR when EPA and DHA were considered separately⁽¹³⁰⁾. The 15-year cumulative incidence of probable dementia was estimated to be 12.1% with high EPA + DHA exposure compared to 14.2% with low EPA + DHA exposure (absolute risk difference = 2.05%).

Ageing and dementia progression are underpinned by total brain atrophy (loss of volume) and in AD the hippocampus is particularly affected. In the Women's Health Initiative Memory study, a 1 SD greater erythrocyte EPA + DHA level was correlated with 2.1 cm³ larger brain volume and greater hippocampal volume (50 mm³), with the effect size purported to be equivalent to 1–2 years of ageing⁽¹⁶⁾. An association between erythrocyte EPA + DHA and medial temporal lobe volume trajectories assessed over a maximum of 10.8 years (median follow-up 4.0 years) was observed in the Three-City study, along with improved global cognition and memory and a 60% increased risk of dementia in Q1 *v.* Q5 of EPA + DHA status⁽¹³²⁾.

The findings from prospective cohort studies have been synthesised into four meta-analyses which focus on fish intake^(133,134), or both fish and LC *n*-3 PUFA intake^(135,136) on a variety of cognitive outcomes, which are further summarised in an umbrella review of meta-analyses⁽¹⁵⁾. Samieri *et al.* pooled the French Three-City study and four US cohorts and included data from *n* 23 688 (88% female) with median follow-ups of 3.9–9.1 years⁽¹³⁴⁾. Higher fish intake was associated with slower decline in both global cognition and episodic memory. The effect of consuming ≥ 4 servings/week *v.* <1 serving/week of fish on episodic memory decline was estimated to be equivalent to 4 years of ageing. Although the Bakre *et al.*'s (*n* 9 studies) analysis does not provide information on actual fish portion consumption per category, a dose-dependent effect was observed with a relative risk (RR) (95% CI) of dementia of 0.84 (0.72, 0.98), 0.78 (0.68, 0.90) and 0.77 (0.61, 0.98) in those with low, middle and high consumption of fish *v.* those with no or lowest consumption of fish, with the corresponding RRs of 0.88 (0.74, 1.04), 0.79 (0.65, 0.96) and 0.67 (0.58, 0.78), respectively for AD⁽¹³³⁾. In the most comprehensive and granular meta-analysis, Zhang *et al.* combine data from twenty-one individual studies (181 580 participants) with 4438 cases, during follow-up periods ranging from 2.1 to 21 years to examine associations between fish, total PUFA and individual PUFA intakes and total dementia and dementia sub-types⁽¹³⁶⁾. The main findings is that an increase in fish of one serving per week is associated with a lower RR (95% CI) of dementia and AD of 0.95 (0.90, 0.99) and 0.93 (0.90, 0.95), with an equivalent RR for a 0.1 g/d increment of dietary DHA intake (but not EPA) of 0.86 (0.76, 0.96) and 0.63 (0.51, 0.76), respectively. This effect size for one portion of fish

and AD is relatively consistent with Wu *et al.*'s analysis who reported that an increment of 100 g/week of fish intake (UK portion is 140 g) was associated with an 11 % lower risk of AD (RR = 0.89, 95 % CI 0.79, 0.99)⁽¹³⁵⁾.

There is conflicting evidence that associations may be influenced by *APOE* genotype status, with some prospective cohorts reporting no influence^(130,132,134), some no benefits of fish or EPA/DHA intake in *APOE4* carriers^(137–139) and some reporting a beneficial association only in *APOE4*⁽¹²⁸⁾. These apparent inconsistencies are likely attributable to a lack of a granular understanding of influencers of brain DHA metabolism in *APOE4*. It is possible that due to a defective brain DHA uptake and metabolism there is a greater DHA need throughout life in *APOE4*. However, with variables such as age, menopause and significant pathology potentially having a greater impact on brain DHA uptake in *APOE4*, beyond a certain physiological stage an increased DHA intake or blood status may have a lower or negligible cognitive benefit in *APOE4* as it will not translate into higher brain DHA levels. More research is needed to identify the optimal DHA intake and supplementation 'window' in *APOE4*.

Therefore overall, in prospective cohort studies high v. low/no fish and LC *n*-3 PUFA consumption is associated with up to a 40 % reduced risk of total dementia, and in particular AD, with effect sizes equivalent to several years of ageing. It is likely the benefits of fish consumption extend beyond the provision of LC *n*-3 PUFAs, with fish also being a rich sources of selenium, vitamin B₁₂ and vitamin D, all of which may enhance cognition⁽¹⁸⁾. For a dietary component such as DHA/fish which is considered a signature of affluence and an overall healthy diet and lifestyle⁽¹³²⁾ the possibility of residual confounding should be considered, with some of the cognitive benefits seen in prospective cohorts, potentially due to as yet unknown factors which are not fully corrected for in the statistical models.

Fish and DHA intake and status and cognition: randomised controlled trial evidence

To the best of our knowledge, there is currently no RCT which has investigated the impact of fish intake in isolation (i.e. not as part of a multi-food or whole diet intervention) on cognitive outcomes. Prospective cohort evidence where EPA and DHA are predominantly derived from fish, have examined the impact on dementia risk and cognition over follow-up periods up to 20 years. In contrast, RCTs have intervened with a mixed LC *n*-3 PUFA or DHA-rich supplement for up to 3 years, but typically 6 months, which have produced mixed and often null findings (Table 1).

Cognitive benefits of EPA + DHA supplementation have not been observed in AD patients⁽¹⁴⁰⁾. In the Alzheimer's Disease Cooperative study, supplementation with 2.0 g DHA for 18 months in those with mild to moderate AD, did not affect the co-primary outcome measures, the cognitive subscale of the Alzheimer's Disease Assessment Scale (ADAS-cog) and the clinical dementia rating sum of boxes⁽¹⁴⁰⁾. An effect of

intervention on the cognitive subscale of the Alzheimer's Disease Assessment Scale and the minimal state examination was, however, observed in *APOE4* non-carriers. In a 2008 RCT in three-hundred and two cognitively healthy individuals, no effect of doses of 400 or 1900 mg EPA + DHA daily on a range of cognitive outcomes was evident⁽¹⁴¹⁾. This is in contrast to the findings of Stonehouse and colleagues who observed a significant impact of 1160 mg DHA + 170 mg EPA daily over 6 months, on the speed of episodic and working memory, and episodic memory performance in women only, with low habitual EPA and DHA intake at baseline (<200 mg/week)⁽¹⁴²⁾ with Yurko-Mauro *et al.*, also observing improvements in a number of cognitive outcomes supplementing with 900 mg daily in those with subjective memory complaints for 6 months⁽¹⁴³⁾. This is in contrast to the Multidomain Alzheimer Preventive Trial (MAPT) (3 years, *n* 1680)⁽¹⁴⁴⁾ and a more recent RCT (18 m, *n* 403)⁽¹⁴⁵⁾ who observed no impact of 800 mg DHA plus 225 mg EPA daily or 1720 mg DHA plus 600 mg EPA, respectively, on cognitive performance. The MAPT intervention highlights the importance of participant selection, with a much higher mean education attainment in the study group relative to a general French population, which may provide cognitive reserve and have contributed to the lower than expected decline in cognitive function in the MAPT control group⁽¹⁴⁴⁾ and also a number of other RCTs^(146,147), which impacts study power. A secondary analysis in MAPT, in the control group showed the greatest cognitive decline in participants with the lowest quartile (Q1) v. Q2–Q4 of baseline erythrocyte EPA + DHA *n*-3 index (EPA + DHA), with the optimal *n*-3 index cut-off for predicting notable cognitive decline calculated as 5.3 %⁽¹⁴⁸⁾. Using this cut-off, there was a consistent but non-significant difference in 3-year cognitive decline between EPA + DHA treated and placebo subjects with 'low' baseline *n*-3 index. The authors concluded that those with an *n*-3 index below approximately 5 % are at an increased risk of cognitive decline and could be a good target for recruiting a responsive population subgroup.

Intervention studies which target cognition have predominantly fed DHA-rich or DHA-only supplements as the bioactive LC *n*-3 PUFA, based on the observation that brain DHA levels are >250 higher than EPA⁽¹⁴⁹⁾. EPA does enter the brain with uptake efficiencies equivalent to DHA, but is thought to be rapidly metabolised following entry, although concentrations of EPA are higher than DHA in microglial cells⁽¹⁴⁹⁾. The impact of EPA on cognition and depression is being increasingly recognised^(149–151). In a recent intervention, Patan and colleagues observed a significant effect of an EPA-rich oil on cognitive global accuracy and speed relative to a DHA-rich or placebo oil fed for 6 months (Table 1). Future interventions should not only consider what dose, but also what DHA:EPA ratio of the supplement and its chemical form (ethyl ester, TAG and phospholipid LPC)⁽¹⁵²⁾.

The prodrome of AD is thought to be 20–30 years or potentially longer⁽¹⁵³⁾. Therefore, cognitive assessment or

Table 1. Select RCTs of nutrition interventions to improve cognition

Study	Population	Intervention	Duration	Outcome	Comment
van de Rest <i>et al.</i> ⁽¹⁴¹⁾	Cognitively healthy (<i>n</i> 302, 70 years)	High dose: 847 mg DHA + 1093 mg EPA daily Low dose: 176 mg DHA + 226 mg EPA daily	6 months	No effect on a range of cognitive domains	Plasma concentrations of EPA + DHA increased by 238% in the high-dose and 51% in the low-dose groups compared with placebo
Dangour <i>et al.</i> ⁽¹⁴⁶⁾	Cognitively healthy (<i>n</i> 867, 74 years)	500 mg DHA + 200 mg EPA daily	24 months	No effect on the California Verbal Learning Test	No effect on global cognitive function, memory, processing, executive and global delay z scores High fish intake at baseline in some (Table 1) Lack of expected cognitive decline in the control arm
Quinn <i>et al.</i> ⁽¹⁴⁰⁾	Mild to moderate AD (<i>n</i> 402, 76 years)	2 g DHA daily	18 months	No effect on the Alzheimer's Disease Assessment-cognition Scale (ADAS-cog) No effect on CDR	Supplementation did not slow cognitive decline in patients with AD MMSE and ADAS-cog improved in APOE4 non-carriers
Yurko-Mauro <i>et al.</i> ⁽¹⁴³⁾	SMC (<i>n</i> 485, 70 years)	900 mg DHA daily	6 months	Improved CANTAB Paired Associate Learning, a visuospatial learning and episodic memory test	DHA also improved immediate and delayed verbal recognition memory scores but not working memory
Stonehouse <i>et al.</i> ⁽¹⁴²⁾	Healthy adults (<i>n</i> 228, 33 years)	1160 mg DHA + 170 mg EPA daily	6 months	Effect on memory accuracy and reaction time	Screening process only recruited those with a low habitual intake of EPA + DHA of <200 mg/week
Andrieu <i>et al.</i> ⁽¹⁴⁴⁾	SMC (<i>n</i> 1680, 70 years)	Multi-domain intervention with 800 mg DHA and 225 mg EPA daily or EPA + DHA alone	36 months	No effect of the multi-domain intervention and EPA + DHA alone	Cognitive tests included a composite score on the free and cued selective reminding test, ten mini-mental state examination orientation items, digit symbol substitution test and category naming test
Soininen <i>et al.</i> ^(147,155)	Prodromal AD (<i>n</i> 311, 71 years)	1200 mg DHA, 300 mg EPA + phospholipids, uridine monophosphate, choline, vitamins B ₁₂ , B ₆ , folic acid, C, E and selenium	24 months 36 months	No effect on the Neurocognitive Test Battery (NTB) primary outcome Effect on clinical dementia rating (CDR) Effect on hippocampal volume Effect on NTB Effect on CDR and memory Effect on hippocampal and whole brain volume	Unexpectedly lower rate of cognitive decline in the control group No effect on whole brain volume or memory No effect on executive function

Zhang <i>et al.</i> ⁽¹⁴⁶⁾	MCI (n 240, 74 years)	2 g DHA daily	24 months	Effect on intelligence quotient, and information and digit span Effect on Aβ-42 level No effect of treatment on reasoning, working memory, short-term memory, retrieval fluency and cognitive speed-related constructs	Daily DHA may improve cognition and change Aβ-mediated autophagy
Danthiir <i>et al.</i> ⁽¹⁴⁵⁾	Cognitively healthy, (n 403, 73 years)	1720 mg DHA and 600 mg EPA daily	18 months		A negative main effect was found on psychomotor speed some sex and APOE genotype interactions evident
Patan <i>et al.</i> ⁽¹⁵⁰⁾	Healthy adults (n 310, 36 years)	900 mg DHA and 270 mg EPA daily (DHA-rich oil), 360 mg DHA and 900 mg EPA daily (EPA-rich oil)	6 months	Both global accuracy and speed improved with EPA-rich oil compared with placebo and DHA-rich oil	Accuracy of memory was improved with EPA- compared with DHA-rich oil Both EPA- and DHA-rich oils showed trends towards reduced prefrontal cortex oxygenated Hb

AD, Alzheimer's disease; Aβ, amyloid-β; CANTAB, Cambridge neuropsychological test automated battery; MCI, mild cognitive impairment; MMSE, mini-mental state examination; RCT, randomised controlled trial; SMC, subjective memory complaints. This table is by no means exhaustive. The RCTs included were >6 months in duration and were selected to demonstrate the discordance between individual study findings.

brain volume and atrophy (assessed by MRI) rather than incident disease have to date been exclusively used as primary RCT outcomes. Cognitive questionnaires and other assessment tools historically may not have been fully fit for purpose, lacking the specificity and sensitivity to detect subtle effect of intervention on specific cognitive domains. The variability and lack of a standard battery of cognitive tests employed in cognitive RCTs is likely to be a large contributor to the heterogeneity in finding, between trials.

Furthermore, due attention is not given to the length of the intervention period. As brain DHA half-life is estimated to be 2.5 years⁽¹⁵⁴⁾, supplementation periods of at least 1 year are likely to be needed to detect the cognitive benefits associated with DHA enrichment of neuronal cells, such as effects on dendrite outgrowth and spine density, synaptic function and Aβ processing. The impact of intervention period on the study conclusions is evidenced by comparing the 24 month and 36 month findings from the LipiDiDiet study which fed the Souvenaid (Fortasyn Connect) medicinal food, which combines 1200 mg DHA and 300 mg EPA with phospholipids, uridine monophosphate, choline, vitamins B₁₂, B₆, folic acid, C, E and selenium. At 24 months, although an effect of Souvenaid on secondary outcomes was observed (hippocampal volume and clinical dementia rating score) no effect of the intervention on the primary outcome, the neurocognitive test battery performance, was evident. By 36 months the intervention had significantly increased the neurocognitive test battery test score by 60% relative to the control group, with the greatest cognitive benefits (based on the clinical dementia rating score) evident in those with the highest cognitive status at baseline (mini-mental state examination ≥29).

There is a strong justification and need to conduct a future RCT in 'at-risk' cognitively healthy participants with incident dementia or AD as the primary outcome. Such a trial is likely to require at least a 5-year intervention period and several thousand per intervention arm, given the long prodrome and AD incident rates. Careful enrichment of the trial with an at-risk responsive population based on such factors at APOE genotype status, cardiovascular risk profile, baseline EPA + DHA status, education attainment, brain imaging and blood biomarker profiles is key to success. Consideration should also be given not only to the LC n-3 PUFA dose, but also the DHA:EPA ratio and chemical form.

Final thoughts

The prospective and experimental evidence for the role of DHA in brain function and the cognitive benefits of increased fish, and LC n-3 PUFA intakes are convincing with large effect sizes. It is likely that EPA and DHA have complementary neurophysiological benefits, which includes an effect on oxylipin production, and should be co-supplemented or ideally consumed as oily rich fish. Confirmation of the cognitive benefits is needed from the well-designed RCTs which include large population subgroups who are likely to be most responsive

and gain most benefit. Accumulating evidence suggests that *APOE4* carriers have a lower brain uptake and status and would particularly benefit from DHA intervention prior to any significant neuropathology, which affects brain DHA uptake.

Financial Support

Financial support for our work included in the review has been provided by the BBSRC (BB/M004449/1), EU-JPI/BBSRC, (BB/P028233/1) Alzheimer's Society (AS-PhD-2015-023), Alzheimer's Research UK (PRRF2017-006), the UK Foods Standards Agency (N05065 and N05066) and the UK Medical Research Council (U105960389 and U1052.00.014).

Conflict of Interest

None.

Authorship

The authors had sole responsibility for all aspects of preparation of this paper.

References

1. Patterson C (2018) World Alzheimer Report. Alzheimer's Disease International (ADI), London.
2. (2019) Global, regional, and national burden of Alzheimer's disease and other dementias, 1990–2016: a systematic analysis for the global burden of disease study 2016. *Lancet Neurol* **18**, 88–106.
3. Prince M, Knapp M, Guerchet M *et al.* (2014) Dementia UK: Update. Alzheimer's Society, with King's College London and the London School of Economics.
4. ONS (2018) Deaths registered in England and Wales: 2017. In: Statistics OoN, editor.
5. Ferretti MT, Iulita MF, Cavedo E *et al.* (2018) Sex differences in Alzheimer disease – the gateway to precision medicine. *Nat Rev Neurol* **14**, 457–469.
6. Fisher DW, Bennett DA & Dong H (2018) Sexual dimorphism in predisposition to Alzheimer's disease. *Neurobiol Aging* **70**, 308–324.
7. Oveisgharan S, Arvanitakis Z, Yu L *et al.* (2018) Sex differences in Alzheimer's disease and common neuropathologies of aging. *Acta Neuropathol* **136**, 887–900.
8. Neu SC, Pa J, Kukull W *et al.* (2017) Apolipoprotein E genotype and sex risk factors for Alzheimer disease: a meta-analysis. *JAMA Neurol* **74**, 1178–1189.
9. Pontifex M, Vauzour D & Minihane AM (2018) The effect of APOE genotype on Alzheimer's disease risk is influenced by sex and docosahexaenoic acid status. *Neurobiol Aging* **69**, 209–220.
10. Pontifex MG, Martinsen A, Saleh RNM *et al.* (2021) APOE4 genotype exacerbates the impact of menopause on cognition and synaptic plasticity in APOE-TR mice. *FASEB J* **35**, e21583.
11. Rahman A, Schelbaum E, Hoffman K *et al.* (2020) Sex-driven modifiers of Alzheimer risk: a multimodality brain imaging study. *Neurology* **95**, e166–e178.
12. Walsh S, Merrick R, Milne R *et al.* (2021) Aducanumab for Alzheimer's disease? *Br Med J* **374**, n1682.
13. Livingston G, Huntley J, Sommerlad A *et al.* (2020) Dementia prevention, intervention, and care: 2020 report of the lancet commission. *Lancet* **396**, 413–446.
14. Solfrizzi V, Custodero C, Lozupone M *et al.* (2017) Relationships of dietary patterns, foods, and micro- and macronutrients with Alzheimer's disease and late-life cognitive disorders: a systematic review. *J Alzheimer's Dis: JAD* **59**, 815–849.
15. Barbaresko J, Lellmann AW, Schmidt A *et al.* (2020) Dietary factors and neurodegenerative disorders: an umbrella review of meta-analyses of prospective studies. *Adv Nut* **11**, 1161–1173.
16. Pottala JV, Yaffe K, Robinson JG *et al.* (2014) Higher RBC EPA + DHA corresponds with larger total brain and hippocampal volumes: WHIMS-MRI study. *Neurology* **82**, 435–442.
17. SACN (2018) SACN statement on diet, cognitive impairment and dementias. In *Scientific Advisory Commission on Nutrition DoH*, London: The Stationary Office (UK Government).
18. Scarmeas N, Anastasiou CA & Yannakoulia M (2018) Nutrition and prevention of cognitive impairment. *Lancet Neurol* **17**, 1006–1015.
19. Shishtar E, Rogers GT, Blumberg JB *et al.* (2020) Long-term dietary flavonoid intake and change in cognitive function in the Framingham offspring cohort. *Public Health Nutr* **23**, 1–13.
20. Jennings A, Cunnane SC & Minihane AM (2020) Can nutrition support healthy cognitive ageing and reduce dementia risk? *Br Med J* **369**, m2269.
21. Melo van Lent D, O'Donnell A, Beiser AS *et al.* (2021) Mind diet adherence and cognitive performance in the Framingham heart study. *J Alzheimer's Dis: JAD* **82**, 827–839.
22. Shannon OM, Stephan BCM, Granic A *et al.* (2019) Mediterranean diet adherence and cognitive function in older UK adults: the European prospective investigation into cancer and nutrition-Norfolk (EPIC-Norfolk) study. *Am J Clin Nutr* **110**, 938–948.
23. Wu L & Sun D (2017) Adherence to Mediterranean diet and risk of developing cognitive disorders: an updated systematic review and meta-analysis of prospective cohort studies. *Sci Rep* **7**, 41317.
24. Livingston G, Sommerlad A, Orgeta V *et al.* (2017) Dementia prevention, intervention, and care. *Lancet* **390**, 2673–2734.
25. WHO Guidelines (2019) Risk reduction of cognitive decline and dementia: Report No.: ISBN-13: 978-92-4-155054-3.
26. Burdge GC, Finnegan YE, Minihane AM *et al.* (2003) Effect of altered dietary n-3 fatty acid intake upon plasma lipid fatty acid composition, conversion of [¹³C]alpha-linolenic acid to longer-chain fatty acids and partitioning towards beta-oxidation in older men. *Br J Nutr* **90**, 311–321.
27. Arterburn LM, Hall EB & Oken H (2006) Distribution, interconversion, and dose response of n-3 fatty acids in humans. *Am J Clin Nutr* **83**(6 Suppl), 1467s–1476s.
28. Crawford MA, Casperd NM & Sinclair AJ (1976) The long chain metabolites of linoleic acid linolenic acids in liver and brain in herbivores and carnivores. *Comp Biochem Physiol B* **54**, 395–401.
29. Zhao Z, Nelson AR, Betsholtz C *et al.* (2015) Establishment and dysfunction of the blood–brain barrier. *Cell* **163**, 1064–1078.



30. Bazinet RP & Layé S (2014) Polyunsaturated fatty acids and their metabolites in brain function and disease. *Nat Rev Neurosci* **15**, 771–785.
31. Lacombe RJS, Chouinard-Watkins R & Bazinet RP (2018) Brain docosahexaenoic acid uptake and metabolism. *Mol Aspects Med* **64**, 109–134.
32. Mitchell RW, On NH, Del Bigio MR *et al.* (2011) Fatty acid transport protein expression in human brain and potential role in fatty acid transport across human brain microvessel endothelial cells. *J Neurochem* **117**, 735–746.
33. Ochiai Y, Uchida Y, Ohtsuki S *et al.* (2017) The blood–brain barrier fatty acid transport protein 1 (FATP1/SLC27A1) supplies docosahexaenoic acid to the brain, and insulin facilitates transport. *J Neurochem* **141**, 400–412.
34. Ochiai Y, Uchida Y, Tachikawa M *et al.* (2019) Amyloid beta(25–35) impairs docosahexaenoic acid efflux by down-regulating fatty acid transport protein 1 (FATP1/SLC27A1) protein expression in human brain capillary endothelial cells. *J Neurochem* **150**, 385–401.
35. Chouinard-Watkins R & Bazinet RP (2018) ACSL6 is critical for maintaining brain DHA levels. *Proc Natl Acad Sci USA* **115**, 12343–5.
36. Fernandez RF, Kim SQ, Zhao Y *et al.* (2018) Acyl-CoA synthetase 6 enriches the neuroprotective omega-3 fatty acid DHA in the brain. *Proc Natl Acad Sci USA* **115**, 12525–12530.
37. Chan JP, Wong BH, Chin CF *et al.* (2018) The lysolipid transporter Mfsd2a regulates lipogenesis in the developing brain. *PLoS Biol* **16**, e2006443.
38. Nguyen LN, Ma D, Shui G *et al.* (2014) Mfsd2a is a transporter for the essential omega-3 fatty acid docosahexaenoic acid. *Nature* **509**, 503–506.
39. Alakbarzade V, Hameed A, Quek DQY *et al.* (2015) A partially inactivating mutation in the sodium-dependent lysophosphatidylcholine transporter MFSD2A causes a non-lethal microcephaly syndrome. *Nat Genet* **47**, 814–817.
40. Wong BH & Silver DL (2020) Mfsd2a: a physiologically important lysolipid transporter in the brain and Eye. In *Lipid Transfer in Lipoprotein Metabolism and Cardiovascular Disease*, pp. 223–234 [X-C Jiang, editor]. Singapore: Springer Singapore.
41. Ben-Zvi A, Lacoste B, Kur E *et al.* (2014) Mfsd2a is critical for the formation and function of the blood–brain barrier. *Nature* **509**, 507–511.
42. Eser Ocak P, Ocak U, Sherchan P *et al.* (2020) Insights into major facilitator superfamily domain-containing protein-2a (Mfsd2a) in physiology and pathophysiology. What do we know so far? *J Neurosci Res* **98**, 29–41.
43. Chouinard-Watkins R, Lacombe RJS & Bazinet RP (2018) Mechanisms regulating brain docosahexaenoic acid uptake: what is the recent evidence? *Curr Opin Clin Nutr Metab Care* **21**, 71–77.
44. Yassine HN, Braskie MN, Mack WJ *et al.* (2017) Association of docosahexaenoic acid supplementation with Alzheimer disease stage in apolipoprotein E ε4 carriers: a review. *JAMA Neurol* **74**, 339–347.
45. Zhong N & Weisgraber KH (2009) Understanding the association of apolipoprotein E4 with Alzheimer disease: clues from its structure. *J Biol Chem* **284**, 6027–6031.
46. Frieden C & Garai K (2012) Structural differences between apoE3 and apoE4 may be useful in developing therapeutic agents for Alzheimer's disease. *Proc Natl Acad Sci USA* **109**, 8913–8918.
47. Abondio P, Sazzini M, Garagnani P *et al.* (2019) The genetic variability of APOE in different human populations and Its implications for longevity. *Genes* **10**, 222–250.
48. Liu CC, Liu CC, Kanekiyo T *et al.* (2013) Apolipoprotein E and Alzheimer disease: risk, mechanisms and therapy. *Nat Rev Neurol* **9**, 106–118.
49. Farrer LA, Cupples LA, Haines JL *et al.* (1997) Effects of age, sex, and ethnicity on the association between apolipoprotein E genotype and Alzheimer disease. A meta-analysis. APOE and Alzheimer disease meta analysis consortium. *JAMA* **278**, 1349–1356.
50. Corder EH, Saunders AM, Strittmatter WJ *et al.* (1993) Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families. *Science* **261**, 921–923.
51. Yassine HN & Finch CE (2020) APOE Alleles and diet in brain aging and Alzheimer's disease. *Front Aging Neurosci* **12**, 150.
52. Vandal M, Alata W, Tremblay C *et al.* (2014) Reduction in DHA transport to the brain of mice expressing human APOE4 compared to APOE2. *J Neurochem* **129**, 516–526.
53. Martinsen A, Tejera N, Vauzour D *et al.* (2019) Altered SPMs and age-associated decrease in brain DHA in APOE4 female mice. *FASEB J* **33**, 10315–10326.
54. Chouinard-Watkins R, Conway V, Minihane AM *et al.* (2015) Interaction between BMI and APOE genotype is associated with changes in the plasma long-chain-PUFA response to a fish-oil supplement in healthy participants. *Am J Clin Nutr* **102**, 505–513.
55. Chouinard-Watkins R, Rioux-Perreault C, Fortier M *et al.* (2013) Disturbance in uniformly ¹³C-labelled DHA metabolism in elderly human subjects carrying the apoE ε4 allele. *Br J Nutr* **110**, 1751–1759.
56. Yassine HN, Rawat V, Mack WJ *et al.* (2016) The effect of APOE genotype on the delivery of DHA to cerebrospinal fluid in Alzheimer's disease. *Alzheimer's Res Ther* **8**, 25.
57. Klenk E & Bongard W (1952) Constitution of the unsaturated C20 and C22 fatty acids of the glycerophosphatides of the brain. *Hoppe Seylers Z Physiol Chem* **291**, 104–118.
58. Belayev L, Hong SH, Menghani H *et al.* (2018) Docosanoids promote neurogenesis and angiogenesis, blood-brain barrier integrity, penumbra protection, and neurobehavioral recovery after experimental ischemic stroke. *Mol Neurobiol* **55**, 7090–7106.
59. Bradbury J (2011) Docosahexaenoic acid (DHA): an ancient nutrient for the modern human brain. *Nutrients* **3**, 529–554.
60. Díaz M, Mesa-Herrera F & Marín R (2021) DHA and its elaborated modulation of antioxidant defenses of the brain: implications in aging and AD neurodegeneration. *Antioxidants* **10**, 907–933.
61. Howe PRC, Evans HM, Kuszewski JC *et al.* (2018) Effects of long chain omega-3 polyunsaturated fatty acids on brain function in mildly hypertensive older adults. *Nutrients* **10**, 1412–1427.
62. Hur J, Mateo V, Amalric N *et al.* (2018) Cerebrovascular β-amyloid deposition and associated microhemorrhages in a Tg2576 Alzheimer mouse model are reduced with a DHA-enriched diet. *FASEB J* **32**, 4972–4983.
63. Liu ZH, Chen NY, Tu PH *et al.* (2020) DHA attenuates cerebral edema following traumatic brain injury via the reduction in blood-brain barrier permeability. *Int J Mol Sci* **21**, 6291–6309.
64. Eady TN, Belayev L, Khoutorova L *et al.* (2012) Docosahexaenoic acid signaling modulates cell survival in experimental ischemic stroke penumbra and initiates long-term repair in young and aged rats. *PLoS ONE* **7**, e46151.
65. Belkouch M, Hachem M, Elgot A *et al.* (2016) The pleiotropic effects of omega-3 docosahexaenoic acid on the

- hallmarks of Alzheimer's disease. *J Nutr Biochem* **38**, 1–11.
66. Weiser MJ, Butt CM & Mohajeri MH (2016) Docosahexaenoic acid and cognition throughout the lifespan. *Nutrients* **8**, 99.
 67. Sinclair AJ (2019) Docosahexaenoic acid and the brain – what is its role? *Asia Pac J Clin Nutr* **28**, 675–688.
 68. Kowiański P, Lietzau G, Czuba E *et al.* (2018) BDNF: a key factor with multipotent impact on brain signaling and synaptic plasticity. *Cell Mol Neurobiol* **38**, 579–593.
 69. Wu A, Ying Z & Gomez-Pinilla F (2004) Dietary omega-3 fatty acids normalize BDNF levels, reduce oxidative damage, and counteract learning disability after traumatic brain injury in rats. *J Neurotrauma* **21**, 1457–1467.
 70. Hooijmans CR, Pasker-de Jong PC, de Vries RB *et al.* (2012) The effects of long-term omega-3 fatty acid supplementation on cognition and Alzheimer's pathology in animal models of Alzheimer's disease: a systematic review and meta-analysis. *J Alzheimer's Dis: JAD* **28**, 191–209.
 71. Chataigner M, Martin M, Lucas C *et al.* (2021) Fish hydrolysate supplementation containing *n*-3 long chain polyunsaturated fatty acids and peptides prevents LPS-induced neuroinflammation. *Nutrients* **13**, 824–845.
 72. Serhan CN, Dalli J, Colas RA *et al.* (2015) Protectins and maresins: new pro-resolving families of mediators in acute inflammation and resolution bioactive metabolome. *Biochim Biophys Acta* **1851**, 397–413.
 73. Joffre C, Dinel AL, Chataigner M *et al.* (2020). *n*-3 Polyunsaturated fatty acids and their derivatives reduce neuroinflammation during aging. *Nutrients* **12**, 647–672.
 74. Orr SK, Palumbo S, Bosetti F *et al.* (2013) Unesterified docosahexaenoic acid is protective in neuroinflammation. *J Neurochem* **127**, 378–393.
 75. Song C, Manku MS & Horrobin DF (2008) Long-chain polyunsaturated fatty acids modulate interleukin-1 β -induced changes in behavior, monoaminergic neurotransmitters, and brain inflammation in rats. *J Nutr* **138**, 954–963.
 76. Devassy JG, Leng S, Gabbs M *et al.* (2016) Omega-3 polyunsaturated fatty acids and oxylipins in neuroinflammation and management of Alzheimer disease. *Adv Nut* **7**, 905–916.
 77. Fillit H, Ding W, Buee L *et al.* (1991) Elevated circulating tumor necrosis factor levels in Alzheimer's disease. *Neurosci Lett* **129**, 318–320.
 78. Strauss S, Bauer J, Ganter U *et al.* (1992) Detection of interleukin-6 and α 2-macroglobulin immunoreactivity in cortex and hippocampus of Alzheimer's disease patients. *Lab Invest* **66**, 223–230.
 79. Liu B & Hong JS (2003) Role of microglia in inflammation-mediated neurodegenerative diseases: mechanisms and strategies for therapeutic intervention. *J Pharmacol Exp Ther* **304**(1), 1–7.
 80. Sastre M, Klockgether T & Heneka MT (2006) Contribution of inflammatory processes to Alzheimer's disease: molecular mechanisms. *Int J Dev Neurosci* **24**, 167–176.
 81. Wyss-Coray T (2006) Inflammation in Alzheimer disease: driving force, bystander or beneficial response? *Nat Med* **12**, 1005–1015.
 82. Calsolaro V & Edison P (2016) Neuroinflammation in Alzheimer's disease: current evidence and future directions. *Alzheimer's Dement* **12**, 719–732.
 83. Cai Z, Hussain MD & Yan LJ (2014) Microglia, neuroinflammation, and beta-amyloid protein in Alzheimer's disease. *Int J Neurosci* **124**, 307–321.
 84. Dani M, Wood M, Mizoguchi R *et al.* (2018) Microglial activation correlates *in vivo* with both tau and amyloid in Alzheimer's disease. *Brain* **141**, 2740–2754.
 85. Chandra A, Valkimadi P-E, Pagano G *et al.* (2019) Applications of amyloid, tau, and neuroinflammation PET imaging to Alzheimer's disease and mild cognitive impairment. *Hum Brain Mapp* **40**, 5424–5442.
 86. Fan Z, Okello AA, Brooks DJ *et al.* (2015) Longitudinal influence of microglial activation and amyloid on neuronal function in Alzheimer's disease. *Brain* **138**, 3685–3698.
 87. Jack CR, Knopman DS, Jagust WJ *et al.* (2013) Tracking pathophysiological processes in Alzheimer's disease: an updated hypothetical model of dynamic biomarkers. *Lancet Neurol* **12**, 207–216.
 88. Dinarello CA (2011) Interleukin-1 in the pathogenesis and treatment of inflammatory diseases. *Blood* **117**, 3720–3732.
 89. Calder PC (2017) Omega-3 fatty acids and inflammatory processes: from molecules to man. *Biochem Soc Trans* **45**, 1105–1115.
 90. Simopoulos AP (2002) The importance of the ratio of omega-6/omega-3 essential fatty acids. *Biomed Pharmacother* **56**, 365–379.
 91. Orr SK, Trépanier MO & Bazinet RP (2013). *n*-3 polyunsaturated fatty acids in animal models with neuroinflammation. *Prostaglandins Leukot Essent Fatty Acids* **88**, 97–103.
 92. Layé S, Nadjar A, Joffre C *et al.* (2018) Anti-inflammatory effects of omega-3 fatty acids in the brain: physiological mechanisms and relevance to pharmacology. *Pharmacol Rev* **70**, 12–38.
 93. Fourrier C, Remus-Borel J, Greenhalgh AD *et al.* (2017) Docosahexaenoic acid-containing choline phospholipid modulates LPS-induced neuroinflammation *in vivo* and in microglia *in vitro*. *J Neuroinflammation* **14**, 170.
 94. Chen X, Chen C, Fan S *et al.* (2018) Omega-3 polyunsaturated fatty acid attenuates the inflammatory response by modulating microglia polarization through SIRT1-mediated deacetylation of the HMGB1/NF- κ B pathway following experimental traumatic brain injury. *J Neuroinflammation* **15**, 116.
 95. Vom Berg J, Prokop S, Miller KR *et al.* (2012) Inhibition of IL-12/IL-23 signaling reduces Alzheimer's disease-like pathology and cognitive decline. *Nat Med* **18**, 1812–1819.
 96. Gabbs M, Leng S, Devassy JG *et al.* (2015) Advances in our understanding of oxylipins derived from dietary PUFAs. *Adv Nut* **6**, 513–540.
 97. Schulze MB, Minihane AM, Saleh RNM *et al.* (2020) Intake and metabolism of omega-3 and omega-6 polyunsaturated fatty acids: nutritional implications for cardiometabolic diseases. *Lancet Diabetes Endocrinol* **8**, 915–930.
 98. Anderson EJ & Taylor DA (2012) Stressing the heart of the matter: re-thinking the mechanisms underlying therapeutic effects of *n*-3 polyunsaturated fatty acids. *F1000 Med Rep* **4**, 13.
 99. Buckley CD, Gilroy DW & Serhan CN (2014) Proresolving lipid mediators and mechanisms in the resolution of acute inflammation. *Immunity* **40**, 315–327.
 100. Christie WW & Harwood JL (2020) Oxidation of polyunsaturated fatty acids to produce lipid mediators. *Essays Biochem* **64**, 401–421.
 101. Wada M, DeLong CJ, Hong YH *et al.* (2007) Enzymes and receptors of prostaglandin pathways with arachidonic acid-derived versus eicosapentaenoic acid-derived substrates and products. *J Biol Chem* **282**, 22254–22266.

102. Kodani SD & Hammock BD (2015) The 2014 Bernard B. Brodie award lecture-epoxide hydrolases: drug metabolism to therapeutics for chronic pain. *Drug Metab Dispos* **43**, 788–802.
103. Node K, Huo Y, Ruan X *et al.* (1999) Anti-inflammatory properties of cytochrome P450 epoxygenase-derived eicosanoids. *Science* **285**, 1276–1279.
104. Inceoglu B, Wagner K, Schebb NH *et al.* (2011) Analgesia mediated by soluble epoxide hydrolase inhibitors is dependent on cAMP. *Proc Natl Acad Sci USA* **108**, 5093–5097.
105. Seki H, Fukunaga K, Arita M *et al.* (2010) The anti-inflammatory and proresolving mediator resolvin E1 protects mice from bacterial pneumonia and acute lung injury. *J Immunol* **184**, 836–843.
106. Herová M, Schmid M, Gemperle C *et al.* (2015) ChemR23, the receptor for chemerin and resolvin E1, is expressed and functional on M1 but not on M2 macrophages. *J Immunol* **194**, 2330–2337.
107. Arita M, Bianchini F, Aliberti J *et al.* (2005) Stereochemical assignment, antiinflammatory properties, and receptor for the omega-3 lipid mediator resolvin E1. *J Exp Med* **201**, 713–722.
108. Miyazawa K, Fukunaga H, Tatewaki Y *et al.* (2020) Alzheimer's disease and specialized pro-resolving lipid mediators: do MaR1, RvD1, and NPD1 show promise for prevention and treatment? *Int J Mol Sci* **21**, 5783–5797.
109. Whittington RA, Planel E & Terrando N (2017) Impaired resolution of inflammation in Alzheimer's disease: a review. *Front Immunol* **8**, 1464–1473.
110. Dunn HC, Ager RR, Baglietto-Vargas D *et al.* (2015) Restoration of lipoxin A4 signaling reduces Alzheimer's disease-like pathology in the 3xTg-AD mouse model. *J Alzheimer's Dis: JAD* **43**, 893–903.
111. Terrando N, Gómez-Galán M, Yang T *et al.* (2013) Aspirin-triggered resolvin D1 prevents surgery-induced cognitive decline. *FASEB J* **27**, 3564–3571.
112. Hong S, Gronert K, Devchand PR *et al.* (2003) Novel docosatrienes and 17S-resolvins generated from docosahexaenoic acid in murine brain, human blood, and glial cells: autacoids in anti-inflammation. *J Biol Chem* **278**, 14677–14687.
113. Qu L & Caterina MJ (2018) Accelerating the reversal of inflammatory pain with NPD1 and its receptor GPR37. *J Clin Invest* **128**, 3246–3249.
114. Marcheselli VL, Hong S, Lukiw WJ *et al.* (2003) Novel docosanoids inhibit brain ischemia-reperfusion-mediated leukocyte infiltration and pro-inflammatory gene expression. *J Biol Chem* **278**, 43807–43817.
115. Asatryan A & Bazan NG (2017) Molecular mechanisms of signaling via the docosanoid neuroprotectin D1 for cellular homeostasis and neuroprotection. *J Biol Chem* **292**, 12390–7.
116. Wang X, Zhu M, Hjorth E *et al.* (2015) Resolution of inflammation is altered in Alzheimer's disease. *Alzheimer's Dement* **11**, 40–50, e1–2.
117. Hashimoto M, Katakura M, Tanabe Y *et al.* (2015) n-3 fatty acids effectively improve the reference memory-related learning ability associated with increased brain docosahexaenoic acid-derived docosanoids in aged rats. *Biochim Biophys Acta, Mol Cell Biol Lipids* **1851**, 203–209.
118. Rey C, Delpech JC, Madore C *et al.* (2019) Dietary n-3 long chain PUFA supplementation promotes a pro-resolving oxylipin profile in the brain. *Brain Behav Immun* **76**, 17–27.
119. Ostermann AI & Schebb NH (2017) Effects of omega-3 fatty acid supplementation on the pattern of oxylipins: a short review about the modulation of hydroxy-, dihydroxy-, and epoxy-fatty acids. *Food Funct* **8**, 2355–2367.
120. Saleh RN WA, Ostermann AI, Schebb NH *et al.* (2021) APOE genotype modifies the plasma oxylipin response to omega-3 polyunsaturated fatty acid supplementation in healthy individuals. *Front Nutr* **8**, 723813–723828.
121. Zhao J, Roman MJ, Devereux RB *et al.* (2014) Leukotriene haplotype × diet interaction on carotid artery hypertrophy and atherosclerosis in American Indians: the strong heart family study. *Atherosclerosis* **233**, 165–171.
122. Stephensen CB, Armstrong P, Newman JW *et al.* (2011) ALOX5 gene variants affect eicosanoid production and response to fish oil supplementation. *J Lipid Res* **52**, 991–1003.
123. Kalmijn S, Launer LJ, Ott A *et al.* (1997) Dietary fat intake and the risk of incident dementia in the Rotterdam study. *Ann Neurol* **42**, 776–782.
124. Devore EE, Grodstein F, van Rooij FJ *et al.* (2009) Dietary intake of fish and omega-3 fatty acids in relation to long-term dementia risk. *Am J Clin Nutr* **90**, 170–176.
125. Zhang Y, Zhuang P, He W *et al.* (2018) Association of fish and long-chain omega-3 fatty acids intakes with total and cause-specific mortality: prospective analysis of 421 309 individuals. *J Intern Med* **284**, 399–417.
126. ISSFAL (2004) Report of the Sub-Committee on Recommendations for Intake of Polyunsaturated Fatty Acids in Healthy Adults. International Society for the Study of Fatty Acids and Lipids.
127. SACN (2004) Advice on fish consumption: benefits and risks. London, The Stationary Office: Scientific Advisory Committee on Nutrition (UK Government).
128. Morris MC, Brockman J, Schneider JA *et al.* (2016) Association of seafood consumption, brain mercury level, and APOE ε4 status with brain neuropathology in older adults. *JAMA* **315**, 489–497.
129. Schaefer EJ, Bongard V, Beiser AS *et al.* (2006) Plasma phosphatidylcholine docosahexaenoic acid content and risk of dementia and Alzheimer disease: the Framingham heart study. *Arch Neurol* **63**, 1545–1550.
130. Ammann EM, Pottala JV, Robinson JG *et al.* (2017) Erythrocyte omega-3 fatty acids are inversely associated with incident dementia: secondary analyses of longitudinal data from the Women's Health Initiative Memory Study (WHIMS). *Prostaglandins Leukot Essent Fatty Acids* **121**, 68–75.
131. von Schacky C (2021) Importance of EPA and DHA blood levels in brain structure and function. *Nutrients* **13**, 1074–1092.
132. Thomas A, Baillet M, Proust-Lima C *et al.* (2020) Blood polyunsaturated omega-3 fatty acids, brain atrophy, cognitive decline, and dementia risk. *Alzheimer's Dement* **17**, 407–416.
133. Bakre AT, Chen R, Khutan R *et al.* (2018) Association between fish consumption and risk of dementia: a new study from China and a systematic literature review and meta-analysis. *Public Health Nutr* **21**, 1921–1932.
134. Samieri C, Morris MC, Bennett DA *et al.* (2018) Fish intake, genetic predisposition to Alzheimer disease, and decline in global cognition and memory in 5 cohorts of older persons. *Am J Epidemiol* **187**, 933–940.
135. Wu S, Ding Y, Wu F *et al.* (2015) Omega-3 fatty acids intake and risks of dementia and Alzheimer's disease: a meta-analysis. *Neurosci Biobehav Rev* **48**, 1–9.
136. Zhang Y, Chen J, Qiu J *et al.* (2016) Intakes of fish and polyunsaturated fatty acids and mild-to-severe cognitive impairment risks: a dose-response meta-analysis of 21 cohort studies. *Am J Clin Nutr* **103**, 330–340.
137. Barberger-Gateau P, Samieri C, Féart C *et al.* (2011) Dietary omega 3 polyunsaturated fatty acids and

- Alzheimer's disease: interaction with apolipoprotein E genotype. *Curr Alzheimer Res* **8**, 479–491.
138. Huang TL, Zandi PP, Tucker KL *et al.* (2005) Benefits of fatty fish on dementia risk are stronger for those without APOE epsilon4. *Neurology* **65**, 1409–1414.
 139. Whalley LJ, Deary IJ, Starr JM *et al.* (2008) *n*-3 Fatty acid erythrocyte membrane content, APOE varepsilon4, and cognitive variation: an observational follow-up study in late adulthood. *Am J Clin Nutr* **87**, 449–454.
 140. Quinn JF, Raman R, Thomas RG *et al.* (2010) Docosahexaenoic acid supplementation and cognitive decline in Alzheimer disease: a randomized trial. *JAMA* **304**, 1903–1911.
 141. van de Rest O, Geleijnse JM, Kok FJ *et al.* (2008) Effect of fish oil on cognitive performance in older subjects: a randomized, controlled trial. *Neurology* **71**, 430–438.
 142. Stonehouse W, Conlon CA, Podd J *et al.* (2013) DHA supplementation improved both memory and reaction time in healthy young adults: a randomized controlled trial. *Am J Clin Nutr* **97**, 1134–1143.
 143. Yurko-Mauro K, McCarthy D, Rom D *et al.* (2010) Beneficial effects of docosahexaenoic acid on cognition in age-related cognitive decline. *Alzheimer's Dement* **6**, 456–464.
 144. Andrieu S, Guyonnet S, Coley N *et al.* (2017) Effect of long-term omega 3 polyunsaturated fatty acid supplementation with or without multidomain intervention on cognitive function in elderly adults with memory complaints (MAPT): a randomised, placebo-controlled trial. *Lancet Neurol* **16**, 377–389.
 145. Danthiir V, Hosking DE, Nettelbeck T *et al.* (2018) An 18-mo randomized, double-blind, placebo-controlled trial of DHA-rich fish oil to prevent age-related cognitive decline in cognitively normal older adults. *Am J Clin Nutr* **107**, 754–762.
 146. Dangour AD, Allen E, Elbourne D *et al.* (2010) Effect of 2-y *n*-3 long-chain polyunsaturated fatty acid supplementation on cognitive function in older people: a randomized, double-blind, controlled trial. *Am J Clin Nutr* **91**, 1725–1732.
 147. Soininen H, Solomon A, Visser PJ *et al.* (2017) 24-month Intervention with a specific multinutrient in people with prodromal Alzheimer's disease (LipiDiDiet): a randomised, double-blind, controlled trial. *Lancet Neurol* **16**, 965–975.
 148. Coley N, Raman R, Donohue MC *et al.* (2018) Defining the optimal target population for trials of polyunsaturated fatty acid supplementation using the erythrocyte omega-3 index: a step towards personalized prevention of cognitive decline? *J Nutr Health Aging* **22**, 982–998.
 149. Bazinet RP, Metherel AH, Chen CTS *et al.* (2020) Brain eicosapentaenoic acid metabolism as a lead for novel therapeutics in major depression. *Brain Behav Immun* **85**, 21–28.
 150. Patan MJ, Kennedy DO, Husberg C *et al.* (2021) Supplementation with oil rich in eicosapentaenoic acid, but not in docosahexaenoic acid, improves global cognitive function in healthy, young adults: results from randomized controlled trials. *Am J Clin Nutr* **114**, 914–924.
 151. Dyall SC (2015) Long-chain omega-3 fatty acids and the brain: a review of the independent and shared effects of EPA, DPA and DHA. *Front Aging Neurosci* **7**, 52.
 152. Patrick RP (2019) Role of phosphatidylcholine-DHA in preventing APOE4-associated Alzheimer's disease. *FASEB J* **33**, 1554–1564.
 153. Sperling RA, Aisen PS, Beckett LA *et al.* (2011) Toward defining the preclinical stages of Alzheimer's disease: recommendations from the national institute on aging-Alzheimer's association workgroups on diagnostic guidelines for Alzheimer's disease. *Alzheimer's Dement* **7**, 280–292.
 154. Umhau JC, Zhou W, Carson RE *et al.* (2009) Imaging incorporation of circulating docosahexaenoic acid into the human brain using positron emission tomography. *J Lipid Res* **50**, 1259–1268.
 155. Soininen H, Solomon A, Visser PJ *et al.* (2021) 36-month LipiDiDiet multinutrient clinical trial in prodromal Alzheimer's disease. *Alzheimer's Dement* **17**, 29–40.
 156. Zhang YP, Lou Y, Hu J *et al.* (2018) DHA supplementation improves cognitive function via enhancing A β -mediated autophagy in Chinese elderly with mild cognitive impairment: a randomised placebo-controlled trial. *J Neurol Neurosurg Psychiatry* **89**, 382–388.