Whole body and muscle energy metabolism in preruminant calves: effects of nutrient synchrony and physical activity

Joost J. G. C. van den Borne[1]*, Jean-François Hocquette[2], Martin W. A. Verstegen[1] and Walter J. J. Gerrits[1]

[1]*Animal Nutrition Group, Department of Animal Sciences, Wageningen University, P.O. Box 338, 6700 AH Wageningen, The Netherlands*

[2]*Unité de Recherches sur les Herbivores, INRA, Clermont-Ferrand/Theix, 63122 Saint-Genès-Champanelle, France*

The effects of asynchronous availability of amino acids and glucose on muscle composition and enzyme activities in skeletal muscle were studied in preruminant calves. It was hypothesized that decreased oxidative enzyme activities in muscle would explain a decreased whole body heat production with decreasing nutrient synchrony. Preruminant calves were assigned to one of six degrees of nutrient synchrony, step-wise separating the intake of protein and lactose over the two daily meals. Calves at the most synchronous treatment received two identical meals daily. At the most asynchronous treatment, 85 % of the daily protein and 20 % of the daily lactose supply were fed in one meal and the remainder in the other meal. Daily intakes of all dietary ingredients were identical for all treatments. Oxidative enzyme activities and fat content increased with decreasing nutrient synchrony in *M. Rectus Abdominis* (RA), but not in *M. Semitendinosus*. Cytochrome-*c*-oxidase activity was positively correlated with fat content in RA (r 0·49; $P<0·01$). Oxidative enzyme activities in both muscles were not correlated with average daily heat production, but citrate synthase activity in RA was positively correlated ($P<0·01$) with the circadian amplitude (r 0·53) and maximum (r 0·61) of heat production associated with physical activity. In conclusion, this study indicates that muscle energy stores are regulated by nutrient synchrony. The lack of correlation between muscle oxidative enzyme activities and average daily heat production was in contrast with findings in human subjects. Therefore, oxidative enzyme activity in muscle should not be used as an indicator for whole body heat production in growing calves.

Energy metabolism: Muscle: Nutrient synchrony: Physical activity

Daily nutrient intake has been correlated with muscle enzyme activities in animals (Cassar-Malek *et al.* 2004) and man (Helge & Kiens, 1997), and with intramuscular fat and glycogen contents in animals (Pethick & Rowe, 1996; Gondret *et al.* 2000). Regulation of muscle oxidative enzyme activities and muscle composition also depends on the supply of individual macronutrients (Hocquette *et al.* 1998; Geelen *et al.* 2001; Gondret & Lebret, 2002). Apart from variation in daily nutrient supply, within-day variation in the supply of different nutrients (i.e. synchrony) may induce these effects. Asynchronous absorption patterns can be induced by either a separated intake of protein and carbohydrates in time (e.g. dissociated diets) or by supplying ingredients with different kinetics of digestion and absorption. In preruminants, for example, asynchronous absorption of glucose and amino acids may result from differences in passage behaviour of clotting *v.* non-clotting dietary ingredients (Guilloteau *et al.* 1986; Verdonk *et al.* 1999). Applying a theoretical approach, we separated the intake of glucose and amino acids within the day (i.e. across meals) in heavy preruminant calves (Van den Borne *et al.*

2006*b*). Surprisingly, and in contrast to similar investigations in pigs (JJGC Van den Borne, JW Schrama, MJW Heetkamp, MWA Verstegen and WJJ Gerrits, unpublished results), increased separation of the amino acid and glucose availability within a day decreased whole body heat production (-9 %), and increased whole body fat deposition ($+59$ %), in heavy preruminant calves (Van den Borne *et al.* 2006*b*). Muscle energy metabolism may contribute to these changes as it accounts for about 20 % of the daily heat production in growing farm animals (Ortigues *et al.* 1995). Therefore, muscle energy metabolism may help to spare energy in case of an asynchronous supply of protein and lactose. Effects of nutrient synchrony on the energy metabolism and composition of muscles may also have implications for muscle function, meat quality and development of metabolic disorders in calves.

The regulation of muscle energy metabolism is well documented (e.g. Hocquette *et al.* 1998). In non-growing man, oxidative enzyme activities in muscle are positively correlated with whole body heat production (Zurlo *et al.* 1994; Doucet

et al. 2003). It is not known if a similar relationship exists in growing animals and if increased oxidative enzyme activities in muscle are associated with increased whole body heat production. In addition, it is unknown to what extent the demand for energy in muscle, i.e. for physical activity, drives the activity of the enzymes and hence regulates muscle energy metabolism in farm animals.

The aim of this study was therefore to gain insight in the regulation of whole body heat production by assessing the effects of nutrient synchrony and physical activity on muscle enzyme activities and muscle composition in heavy preruminant calves. Results on whole body protein and energy utilization have been presented elsewhere (Van den Borne *et al.* 2006*b*). It is hypothesized that oxidative enzyme activities decrease and intramuscular fat content increases with decreasing nutrient synchrony in preruminant calves. Oxidative enzyme activities in skeletal muscle were expected to correlate with whole body heat production.

Materials and methods

Animals, housing and experimental treatments

Thirty-six male, preruminant calves of the Holstein Friesian breed were used. The experimental treatments consisted of six degrees of nutrient synchrony (SYN 1 to 6) at each of two meal sequences (SEQ A and B) in a 6×2 factorial arrangement. The experiment was originally designed to study effects of SYN and SEQ on nutrient utilization (Van den Borne *et al.* 2006*b*). Two identical respiration chambers were available and calves were studied in eighteen trials of two calves each. Individual calves of approximately 130 kg body weight (BW) were assigned to an experimental treatment. The degree of nutrient synchrony was varied by a step-wise exchange of protein and lactose between the two daily meals (on a calculated digestible energy basis) which resulted in an equal distribution of digestible energy intake within a day (Table 1). Calves at SYN 1 received two identical meals daily. Calves at the most asynchronous treatment, SYN 6, received 85 % of the daily protein supply in one meal and the remaining 15 % in the other meal. A high protein meal was fed in the morning and a high lactose meal was fed in the evening for calves at SEQ A. For calves at SEQ B, this sequence was reversed. Allocation of calves to SYN and SEQ was balanced in time across trials.

Calves were fed the experimental diets for five weeks. During the first four weeks of study, calves were individually housed in cages (1.85×0.75 m) in a climate-controlled stable. Then they were moved to the respiration chambers where whole body heat production was measured during one week. The experiment complied with the Dutch law regarding the use of experimental animals.

Diets and feeding

Calves were fed according to their metabolic BW ($BW^{0.75}$), adjusted daily for a projected daily gain of 1000 g. Metabolizable energy requirements for maintenance (ME_m) were assumed to be 460 kJ/d per kg $BW^{0.75}$ (Van Es *et al.* 1967; Gerrits *et al.* 1996), and the feeding level was $2.0 \times ME_m$. Two basal milk replacers were produced for the most asynchronous treatment (SYN 6). All other experimental treatments were realized by mixing the basal milk replacers in the appropriate ratio to create the experimental contrast presented in Table 1. The detailed nutrient and ingredient composition of the basal milk replacers are given by Van den Borne *et al.* (2006*b*). Briefly, diets included whey protein concentrate, lactose and coconut and soya oil as main ingredients. Diets provided on a daily basis 20·5 % crude protein, 20·8 % crude fat and 44·9 % lactose. Daily intakes of all nutrients and dietary ingredients were similar between treatments. Milk replacer was reconstituted with water (140 g/l) and supplied at a temperature of about 40°C in a bucket. Roughage was not provided. Calves were fed individually at 06.00 h and 18.00 h, and were allowed 15 min to consume the meal. Feed intake was measured twice daily and BW was measured weekly throughout the experiment. Average daily gain and feed conversion ratio were calculated over the 5 week experimental period.

Measurement of whole body heat production and physical activity

During the fifth week of the study, gas exchange (O_2, CO_2 and CH_4) was continuously measured during 7 d in 6 min intervals by indirect calorimetry for estimation of whole body heat production. From the gaseous exchange, heat production (H_{tot}) was calculated according to the formula of Brouwer (1965) and the RQ was calculated as the CO_2 production divided by the O_2 consumption. Posture of calves was measured every minute by IR beam interruption. Physical activity was recorded with a radar–Doppler device according to the method described by Wenk & Van Es (1976). From these measurements, heat production associated with physical activity (H_{act}) and heat production corrected for physical activity (H_{cor}) were separately estimated (Van den Borne *et al.* 2006*a*). The kinetics of whole body heat production were described by calculation of the hourly means of H_{tot}, H_{cor} and H_{act} for each calf. The circadian maximum and minimum were defined for each calf as the highest and lowest hourly means within a day, respectively. The amplitudes of H_{tot}, H_{cor} and H_{act} were subsequently calculated as the difference between the circadian maximum and minimum for each

Table 1. Experimental treatments; division of the nutrient intake (g/kg $BW^{0.75}$) over two daily meals

Treatment (SYN)...	1	2	3	4	5	6
CON	50	57	64	71	78	85
Morning meal* (06.00 h)						
Protein	4·6	5·3	5·9	6·5	7·1	7·7
Lactose	10·0	8·9	7·8	6·7	5·6	4·5
Fat	4·6	4·6	4·6	4·6	4·6	4·6
Evening meal (18.00 h)						
Protein	4·6	4·0	3·4	2·8	2·2	1·6
Lactose	10·0	11·1	12·2	13·4	14·5	15·6
Fat	4·6	4·5	4·5	4·5	4·5	4·5

CON, contribution of the high protein meal to the daily protein supply (50 % = two identical, balanced meals; 100 % = all daily protein in one meal); SYN, degree of nutrient synchrony.

* Treatments are presented for meal sequence A. For meal sequence B, the nutrient intake in the morning and evening meal was reversed.

calf. For calculating circadian means of heat production by the formula of Brouwer (1965), the nitrogen excretion was assumed to be constant throughout the day.

Measurement of muscle composition and enzyme activities

After five weeks on the experimental treatment, calves were transported to the slaughterhouse (20 min) and killed at 13.00 h (i.e. 7 h after the morning meal) by stunning and exsanguination. Within 15 min post-slaughtering, samples of the *M. Rectus Abdominis* (RA; oxido-glycolytic muscle) and *M. Semitendinosus* (ST; glycolytic muscle) were removed. The samples of skeletal muscle were immediately trimmed of visible fat and connective tissue. Samples were then cut into pieces, immediately frozen in liquid N_2 and stored at $-80°C$ pending analyses. Samples for crude fat analysis (about 30 g for each muscle) were not frozen in liquid N_2, but ice-chilled and subsequently stored at $-20°C$ pending analyses.

Protein (Bradford, 1976) and DNA (Labarca & Paigen, 1980) contents were measured in muscle homogenates. Activity of the oxidative enzyme citrate synthase (CS; EC 4.1.3.7), which is involved in the substrate flux through the tricarboxylic acid cycle, was determined in sonicated homogenates by measuring the rate of initial reaction at 412 nm by means of the DTNB (5·5′-dithiobis(2-nitrobenzoate)) method as described by Shepherd & Garland (1969). Activity of the oxidative enzyme cytochrome-*c*-oxidase (COX; EC 1.9.3.1), which is involved in the substrate flux through the respiratory chain, was determined in sonicated homogenates by measuring the oxidation of reduced cytochrome-*c* as described by Van Hinsberg *et al.* (1978). Activities of CS and COX were expressed in μmol CoA liberated per min per g protein and in μmol cytochrome-*c* oxidized per min per g protein at 25°C respectively. Activity of the glycolytic enzyme lactate dehydrogenase (LDH; EC 1.1.1.2.7), which catalyzes the formation of lactate from pyruvate, was determined as described by Ansay (1974). LDH activity was expressed in μmol NADH oxidized per min per g protein. Glycogen content was measured by the method of Carroll *et al.* (1955). Briefly, glycogen was extracted from the tissue by homogenization with 5 % trichloroacetic acid solution, precipitated from the extract by 95 % ethanol and determined with the anthrone reagent in a colorimeter at 620 nm. Fat content was measured in muscle tissue after freeze-drying according to ISO 6492 (International Organization for Standardization, 1999). Ratios for CS:LDH, COX:LDH (relative enzyme activities), protein:DNA (indicator for muscle fibre size) and COX:CS (indicator for the biochemical properties of the mitochondria) were calculated.

Statistical analysis

The effects of nutrient synchrony and the interaction between nutrient synchrony and meal sequence on muscle traits and whole body energy metabolism traits were analysed by linear regression analysis, using the general linear model procedure of the SAS statistical software package version 9.1 (Statistical Analysis Systems Institute, Cary, NC, USA). The degree of nutrient synchrony was included as a regressor

(Default 1).

$$Y_{ij} = \mu + \beta_1 \times X_j + \beta_{2i} \times S_i X_j + \varepsilon_{ij}, \tag{1}$$

where $Y_{ij} =$ dependent variable over the whole period, $\mu =$ average intercept, $\beta_1 =$ effect of degree of nutrient synchrony, expressed as percentage of the daily protein intake in the high protein meal, $\beta_{2i} =$ interaction between degree of nutrient synchrony and meal sequence i, $S_i =$ fixed effect of meal sequence i, $X_j =$ degree of nutrient synchrony (expressed as percentage of the daily protein intake in the high protein meal) for calf j, $\varepsilon_{ij} =$ error term, $i = 1, 2$, and $j = 1, \ldots, 18$.

Pearson correlation coefficients were calculated for relationships between muscle energy metabolism traits within each muscle, for each trait between the two muscles and between muscle energy metabolism traits and whole body energy metabolism traits. Traits for the two muscles were compared by pairwise comparisons.

Results

Animal performance

Two calves were excluded from analysis, because of feed refusals or illness. The initial BW of the calves was 128 (SEM 1·7) kg and average daily gain during the 5 week experimental period was 885 (SEM 27·1) g. Average daily gain was lower ($P < 0·05$) for calves at SYN 5 and 6 (729 and 815 g respectively) than for calves at SYN 1–4. Calves at SYN 1–4 had a similar daily gain (on average 945 g). Feed intake did not differ between treatments and consequently the feed to gain ratio was higher ($P < 0·01$) for calves at SYN 5 and 6 (2·56 and 2·48 respectively) than for calves at the other treatments (on average 2·02). However, calves at the two most asynchronous treatments had a lower nutrient digestibility. The consequently lower digestible nutrient intake in calves at SYN 5 and 6 complicated interpretation of the results from SYN 1 to 6. Linear regression was therefore also performed for SYN 1 to 4 (*n* 22) separately, because calves at those treatments had identical digestible nutrient intakes.

Muscle enzyme activities

In RA, activities of COX and CS increased ($P = 0·048$) and tended to increase ($P = 0·075$), respectively, with decreasing nutrient synchrony (SYN 1–4; Table 2). Ratios of enzyme activities were not affected by nutrient synchrony. In ST, enzyme activities were not affected by nutrient synchrony (Table 3). The sequence of the high protein and high carbohydrate meals did not affect enzyme activities.

Muscle composition

Intramuscular fat content increased in RA ($P = 0·037$) with decreasing nutrient synchrony at an identical digestible nutrient intake (Table 2), but nutrient synchrony did not affect intramuscular fat content in ST (Table 3). The fat content was substantially higher ($P < 0·001$) in RA (18·9 mg/g tissue) than in ST (6·5 mg/g tissue). The sequence of the high protein and high carbohydrate meals did not affect intramuscular fat content.

Table 2. Effects of nutrient synchrony on muscle enzyme activities and muscle composition in *M. Rectus Abdominis* of heavy preruminant calves

(Values are means for treatments with their standard errors; *n* 5 per treatment for SYN 1 and 2, and *n* 6 per treatment for SYN 3−6)

Treatment (SYN)...	1	2	3	4	5	6	SYN 1−6			SYN 1−4		
CON	50	57	64	71	78	85	SEM	b^*	P value†	SEM	b	P value
Enzyme activity (μmol/g protein per min)												
LDH	4317	4269	4502	4447	4358	4076	286·2	-4.5×10^{-3}	0·673	344·4	8.7×10^{-3}	0·710
COX	119	97	119	154	121	99	13·7	0.2×10^{-4}	0·960	14·1	2.0×10^{-3}	0·048
CS	54	52	57	67	58	51	5·3	0.4×10^{-4}	0·850	5·0	0.7×10^{-3}	0·075
Relative enzyme activity												
COX:CS	2·22	1·87	2·16	2·70	2·33	2·00	0·321	2.3×10^{-3}	0·849	0·299	0·01	0·499
COX:LDH, $\times 10^{-3}$	27·8	24·6	27·7	35·3	28·4	24·2	3·92	2.1×10^{-6}	0·989	4·32	0.4×10^{-3}	0·191
CS:LDH, $\times 10^{-3}$	12·4	12·8	12·9	15·9	13·8	12·3	1·53	0.2×10^{-4}	0·728	1·48	0.2×10^{-3}	0·180
Muscle composition												
Glycogen(mg/g fresh tissue)‡	5·9	7·6	6·2	6·0	5·5	6·1	1·14	-0.02	0·630	1·25	-9.0×10^{-3}	0·907
Fat (mg/g fresh tissue)	16·3	13·5	21·9	23·9	16·3	13·7	2·46	-0.03	0·802	2·43	0·44	0·037
Protein (mg/g fresh tissue)	165	175	170	158	162	172	7·0	-0.07	0·764	7·7	-0.45	0·391
DNA (μg/g fresh tissue)	1881	1972	2017	1996	1977	1989	55·1	1·95	0·309	47·9	5·75	0·088
Protein:DNA (mg/mg)	88·1	88·8	84·8	79·4	82·3	87·0	4·39	-0.1×10^{-3}	0·435	4·92	-0.5×10^{-3}	0·140

CON, contribution of the high protein meal to the daily protein supply (50 % = two identical, balanced meals; 100 % = all daily protein in one meal); SYN, degree of nutrient synchrony; LDH, lactate dehydrogenase; COX, cytochrome-*c*-oxidase; CS, citrate synthase.
* Regression coefficient ($y = a + b \times x$), representing the change in response parameter per % increase of the contribution of the high protein meal to the daily protein supply.
† Probability for test if the regression coefficient (*b*) equals 0. Effects of meal sequence and interactions between meal sequence and degree of nutrient synchrony were non-significant and were therefore excluded from the model.
‡ An interaction between CON and the meal sequence on muscle glycogen content was found ($P = 0.023$).

Table 3. Effects of nutrient synchrony on muscle enzyme activities and muscle composition in *M. Semitendinosus* of heavy preruminant calves

(Values are means for treatments with their standard errors; *n* 5 per treatment for SYN 1 and 2, and *n* 6 per treatment for SYN 3−6)

Treatment (SYN)...	1	2	3	4	5	6	SYN 1−6			SYN 1−4		
CON	50	57	64	71	78	85	SEM	b^*	P value†	SEM	b	P value
Enzyme activity (μmol/g protein per min)												
LDH	4167	4618	4500	4738	4381	4562	242·2	5.2×10^{-3}	0·543	272·9	0.2×10^{-4}	0·240
COX	113	94	78	103	90	96	12·7	-0.2×10^{-3}	0·544	12·8	-0.5×10^{-3}	0·525
CS	46	53	43	61	54	58	4·9	0.3×10^{-3}	0·095	5·4	0.5×10^{-3}	0·166
Relative enzyme activity												
COX:CS	2·51	1·80	2·04	1·72	1·68	1·62	0·264	-0.02	0·051	0·326	-0.03	0·199
COX:LDH, $\times 10^{-3}$	29·4	21·2	17·8	22·0	20·9	21·6	3·91	-0.1×10^{-3}	0·347	4·14	-0.3×10^{-3}	0·267
CS:LDH, $\times 10^{-3}$	11·5	11·8	9·7	13·1	12·4	12·8	1·37	0.1×10^{-3}	0·311	1·53	0.6×10^{-4}	0·533
Muscle composition												
Glycogen (mg/g fresh tissue)‡	6·7	6·6	8·2	6·4	6·8	6·8	1·06	-1.4×10^{-3}	0·969	1·14	-0.3×10^{-3}	0·881
Fat (mg/g fresh tissue)	5·6	5·4	7·4	7·6	6·1	6·0	0·98	0·01	0·717	1·19	0·12	0·166
Protein (mg/g fresh tissue)	177	171	171	166	172	175	5·5	-0.04	0·841	6·9	-0.42	0·283
DNA (μg/g fresh tissue)	1964	1880	1908	1950	1944	1844	80·8	-1.57	0·594	84·9	-0.74	0·889
Protein:DNA (mg/mg)	92·7	93·1	89·6	85·9	90·2	94·8	5·69	0.1×10^{-4}	0·961	6·62	0·01	0·475

CON, contribution of the high protein meal to the daily protein supply (50 % = two identical, balanced meals; 100 % = all daily protein in one meal); SYN, degree of nutrient synchrony; LDH, lactate dehydrogenase; COX, cytochrome-*c*-oxidase; CS, citrate synthase.
* Regression coefficient ($y = a + b \times x$), representing the change in response parameter per % increase of the contribution of the high protein meal to the daily protein supply.
† Probability for test if the regression coefficient (*b*) equals 0. Effects of meal sequence and interactions between meal sequence and degree of nutrient synchrony were non-significant and were therefore excluded from the model.
‡ An interaction between CON and the meal sequence on muscle glycogen content was found ($P = 0.007$).

Glycogen content in both muscles was not affected by nutrient synchrony (Tables 2 and 3), but a positive correlation between glycogen content in RA and ST was found (r 0·72; $P<0·001$). Glycogen content averaged 6·24 mg/g in RA and 7·01 mg/g in ST. An interaction between SEQ and SYN was found for intramuscular glycogen content in both muscles ($P<0·05$), indicating that glycogen content was higher after feeding the high protein diet than after the high lactose diet. The difference in glycogen content between meal sequences was more pronounced at SYN 2 (3·08 mg/g in RA and 6·08 mg/g in ST) and SYN 3 (4·63 mg/g in RA and 4·96 mg/g in ST) than at the other treatments.

Whole body heat production

Average heat production and H_{cor} decreased with decreasing nutrient synchrony ($P<0·05$), while the RQ increased ($P=0·028$; Table 4). A detailed description of whole body energy partitioning and protein deposition is presented and discussed elsewhere (Van den Borne et $al.$ 2006b). The maxima of the circadian patterns of H_{tot} and H_{cor} decreased with decreasing nutrient synchrony ($P<0·05$), but the maximum of the circadian pattern of H_{act} was not affected by nutrient synchrony. The amplitudes of the circadian patterns of H_{tot}, H_{cor} and H_{act} were not affected by SYN. The sequence of the high protein and high carbohydrate meals did not affect any of the traits of whole body energy metabolism.

Correlations within muscle

In both muscles, CS and COX activities were positively correlated (r 0·35–0·36; $P<0·05$). Intramuscular fat content showed a positive correlation (r 0·49; $P<0·01$) with COX (and COX:LDH) in RA (data not shown).

Correlations between muscle energy metabolism and whole body energy metabolism

The decreased H_{tot} and H_{cor} with decreasing nutrient synchrony did not correspond with the increased (or unaffected) oxidative enzyme activities in muscle. In accordance, simple correlation analysis showed that average H_{tot}, H_{cor} and H_{act} were not correlated with enzyme activities in RA (Table 5) and ST (data not shown). Intramuscular fat content in RA was negatively correlated with average H_{tot} and H_{cor}.

The maximum of H_{act} and the amplitude of H_{tot} and H_{act} increased with increasing CS activity in RA in preruminant calves, resulting in positive correlations (Table 5). In ST, oxidative enzyme activities were not related to the maxima and amplitudes of heat production traits (data not shown).

Discussion

Animal performance

Feeding more than 71 % of the daily protein in one meal and more than 68 % of the daily lactose in the other one (i.e. SYN 5–6) induced diarrhoea, which was indicated by a lower faecal dry matter content, a lower faecal pH and a lower nutrient digestibility (Van den Borne et $al.$ 2006b). The lower daily gain and higher feed to gain ratio for calves at SYN 5 and 6

were therefore likely due to a decreased digestible nutrient intake. Comparison of calves at SYN 1–4 allows studying the effects of nutrient synchrony at identical intakes of digestible nutrients. Therefore, this discussion focuses on treatments SYN 1–4.

Effects of nutrient synchrony on muscle enzyme activities

Although the quantity (Brandstetter et $al.$ 1998; Cassar-Malek et $al.$ 2004) and the composition (Helge & Kiens, 1997; Geelen et $al.$ 2001; Cuvelier et $al.$ 2006) of the daily feed supply are known to affect oxidative and glycolytic enzyme activities in muscles, this study is one of the first to describe the effect of within-day distribution of nutrient availability on muscle metabolism. Ortigues-Marty et $al.$ (2003) found increased oxidative enzyme activities in muscle (ST) of preruminant calves when skimmed milk protein was replaced by soluble wheat and whey proteins. Feeding soluble wheat and whey proteins resulted in larger circadian fluctuations of the amino acid availability compared with skimmed milk protein. Therefore, it was suggested that a transiently high amino acid availability may require an increased capacity of oxidative enzymes in muscle to provide energy for protein synthesis when amino acids are available (Ortigues-Marty et $al.$ 2003). However, potential reasons for the affected oxidative capacity in the study of Ortigues-Marty et $al.$ (2003) also include a lower intake of the skimmed milk diet than of the wheat and whey protein diet and the use of different dietary ingredients with different kinetics of fatty acid absorption (Petit et $al.$ 1988; Cruywagen et $al.$ 1990; Ortigues-Marty et $al.$ 2003).

A decreased nutrient synchrony (i.e. increased circadian fluctuations in nutrient supply) also increased oxidative enzyme activities in RA in the current study (Table 2), but not in ST. Separating the availability of amino acids and glucose may have required temporarily high oxidative enzyme activities to provide energy for protein synthesis (see Ortigues-Marty et $al.$ 2003) or to oxidize excessively available amino acids or glucose. As net protein utilization was unaffected and fat deposition was increased (Van den Borne et $al.$ 2006b), temporarily increased rates of glucose and/or amino acid oxidation should be accompanied by decreased oxidation rates of oxidation during the remainder of the day.

It is noteworthy that nutrient synchrony only affected muscle enzyme activities in an oxidative muscle (RA), but not in a glycolytic muscle (ST). This corresponds with previous studies which demonstrated a higher response of oxidative (mitochondria-rich) than glycolytic muscles to nutrient supply (Cassar-Malek et $al.$ 2004; Jurie et $al.$ 2006).

Effects of nutrient synchrony on muscle composition

Intramuscular fat content was almost threefold higher in RA than in ST, which agrees with the generally described higher fat content in oxidative than in glycolytic muscles (Gondret et $al.$ 1998; Hocquette et $al.$ 2003). The increased intramuscular fat content in RA with decreasing nutrient synchrony corresponds to an increased whole body fat deposition in these calves (Van den Borne et $al.$ 2006b) indicating that the extra fat is at least partly deposited in muscles. Intramuscular fat generally develops after abdominal, intermuscular and

Table 4. Effects of nutrient synchrony on whole body energy metabolism (kJ/d per kg $BW^{0.75}$) in heavy preruminant calves

(Values are means for treatments with their standard errors; n 5 per treatment for SYN 1 and 2, and n 6 per treatment for SYN 3–6)

Treatment (SYN)...	1	2	3	4	5	6	SYN 1–6			SYN 1–4		
CON	50	57	64	71	78	85	b*	SEM	P value†	b	SEM	P value
Average												
Heat production	691	677	666	629	652	631	−1.66	6.9	0.003	−2.80	8.9	0.012
Activity related heat production	72	79	67	61	66	69	−0.23	2.7	0.328	−0.64	3.4	0.151
Activity corrected heat production	619	598	599	567	586	562	−1.43	5.9	0.003	−2.16	7.8	0.024
Respiratory quotient	0.85	0.86	0.87	0.87	0.86	0.85	-0.02×10^{-3}	0.002	0.892	0.71×10^{-3}	0.003	0.028
Maximum												
Heat production	815	801	786	749	776	743	−1.90	8.6	0.007	−3.03	10.9	0.027
Activity related heat production	158	174	153	156	141	157	−0.38	5.6	0.410	−0.40	6.8	0.670
Activity corrected heat production	721	688	680	642	680	619	−2.31	9.1	0.002	−3.46	11.0	0.011
Respiratory quotient	0.89	0.90	0.91	0.92	0.91	0.92	0.54×10^{-3}	0.003	0.036	1.09×10^{-3}	0.003	0.006
Amplitude (Δ)												
Heat production	242	228	227	226	236	204	−0.67	5.8	0.177	−0.66	7.5	0.511
Activity related heat production	138	151	135	142	120	130	−0.51	5.3	0.244	0.05	6.4	0.956
Activity corrected heat production	187	167	165	164	176	131	−1.04	4.7	0.007	−1.01	4.7	0.101
Respiratory quotient	0.10	0.10	0.09	0.11	0.11	0.12	0.75×10^{-3}	0.008	0.005	0.42×10^{-3}	0.008	0.304

CON, contribution of the high protein meal to the daily protein supply (50 % = two identical, balanced meals; 100 % = all daily protein in one meal); SYN, degree of nutrient synchrony.

* Regression coefficient ($y = a + b \times x$), representing the change in response parameter per % increase of the contribution of the high protein meal to the daily protein supply.

† Probability for test if the regression coefficient (b) equals 0. Effects of meal sequence and interactions between meal sequence and degree of nutrient synchrony were non-significant and were therefore excluded from the model.

subcutaneous fat deposition (Vernon, 1981), which suggests that nutritional regulation of partitioning of fat into muscle is unlikely in relatively young animals (approximately 18 weeks of age). On the other hand, fat deposition in muscle may have been selectively stimulated by the increased glucose supply after a high lactose meal. Smith & Crouse (1984) showed that glucose provides only 1–10 % of the acetyl units to in vitro lipogenesis in subcutaneous adipose tissue of steers, but 50–75 % of the acetyl units in the intramuscular adipocytes. This may be true for preruminants as well, thus providing an explanation for the effect of nutrient synchrony on intramuscular fat content of RA. However, care should be taken in extrapolating these results to the in vivo situation of preruminants, as they absorb much larger amounts of long-chain fatty acids and monosaccharides (glucose and galactose) than mature ruminants.

The intramuscular glycogen content was higher in both muscles after the high protein diet than after the high lactose diet, but interpretation of this interaction between SYN and SEQ on glycogen content is complicated by its non-linearity. The lower glycogen content when protein and carbohydrate intakes are further separated may be explained by a lower insulin response in more asynchronously fed calves (T Vicari, JJGC Van den Borne, WJJ Gerrits, Y Zbinden and JW Blum, unpublished results). This corresponds with human studies in which the insulin response as well as muscle glycogen accumulation was higher after a mixed protein and carbohydrate meal than after a carbohydrate meal (Zawadzki et al. 1992; Van Loon et al. 2000), but does not explain the low muscle glycogen content at the most synchronous treatment treatments.

Correlations within muscle

Cytochrome-c-oxidase activity was positively correlated with muscle composition in RA (i.e. intramuscular fat) in the current study. This relationship between an oxidative enzyme and muscle fat content in RA seems paradoxical, but similar results were obtained in growing rabbits (Gondret et al. 2004) and steers (Hocquette et al. 2003). A high fat deposition and a high fatty acid oxidation can go together as the result of a high fatty acid flux. Although the fat intake with the two daily meals did not differ between treatments, the degree of nutrient synchrony may have (temporarily) increased the fatty acid flux by affecting the kinetics of fat digestion and/or the partitioning of fatty acids to individual tissues.

Correlations between muscle enzyme activities and whole body energy metabolism

In human subjects, positive correlations between muscle enzyme activities and 24 h heat production or sleeping metabolic rate were found (Zurlo et al. 1994; Doucet et al. 2003). In the present study, however, average whole body heat production was not correlated with enzyme activities in RA and ST. Our initial hypothesis that the decreased whole body heat production with decreasing nutrient synchrony is associated with decreased muscle oxidative enzyme activities therefore has to be rejected. Oxidative enzyme activity in RA

Table 5. Pearson correlation coefficients between oxidative enzyme activities and muscle composition in *M. Rectus Abdominis* and whole body energy metabolism traits in heavy preruminant calves (*n* 22)

Trait	CS	COX	CS:LDH	COX:LDH	Fat	Glycogen
Average						
Heat production	−0·06	−0·33	−0·22	−0·42	−0·49*	−0·06
Activity related heat production	0·04	−0·41	0·06	−0·29	−0·43*	0·09
Activity corrected heat production	−0·09	−0·20	−0·28	−0·36	−0·37	−0·11
Maximum						
Heat production	0·28	−0·29	0·19	−0·27	−0·35	0·08
Activity related heat production	0·53**	−0·12	0·58**	−0·10	−0·22	0·26
Activity corrected heat production	−0·12	−0·26	−0·27	−0·39	−0·44*	−0·14
Amplitude						
Heat production	0·63**	0·08	0·66***	0·10	−0·08	0·16
Activity related heat production	0·61**	−0·14	0·58**	−0·11	−0·20	0·19
Activity corrected heat production	−0·04	−0·14	−0·05	−0·14	−0·21	−0·26

LDH, lactate dehydrogenase; COX, cytochrome-*c*-oxidase; CS, citrate synthase.
* $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$.

was positively correlated with the circadian maximum and amplitude of H_{act} in growing calves. A similar correlation was found with H_{tot}, which can be explained by the high contribution of variation in H_{act} to variation in H_{tot}.

The high feeding level in preruminant calves may explain why oxidative enzyme activity in muscle showed a better relationship with within-day variation in H_{act} than with average H_{tot}. Apart from muscle, several other tissues contribute to whole body heat production. Feeding large amounts of nutrients results in a high diet-induced heat production predominantly caused by portal drained viscera and liver (Ortigues *et al.* 1995). This may also explain the different results in growing calves and non-growing man. The diet-induced heat production usually contributes for 30 to 35 % to H_{tot} in growing farm animals (Collin *et al.* 2001; Le Bellego *et al.* 2001; Van Milgen *et al.* 2001), but only for 5 to 15 % to H_{tot} in adult human subjects (Westerterp, 2004). Consequently, the overall contribution of H_{act} and muscle energy metabolism to average H_{tot} is likely to be higher in man than in calves. The individual housing conditions in the current study restricted the physical activity of calves, whereas calves under practical circumstances are housed in groups. Although group-housing only marginally increased H_{act} compared with individual housing as shown in pigs (Rijnen, 2003), this may have contributed to the lack of correlation between oxidative enzyme activity and H_{tot} in this study.

Oxidative enzyme activities in muscle were not correlated with the average H_{act} but only with the circadian fluctuations in H_{act} (maximum and amplitude) in preruminant calves. This may be explained by the measurement of enzyme activity, which truly reflects the maximum enzyme activity rather than the enzyme activity *in vivo*. Variation in fatty acid oxidation, for example, is not necessarily associated with variation in oxidative enzyme activities in muscle (Piot *et al.* 1998). Enzyme activities depend not only on the capacity of enzymes to convert substrates into new (co or end) products, but also on the availability of substrates. The availability of substrates for oxidative enzymes within a day is affected by nutrient supply, but also by particular energy requiring processes like physical activity. It can therefore be expected that the maximum oxidative enzyme activity

is regulated by the kinetics of substrate availability rather than by the daily average substrate availability. Although the major aim of this study was to determine effects of variation in nutrient supply on muscle energy metabolism, it appeared that variation in H_{act} was positively correlated with CS activity in RA.

The positive correlation between physical activity and CS activity in muscle corresponds with results obtained in man (Rimbert *et al.* 2004), rats (Spangenberg *et al.* 2005), pigs (Petersen *et al.* 1997) and steers (Jurie *et al.* 2006). Similarly, muscle CS activity was 23 % higher in group-housed calves than in individually housed calves (Ortigues-Marty *et al.* 2003) which likely relates to increased physical activity. Previous studies have, however, not related the muscle enzyme activity to *in vivo* energy expenditure. Therefore, the current study is one of the first to relate CS activity in muscle to heat production associated with physical activity (and hence to contractile activity in muscle tissue).

Conclusions

From the results of the present study it was concluded that a decreased nutrient synchrony increased oxidative enzyme activity and intramuscular fat content in the oxidative muscle RA of preruminant calves. This indicates that the within-day distribution of macronutrient availability can affect muscle composition and properties which may have consequences for muscle function and meat quality. However, the decrease in whole body heat production with increasing separation of protein and lactose intake could not be explained by a decrease in oxidative enzyme activities in muscle.

In human subjects, muscle enzyme activities relate to whole body energy expenditure. Consequently, enzyme activities can be used as predictors for energy expenditure. Our data indicate that this relationship is present only for the activity associated component of energy expenditure in growing preruminant calves. Therefore, this relationship loses significance when subjects are fed at higher intake levels. Consequently, muscle enzyme activities are probably useless as an indicator for whole body energy expenditure in rapidly growing animals.

Acknowledgements

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