

5. Kado C I, Liu ST. Rapid procedure for detection and isolation of large and small plasmids. *J Bacteriol* 1981;145:1365–1373.
6. Ho PL, Lo WU, Yeung MK, et al. Complete sequencing of pNDM-HK encoding NDM-1 carbapenemase from a multidrug-resistant *Escherichia coli* strain isolated in Hong Kong. *PLoS One* 2011;6:e17989.

Menacing Emergence of Fosfomycin Resistance Among *Klebsiella pneumoniae* Carbapenemase–2-Producing *K. pneumoniae* Driven by Prior Use in Critically Ill Patients

To the Editor—Owing to the widespread prevalence of carbapenemase-producing Enterobacteriaceae resistant to last-resource therapeutic options, including extended-spectrum β -lactams, fluoroquinolones, and aminoglycosides, an interest in old antimicrobial agents, such as polymyxins and fosfomycin, has reignited.¹ The latter is an agent that acts inhibiting the formation of a precursor of peptidoglycan (ie, a cell wall-acting agent); it was first used in the treatment of uncomplicated urinary tract infections but, nowadays, is being used (still on a small scale in our institution) as an adjunct to other active agents for the treatment of *Klebsiella pneumoniae* carbapenemase (KPC)–2-producing *K. pneumoniae* (KPC-2-Kp) infections.²

Although high in vitro frequency of fosfomycin resistance mutations has been reported,³ resistance rates to this agent have remained relatively low since its introduction in clinical practice. On the other hand, a substantially higher resistance rate has been noted when carbapenemase producers are considered.^{3,4} Furthermore, a report by Karageorgopoulos et al⁵ identified patients who were treated with fosfomycin for an initially fosfomycin-susceptible KPC-2-Kp bacteremia but from whom a fosfomycin-resistant isolate was subsequently

collected. However, the potential of in vivo emergence of fosfomycin-resistance among KPC-2-Kp isolates has not been systematically investigated so far.

Thus, this study aimed to perform a survey on the subsequent emergence of fosfomycin resistance among intensive care unit patients from whom a fosfomycin-susceptible KPC-2-Kp isolate was previously collected at a tertiary hospital in southern Brazil from April 1, 2013, through May 31, 2015. KPC-2-Kp was defined according to carbapenem resistance patterns, with phenotypic testing results determined as proposed by Clinical and Laboratory Standards Institute guidelines⁶ and through *bla*_{KPC-2} gene detection by polymerase chain reaction as previously reported.⁷ Cases were defined as patients from whom a fosfomycin-resistant isolate was recovered from urine and/or blood cultures more than 48 hours but less than 90 days after the day a urinary fosfomycin-susceptible isolate was collected. Data on antibiotic exposures between the first KPC-2-Kp isolation and the day on which a positive culture for a fosfomycin-resistant isolate was obtained were recorded.

Eighty-five patients had a urinary KPC-2-Kp isolate collected during the period of this study and 35 of them (41.2%; 95% CI, 31.3%–51.8%) had a subsequent isolation of this same pathogen: 20 patients with a recurrent bacteriuria, 10 patients presenting a bloodstream infection, and 5 patients with both. Each patient was considered only once as a case and therefore for those patients in whom a KPC-2-Kp was recovered from a recurrent bacteriuria and blood, only the latter was considered. Among these 35 patients, in 32 (91.4%) a fosfomycin-susceptible KPC-2-Kp isolate had been previously recovered. For the 3 patients presenting a fosfomycin-resistant KPC-2-Kp isolate, the subsequent fosfomycin susceptibility remained unaltered. On the other hand, for those 32 patients with a prior fosfomycin-susceptible isolate, in 8 patients (25%) the subsequent KPC-2-Kp isolate was resistant to fosfomycin. When evaluating the previous use of antibiotics, 5 of these 8 patients (62.5%; odds ratio, 9.6 [95% CI, 1.6–56.9], *P* = .013) had already received fosfomycin to treat the first urinary KPC-2-Kp isolate (Table 1).

TABLE 1. Microbiologic Features and Patients' Data in Study of Emergence of Fosfomycin Resistance

Patients	Fosfomycin Etest MIC, mg/L ^a		Antibiotic used to treat the first isolate	Clinical site
	First ^b	Subsequent ^c		
1	12.0	128	None	Urine
2	6.0	128	Polymyxin B / meropenem	Urine
3	32.0	>1024	Fosfomycin	Urine and blood
4	32.0	>1024	Fosfomycin	Urine
5	16.0	1024	Fosfomycin	Urine
6	32.0	>1024	Polymyxin B / meropenem / ertapenem	Urine and blood
7	32.0	>1024	Fosfomycin	Urine
8	32.0	>1024	Fosfomycin	Urine

NOTE. All isolates were *Klebsiella pneumoniae* with carbapenem resistance via *bla*_{KPC-2}. MIC, minimum inhibitory concentration.

^aConsidering ≤ 64 mg/L and >64 mg/L as susceptible and resistant, respectively.

^bFirst urinary *K. pneumoniae* carbapenemase (KPC)–2-producing *K. pneumoniae* (KPC-2-Kp) isolate.

^cSubsequent KPC-2-Kp isolate, considering a hospitalization period of >48 hours and <90 days following first isolation.

Although fosfomycin used to be primarily designated for urinary tract infection treatments, the lack of available antibiotics to treat carbapenemase producers has given fosfomycin an important adjuvant role, mainly in more severe infection cases. Despite that, according to results reported by Karageorgopoulos et al⁵ as well as this present study where the emergence of fosfomycin resistance was reported just shortly after its introduction in clinical practices (mid-2014), fosfomycin resistance has become a concern because the endemic level reached by the KPC-2-Kp is due to its great ability to adapt and survive,^{8,9} characteristics that came as an advantage mainly through antimicrobial selective pressure, strongly driven by the previous use, showing the need to establish a rigorous protocol for antimicrobial consumption.

The limitation of this study is due to the unknown genetic background information on which mechanism is involved to confer resistance to fosfomycin. So, further studies should be performed in order to detect possible genetic targets, such as *fosA3* gene, that encode for a specific enzyme and which have recently resulted in a high resistance level to fosfomycin among European KPC-producers.¹⁰

In conclusion, this study reports a significant emergence of fosfomycin resistance among KPC-2-Kp isolates in a relatively short period after the introduction of this antibiotic as an effective agent to treat KPC infections. Strict control practices are urgently required in order to avoid the resistance rate increase, regardless of the mechanism by which it occurs.

ACKNOWLEDGMENTS

Financial support. Conselho Nacional de Desenvolvimento Científico e Tecnológico, Brazil.

Potential conflicts of interest. The author reports no conflicts of interest relevant to this article.

Leandro Reus Rodrigues Perez, PhD

Affiliations: Microbiologia, Hospital Mãe de Deus, Porto Alegre, Brazil; and Laboratório de Pesquisa em Resistência Bacteriana, Centro de Pesquisa Experimental, Hospital de Clínicas de Porto Alegre, Porto Alegre, Brazil.

Address correspondence to Leandro Reus Rodrigues Perez, PhD, Laboratório de Pesquisa em Resistência Bacteriana, Centro de Pesquisa Experimental, Hospital de Clínicas de Porto Alegre, Ramiro Barcelos Street, 2350, Porto Alegre, RS, Brazil 90.035-003 (leandro.reus@gmail.com).

Infect Control Hosp Epidemiol 2016;37:748–749

© 2016 by The Society for Healthcare Epidemiology of America. All rights reserved. 0899-823X/2016/3706-0031. DOI: 10.1017/ice.2016.68

REFERENCES

1. Giske CG. Contemporary resistance trends and mechanisms for the old antibiotics colistin, temocillin, fosfomycin, mecillinam and nitrofurantoin. *Clin Microbiol Infect* 2015;21:899–905.
2. Michalopoulos A, Virtzili S, Rafailidis P, et al. Intravenous fosfomycin for the treatment of nosocomial infections caused by carbapenem-resistant *Klebsiella pneumoniae* in critically ill patients: a prospective evaluation. *Clin Microbiol Infect* 2010;16:184–186.

3. Karageorgopoulos DE, Wang R, Yu XH, Falagas ME. Fosfomycin: evaluation of the published evidence on the emergence of antimicrobial resistance in gram-negative pathogens. *J Antimicrob Chemother* 2012;67:255–268.
4. Jiang Y, Shen P, Wei Z, et al. Dissemination of a clone carrying a *fosA3*-harbouring plasmid mediates high fosfomycin resistance rate of KPC-producing *Klebsiella pneumoniae* in China. *Int J Antimicrob Agents* 2015;45:66–70.
5. Karageorgopoulos DE, Miriagou V, Tzouveleki LS, Spyridopoulou K, Daikos GL. Emergence of resistance to fosfomycin used as adjunct therapy in KPC *Klebsiella pneumoniae* bacteraemia: report of three cases. *J Antimicrob Chemother* 2012;67:2777–2779.
6. Clinical and Laboratory Standards Institute (CLSI). Performance standards for antimicrobial susceptibility testing: 25th informational supplement. CLSI document. Wayne, PA: CLSI; 2015: M100-S25.
7. Perez LR, Rodrigues D, Dias CG. Evaluation of phenotypic tests to detect carbapenem-resistant Enterobacteriaceae in colonized patients hospitalized in intensive care units. *Braz J Infect Dis* 2015;19:436–438.
8. Perez LR. Carbapenem-resistant Enterobacteriaceae: a major prevalence difference due to the high performance of carbapenemase producers when compared to the nonproducers. *Infect Control Hosp Epidemiol* 2015;36:1480–1482.
9. Perez LR, Dias CG. Emergence of infections due to a polymyxin B-resistant KPC-2-producing *Klebsiella pneumoniae* in critically ill patients: what is the role of a previous colonization? *Infect Control Hosp Epidemiol* 2016;37:240–241.
10. Mendes AC, Rodrigues C, Pires J, et al. Importation of fosfomycin resistance *fosA3* gene to Europe. *Emerg Infect Dis* 2016;22:346–348.

Prevalence of Asymptomatic Bacteriuria in Hospitalized Patients

To the Editor—The prevalence of asymptomatic bacteriuria (ASB) varies widely based on the studied population. Currently, the prevalence of ASB in patients hospitalized in acute care institutions is unknown. Awareness of the prevalence of ASB in this setting would be useful in both medical decision making as well as public reporting of hospital-acquired urinary tract infections. In this prevalence study, 200 randomly selected patients admitted in April/May 2013 to a tertiary care academic center had urine samples collected for culture within 24 hours of being admitted. Data from the medical records were collected during these hospitalizations up to 30 days post-enrollment. The objective was to determine the prevalence of ASB. Of the 200 patients, 17 were found to have ASB for a prevalence of 8.5%.

Because infections acquired during a hospital stay are not always reimbursed by insurers, knowing what conditions were present on admission can be relevant from the hospital's perspective. ASB, usually defined as 1 (in men) or 2 separate