DOI: 10.1079/BJN20041142

British Journal of Nutrition (2004), **92**, Suppl. 1, S47–S57 © The Authors 2004

Peripheral and central signals in the control of eating in normal, obese and binge-eating human subjects

Per M. Hellström¹, Allan Geliebter², Erik Näslund³, Peter T. Schmidt¹, Eric K. Yahav², Sami A. Hashim² and Martin R. Yeomans⁴*

The worldwide increase in the incidence of obesity is a consequence of a positive energy balance, with energy intake exceeding expenditure. The signalling systems that underlie appetite control are complex, and the present review highlights our current understanding of key components of these systems. The pattern of eating in obesity ranges from over-eating associated with binge-eating disorder to the absence of binge-eating. The present review also examines evidence of defects in signalling that differentiate these sub-types. The signal-ling network underlying hunger, satiety and metabolic status includes the hormonal signals leptin and insulin from energy stores, and cholecystokinin, glucagon-like peptide-1, ghrelin and peptide YY3-36 from the gastrointestinal tract, as well as neuronal influences via the vagus nerve from the digestive tract. This information is routed to specific nuclei of the hypothalamus and brain stem, such as the arcuate nucleus and the solitary tract nucleus respectively, which in turn activate distinct neuronal networks. Of the numerous neuropeptides in the brain, neuropeptide Y, agouti gene-related peptide and orexin stimulate appetite, while melanocortins and α-melanocortin-stimulating hormone are involved in satiety. Of the many gastrointestinal peptides, ghrelin is the only appetite-stimulating hormone, whereas cholecystokinin, glucagon-like peptide-1 and peptide YY3-36 promote satiety. Adipose tissue provides signals about energy storage levels to the brain through leptin, adiponectin and resistin. Binge-eating has been related to a dysfunction in the ghrelin signalling system. Moreover, changes in gastric capacity are observed, and as gastric capacity is increased, so satiety signals arising from gastric and post-gastric cues are reduced. Understanding the host of neuropeptides and peptide hormones through which hunger and satiety operate should lead to novel therapeutic approaches for obesity; potential therapeutic strategies are highlighted.

Signalling systems: Obesity: Hormones

Energy balance is a metabolic state that exists when total energy expenditure equals dietary energy intake. Normally, energy balance is very well regulated. However, in some individuals there is an imbalance between energy intake and energy expenditure in favour of intake, resulting in weight gain and, ultimately, obesity. The prevalence of obesity, associated with chronic diseases such as diabetes and heart disease, keeps increasing in the USA (Flegal et al. 2002) and has reached epidemic proportions globally (Hill & Peters, 1998). Obesity is highly resistant to treatment, with most lost weight regained within 5 years. Obesity and the associated insulin resistance are also intimately linked with various medical complications and illnesses, such as hypertension, non-insulin-dependent diabetes mellitus, cancer, IHD and osteoarthritis among others, and represent a major and growing health problem worldwide.

Early research on obesity concentrated almost exclusively on hypothalamic controls of food intake. However,

as early as 1952, Kennedy postulated that the maximum daily intake of food during hyperphagia was determined by some limiting factor in addition to hypothalamic mechanisms (MacDougald & Mandrup, 2002). Today it is accepted that the control of feeding involves not only the central nervous system (CNS), but also the adrenal glands, the pancreas and the gastrointestinal (GI) tract. In addition, recent work has shown that adipose tissue has an important role in the regulation of food intake, as it secretes a number of endocrine and paracrine mediators, including leptin, adiponectin, resistin and TNF-α, which have been shown to influence appetite (Fig. 1). Our understanding of factors regulating food intake has increased greatly during the last 10 years, partly as a result of the discovery of leptin (Zhang et al. 1994). That study not only demonstrated that our main store of energy (fat) can signal energy reserves to the CNS, but also spawned an increased interest in the whole field of appetite regulation.

¹Department of Gastroenterology and Hepatology, Karolinska Hospital, Karolinska Institutet, Stockholm, Sweden

²Departments of Psychiatry and Medicine, NY Obesity Research Center, Babcock 10-A, St Luke's Roosevelt Hospital Center, Columbia University, College of Physicians and Surgeons, 1111 Amsterdam Ave, NY 10025, USA

³Division of Surgery, Karolinska Institutet Danderyd Hospital, Karolinska Institutet, Stockholm, Sweden

⁴Department of Psychology, School of Life Sciences, University of Sussex, Brighton, Sussex BN1 9QG, UK

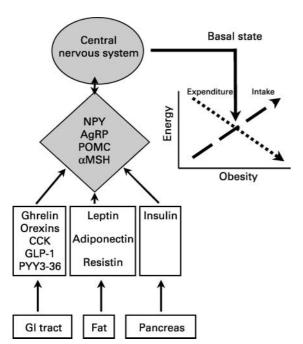


Fig. 1. Major neuropeptide and peptide hormone controls of food intake. The graph illustrates the balance between energy intake and expenditure in order to maintain a stable fat mass. NPY, neuropeptide Y; AgRP, agouti gene-related peptide; POMC, pro-opiomelanocortin; α MSH, α -melanocyte stimulating hormone; CCK, cholecystokinin; GLP, glucagon-like peptide; PYY, peptide YY; GI, gastrointestinal.

The resulting research has improved our knowledge of the signals underlying energy balance, and the main aim of the present review is to summarise our current understanding of peripheral and central signals in the regulation of food intake in normal and obese human subjects. The focus will be on the various peptide hormones and neuropeptides, and GI signals. However, other signals and agents, such as blood sugar and insulin, circulating NEFA, central catecholamines, 5-hydroxytryptamine and α_2 -adrenoceptors are also important in the control of food intake, but beyond the scope of the present review.

A sizable subset of obese subjects, 18-46% of those enrolling in weight-loss programmes, has binge-eating disorder (BED) (De Zwaan et al. 1994), characterised by ingestion of very large meals with loss of control, not followed by purging as in bulimia nervosa (Yanovski, 1993). As noted by Yanovski (1995), 'binge-eating disorder provides an opportunity to study the causes and concomitants of binge-eating in the absence of compensatory behaviors." BED, characterised relatively recently, is listed in the appendix of the Diagnostic and Statistical Manual IV-TR of the American Psychiatric Association. BED is found primarily in obese individuals, and patients with BED are generally more resistant to weight-loss treatment, have higher dropout rates, and show greater recidivism after weight loss (Yanovski, 1993) than other obese subjects. With the rising incidence of BED, it is important to study its aetiology and pathophysiology. Currently, the biological mechanisms underlying BED, including the role of the gut-brain axis, are poorly understood. Therefore, another aim of the present review is to examine whether sufferers from BED and related disorders display disturbances in any of the main signalling systems underlying appetite control.

Peripheral signals for hunger and satiety

There are three different sets of signals from the periphery, one from adipose tissue that exerts long-term regulatory mechanisms on food intake, and the other two from the GI tract and pancreas, with orexigenic as well as anorexigenic properties that exert primarily short-term effects on food intake (Fig. 1).

Adipocyte signalling

Leptin, secreted from adipose tissue, acts on neurons in the hypothalamus, leading to diminished food intake in animals. Leptin is considered to be part of a feedback loop where low levels of leptin signal to the CNS that energy stores are depleted (Zhang et al. 1994). Leptin, similar to insulin, decreases appetite by inhibiting CNS neurons containing neuropeptide Y (NPY) and agouti gene-related peptide. Brain receptors for leptin, as well as insulin, are found (together with receptors for glucocorticosteroids, oestrogen and progesterone) in the periventricular arcuate nucleus in the baso-medial hypothalamus next to the third ventricle, where exchange of molecules across the blood-brain barrier can occur. This area of the brain can be reached by hunger- and satiety-signalling hormones from the bloodstream. Moreover, both leptin and the leptin receptor proteins are present in the fundus of the gastric corpus in man. This suggests a paracrine link by which gastric epithelial cells may be direct targets for leptin (Sobhani et al. 2000). In addition, vagal stimulation causes leptin release from the stomach, suggesting that leptin is released during the cephalic phase of gastric secretion to exert another, still unknown, control mechanism (Sobhani et al. 2002). The released leptin may also signal satiety to the brain (Bray, 2000).

Since its discovery (Zhang et al. 1994), leptin has been shown to be a major regulator of food intake in rodents and genetically engineered animals. However, in human subjects, the balance between control of food intake and metabolism for stable weight control appears far more complex. In human subjects, obesity is associated with high plasma concentrations of leptin. In clinical trials using leptin as an anorexigenic agent, the term 'leptin resistance' has been coined for obese people who, in spite of adequate or even high plasma levels of leptin, do not respond to leptin in an orderly fashion resulting in decreased food intake (Heymsfield et al. 1999). Dysfunction of leptin systems may explain obesity only in a small number of cases where a specific genetic abnormality to the leptin system has been identified (O'Rahilly, 2003), requiring further exploration of alternative signals.

Another possible signalling factor is adiponectin (ARCP30, AdipoQ, apM1, GBP28), a peptide produced and released to the circulation exclusively by adipose tissue (Scherer *et al.* 1995). Adiponectin is a member of the complement C1q factor family and displays structural

homology with the TNF family (Shapiro & Scherer, 1998). There are important differences between adiponectin, insulin and leptin. Unlike leptin, plasma levels of adiponectin remain relatively constant throughout the day and are not affected by food intake (Hotta et al. 2000). Expression of adiponectin is decreased in obese and leptin-deficient ob/ ob mice (Hu et al. 1996; Pajvani et al. 2003), and in human subjects there is a negative correlation between BMI and plasma levels of adiponectin (Matsubara et al. 2002). Obese diabetic subjects have even lower plasma levels of adiponectin than non-diabetic obese people (Hotta et al. 2000; Berg et al. 2002; Hug & Lodish, 2002; Tsao et al. 2002; Ukkola & Santaniemi, 2002), suggesting that diminished adiponectin plays a role in the development of insulin resistance in non-insulin-dependent

Resistin, also known as adipose tissue-specific secretory factor, is another hormone secreted by adipocytes and acts on skeletal muscle myocytes, hepatocytes and adipocytes. Opposite in directional effects to adiponectin, higher resistin may decrease insulin sensitivity, leading to non-insulindependent diabetes mellitus (Steppan *et al.* 2001).

Two additional peptides have been described recently, fasting-induced adipose factor, also known as hepatic fibrinogen-angiopoeitin-related protein (Kersten et al. 2000; Kim et al. 2000) and the perilipins (Londos et al. 1996; Tansey et al. 2001). Animal studies have shown that the fasting-induced adipose factor peptide is predominantly expressed in adipose tissue, and that fasting-induced adipose factor in white adipose tissue and liver is strongly upregulated by fasting. Fasting-induced adipose factor might also act as an apoptosis survival factor for vascular endothelial cells and has therefore been discussed in the context of metabolic and cardiovascular complications of obesity (Trayhurn & Beattie, 2001). The perilipins are the most abundant proteins coating the surfaces of lipid droplets in adipocytes and are found at lower levels surrounding lipid droplets in steroidogenic cells. Perilipins drive the fat accumulation in cells through triacylglycerol storage in adipocytes by regulating the rate of basal lipolysis. Perilipins are also required to maximise hormonally stimulated energy utilisation by lipolysis (Londos et al. 1996; Tansey et al. 2001). Whether these peptides also play a role in signalling energy stores to the CNS like leptin has yet to be determined.

Gastrointestinal signalling

diabetes mellitus.

There are now a number of GI peptide hormones whose activity can be related to a variety of food-related gastric and intestinal cues (Read *et al.* 1994). Distention of the stomach activates gastric stretch receptors and mechanoreceptors that transmit satiety signals (Schwartz *et al.* 1993). The stomach also serves as a food reservoir, with a capacity that can limit meal intake. Although the specific relationship between these gastric signals and the release of GI neuropeptides related to appetite has not been elucidated adequately, recent research has clarified the effects of many of the neuropeptides.

The only known GI peptide hormone with confirmed orexigenic properties is ghrelin, first identified in the

gastric mucosa (Kojima et al. 1999). Ghrelin, a ligand for growth hormone secretagogue receptor, is produced mainly by the stomach and, when administered, increases food intake in animals (Tschop et al. 2000) and human subjects (Wren et al. 2001) without affecting gastric emptying. Plasma levels of ghrelin peak before a meal, then decrease to a nadir and again increase to a peak that is reduced by the next meal (Cummings et al. 2001). Effects of ghrelin on eating may be mediated via the arcuate nucleus and the solitary tract nucleus (nucleus tractus solitarius) in the brain. Ghrelin also stimulates the release of growth hormone (Takaya et al. 2000) and is involved in the hypothalamic regulation of metabolic control and energy balance (Cummings & Foster, 2003). Ghrelin opposes leptin by stimulating different neuropeptides in the hypothalamus (Nakazato et al. 2001). The finding that appetite-stimulating effects of ghrelin have little impact on GI functions (Wren et al. 2001) suggests a central nervous effect of ghrelin. Ghrelin concentration is 27 % lower in obese subjects than in normal-weight subjects (Tschop et al. 2001) and is negatively correlated with BMI (Shiiya et al. 2002). Fasting ghrelin concentration is greatest in anorexia nervosa, consistent with ghrelin concentration rising in animals during starvation (Asakawa et al. 2001), while weight gain in anorexia nervosa decreases plasma ghrelin concentration (Otto et al. 2001). The increase in ghrelin concentration during starvation may promote eating and the fall in ghrelin concentration in obesity may be a secondary response to over-eating (Tschop et al. 2001) rather than causal (Geliebter, 2002). Indeed, fasting ghrelin concentration is negatively correlated with body fat (%) and fasting insulin and fasting leptin concentrations, all of which are elevated in obesity (Tschop et al. 2001; Shiiya et al. 2002). Even if ghrelin is not a primary causal factor in obesity, a ghrelin antagonist may still help reduce food intake in obesity.

Orexins are novel appetite stimulating neuropeptides that appear to play a role in regulation of food intake, arousal and energy homeostasis (Kirchgessner, 2002). Initial reports suggested that orexin A (OXA) and orexin B (OXB) were produced exclusively by a small group of neurons in the lateral hypothalamic area (Sakurai et al. 1998). However, neurons in the GI submucosal and myenteric plexuses, and endocrine cells in the intestinal mucosa and pancreatic islets have recently been shown to display OXA and orexin receptor immunoreactivity (Kirchgessner & Liu, 1999; Näslund et al. 2002). OXA inhibits fasting GI motility in the rat and modulates both insulin and glucagon release from the endocrine rat pancreas (Nowak et al. 2000; Ouedrago et al. 2003). Moreover, orexin-positive neurons in the gut, like those in the hypothalamus, are activated by fasting, indicating a functional response to the nutritional status of these cells (Kirchgessner & Liu, 1999). Recently, orexin receptors have been found on vagal afferent neurons in both rats and humans subjects and OXA inhibited vagal responses to cholecystokinin (CCK; Burdyga et al. 2003). Plasma concentrations of OXA are increased during fasting in human subjects (Komaki et al. 2001) and are lower in obese subjects compared with normal-weight subjects in the fasted state (Adam et al. 2002). Taken together, these results imply a role for OXA in the control of food intake, but the precise nature of any such involvement remains to be clarified.

Conversely, a number of different GI peptide hormones are known to be anorexigenic, promoting early termination of the meal. Of these, CCK, glucagon-like peptide (GLP)-1 and peptide YY (PYY) 3-36 have received recognition as physiological regulators of food intake. CCK is the most widely studied GI peptide regulating food intake control, and research on CCK has been widely reviewed (Ritter et al. 1999). After a meal, CCK is released into the bloodstream from endocrine I cells of the duodenum and the jejunum (Buchan et al. 1978). Studies in human subjects have shown repeatedly that CCK inhibits food intake (Kissileff et al. 1981; Ballinger et al. 1995), although a concurrent gastric preload is generally necessary to achieve a satiety effect with CCK in obese and normal subjects (Lieverse et al. 1993). Peripherally administered CCK also acts on CCKA receptors in the gastric antrum; these receptors are involved in the CCK-mediated inhibition of gastric emptying (Reubi et al. 1997). CCKA receptors are also found in the abdominal part of the vagus nerve (Mercer & Lawrence, 1992) and vagotomy abolishes the satiating effect of the peptide in rats. A relationship between decreased plasma levels of CCK and hunger, as well as decreased fullness, has also been reported in

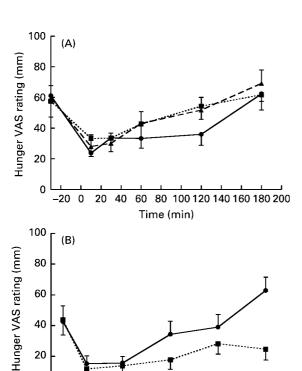


Fig. 2. Hunger ratings in two sets of experiments with intravenously administered glucagon-like peptide-2 (A) at 0.75 (■) or (▲) 2.25 pmol/kg per min or glucagon-like peptide-1 (B) at 0.75 pmol/kg per min (■) or sodium chloride solution (9 g/l; •). VAS, visual analoque scale. Values are means with their standard errors shown by vertical bars. Mean values for glucagon-like peptide-1 were significantly different from those for sodium chloride solution: *P<0.05 (From Näslund et al. 1999b; Schmidt et al. 2003).

100

Time (min)

140

180

220

260

20

-20

20

60

human subjects (French et al. 1993), supporting a role for CCK as a physiological mediator of satiety.

GLP-1 and GLP-2 are produced and secreted from endocrine L-cells found in the mucosa of the ileum and colon. After food ingestion, both peptides are released in equimolar amounts to the circulation (Bell et al. 1983; Holst, 1997). GLP-1 is a major contributor to the ileal brake mechanism of the upper GI tract, thereby modulating gastric emptying and acid secretion (Näslund et al. 1999b). GLP-1 also exerts dual actions in regulation of blood glucose concentrations through insulinotropic and glucagonostatic actions (Ørskov, 1992). Since GLP-1 slows gastric emptying of liquid as well as solid meals (Wettergren et al. 1993; Näslund et al. 1999b), metabolic requirements for insulin after food intake are reduced (Nauck et al. 1997). Accumulating evidence indicates that effects of GLP-1 on GI functions are mediated via the vagus nerve both in animals and human subjects (Imeryuz et al. 1997; Wettergren et al. 1997; Tolessa et al. 1998; Wettergren et al. 1998).

GLP-1 also exerts dual actions on feeding behaviour and satiety: intracerebroventricular injection of GLP-1 in rats inhibits food and water intake (Lambert et al. 1994; Tang-Christensen et al. 1996; Turton et al. 1996) and induces c-fos expression in the paraventricular nucleus of the hypothalamus (Turton et al. 1996). Administration of the GLP-1 receptor antagonist, exendin(9-39)amide, intracerebroventricularly to satiated rats resulted in increased food intake, but not in fasted rats (Turton et al. 1996). Furthermore, rats receiving continuous intracerebroventricular treatment with exendin(9-39)amide not only increased food intake, but also significantly increased their body weight (Bloom, 1997). No effect was seen with intraperitoneal injections, suggesting central signalling as the mode of action of GLP-1 in satiety. In human subjects, most, but not all (Long et al. 1999), studies report decreased food intake after administration of GLP-1, including studies in normal weight, diabetic and obese subjects (Flint et al. 1998, 2000; Näslund et al. 1998b, 1999b; Gutzwiller et al. 1999a,b; Toft-Nielsen et al. 1999) (Fig. 2). Reduced food intake is consistent with ratings of reduced hunger, and increased fullness following GLP-1 infusion (Gutzwiller et al. 1999b; Näslund et al. 1999a).

In contrast to GLP-1, the role of GLP-2 in regulation of food intake is inconsistent. GLP-2 is primarily known to exert trophic effects on the intestinal mucosa (Tsai et al. 1997), leading to advances in the treatment of short bowel syndrome (Lovshin & Drucker, 2000). GLP-2 has been reported to affect gastric motor activity in pigs (Wøjdemann et al. 1998). In addition, GLP-2-containing neurons connect the nucleus tractus solitarius with the medial hypothalamus, where GLP-2 receptor mRNA is expressed (Tang-Christensen et al. 2000). Furthermore, central administration of GLP-2 to rodents reduced food intake compared with placebo, suggesting that GLP-2 may be another satiety-promoting peptide (Tang-Christensen et al. 2000). However, this does not seem to be the case in human subjects. Gastric emptying studies including ratings of satiety have failed to verify any effect of GLP-2 (Schmidt et al. 2003) (Figs 2 and 3), and peripheral administration of GLP-2 did not influence appetite and ad libitum

food intake (Sørensen *et al.* 2003). Why the effects of GLP-2 on appetite in rodents and human subjects seem to differ is unclear and requires substantiation.

PYY itself is well defined as an orexigenic neuropeptide (Morley et al. 1985; Hagan, 2002). However, the truncated form of PYY, PYY3-36, has anorexigenic effects, is released postprandially from the GI tract in proportion to the energy content of a meal and induces satiety (Batterham et al. 2002). PYY3-36 exerts its action as an agonist on Y₂ receptors, thereby suppressing NPY-driven hunger feelings. PYY infusion was shown recently to reduce food intake in obese and lean subjects; moreover, fasting plasma PYY was lower in the obese than the lean subjects (Batterham et al. 2003).

Neuropeptidergic control of appetite is also regulated through insulin from the β -cells of the pancreatic endocrine islets of Langerhans, which, like leptin, downregulate NPY and agouti gene-related peptide, resulting in decreased appetite and food intake. Insulin also stimulates melanocortin-producing neurons in the hypothalamus (Porte *et al.* 2002).

Distortion of gastric satiety cues in obesity and binge-related disorders

As we have seen, except for PYY3-36, which is reduced in obesity, there is no clear evidence of differences in peptide hormones between normal and obese subjects that could explain higher food intake as a component of obesity (the lower levels of ghrelin would suggest reduced appetite). Importantly, obese subjects in general have a larger than normal gastric capacity (Geliebter, 1988) as do normal-weight individuals with bulimia nervosa (Geliebter et al. 1992). A stomach with a large capacity may require a larger meal than usual to generate satiety signals, including release of satiety-related GI peptides and, indeed, gastric capacity correlates highly with test-meal intake (Geliebter et al. 1992). Further evidence for the importance of gastric

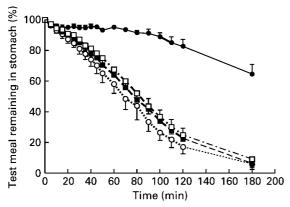


Fig. 3. Gastric emptying curves from two different sets of experiments where glucagon-like peptide-1 (0·75 pmol/kg per min; ●) or sodium chloride (9 g/l; ○) and glucagon-like peptide-2 (0·75 pmol/kg per min; ■) or sodium chloride solution (9 g/l; □) were given intravenously for 180 min to eight healthy volunteers. Values are means with their standard errors shown by vertical bars. Mean values for glucagon-like peptide-1 were significantly different from those for sodium chloride solution: P < 0.001 for time effect, treatment effect and time × treatment interaction for glucagon-like peptide-1 (From Näslund $et\ al.\ 1999\ b$; Schmidt $et\ al.\ 2003$).

capacity is the observation that surgical reduction in gastric capacity reduces meal size and induces marked weight loss (Brolin, 1992). Surgical reduction in capacity can also eliminate binge-eating in BED (Adami *et al.* 1999), confirming that a large stomach capacity helps maintain binge-eating. Moreover, non-surgical reduction of stomach capacity by external abdominal pressure (Geliebter *et al.* 1986) or by filling an intragastric balloon also reduces meal intake, especially in the short term (Geliebter, 1988).

A stomach with increased capacity may also empty fixed liquid meals more slowly, and slower emptying has been seen in bulimia nervosa (Geracioti & Liddle, 1988; Geliebter et al. 1992). When gastric capacity was contrasted between subjects with bulimia nervosa, obese subjects and matched controls in the same laboratory, gastric capacity was largest in the subjects with bulimia nervosa, intermediate in obese subjects and smallest in normalweight subjects (Geliebter & Hashim, 2001). Delayed gastric emptying by promoting greater gastric distension is generally associated with greater satiety, but it may also diminish intestinal satiety cues, including delaying the duodenal release of CCK as in bulimia nervosa (Geracioti & Liddle, 1988). Although impaired CCK effects in obesity in general have not been found, the observation that obesity includes a number of clinically distinct sub-groups, most clearly non-BED and BED, raises the possibility that obese subjects with BED have similar changes in gastric capacity and CCK release to bulimia nervosa. Indeed, when obese subjects were sub-classified as binge-eaters and non-binge-eaters, binge-eaters were similar in stomach capacity to bulimia nervosa patients, while the non-bingeeaters were similar to normal subjects (Geliebter et al. 1992). However, only a few of the subjects with bingeeating in that study had full-fledged BED. The possibility that obese patients who meet full diagnostic criteria for BED had even greater changes in gastric capacity and consequent satiety signalling was assessed in a recent study of BED (Geliebter et al. 2004). In that study, and in a number of the studies referred to in the present review, gastric emptying of liquids was estimated with the tracer paracetamol, which is rapidly absorbed from the duodenum after leaving the stomach (Sanaka et al. 1997, 1998), and correlates well with radiolabelling and intragastric suction. In brief, gastric

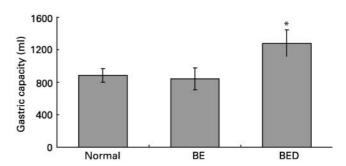


Fig. 4. Gastric capacity, based on volume tolerated, in obese patients classified as having binge-eating disorder (BED), subthreshold BED (BE) or non-bingeing (normal). Values are means with their standard errors shown by vertical bars. Mean value was significantly greater than those of BE or normal groups: *P <0.05. (From Geliebter *et al.* 2004.)

capacity and gastric emptying, and levels of glucose, insulin, leptin, CCK and ghrelin, were assessed while fasting and after a test meal in obese patients who were diagnosed with BED (n 11), binge-eating but not full BED (n 13) or normal without binge-eating (n 13). Gastric capacity was greater in the BED than binge-eating or normal groups, as assessed by filling the stomach with an intragastric balloon until maximal tolerance and by measuring intragastric pressure to determine compliance, although not as great as that reported previously in bulimia nervosa (Geliebter et al. 1992) (Fig. 4). Test-meal size correlated with gastric capacity across all subjects. Gastric emptying did not differ significantly among groups. The only fasting hormone concentration to differ significantly between groups was ghrelin, which was lowest during fasting in BED, intermediate in binge-eaters and highest in the normal subjects. Ghrelin decreased after a fixed meal as expected, but decreased the least in BED from a lower baseline. Ghrelin at 30 min correlated inversely with gastric capacity: r -0.36, P < 0.05. The lower fasting ghrelin suggests that binge-eating results in downregulation of ghrelin, consistent with lower levels in obese subjects as compared with lean subjects (Tschop et al. 2001). If the extent of the fall in ghrelin after a meal is itself a signal for satiety, the smaller relative decrease in ghrelin postprandially may nevertheless contribute to the binge-eating in BED. Overall, these results demonstrate a greater gastric capacity and abnormal ghrelin pattern in BED.

Another peptide that could contribute to obesity is GLP-1. It has been reported that the plasma increase of GLP-1 after a meal is attenuated in obese subjects (Ranganath *et al.* 1996, 1999; Näslund *et al.* 1998a). As GLP-1 strongly inhibits gastric emptying by about 50 % 3 h after

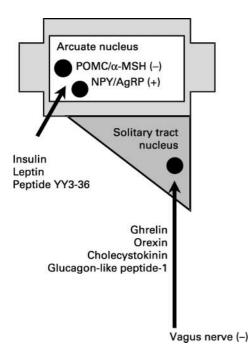


Fig. 5. Schematic illustration of peripheral and central signalling for the control of food intake. POMC, pro-opiomelanocortin; α MSH, α -melanocyte stimulating hormone; NPY, neuropeptide Y; AgRP, agouti gene-related peptide.

a meal (Fig. 3), low GLP-1 concentrations in obese subjects may promote earlier onset of subsequent meals. Moreover, when obese patients undergo jejuno-ileal bypass, their gastric emptying rate is slowed and GLP-1 response to the ingested meal restored (Näslund et al. 1998a). In families with morbid obesity, there is a genetic linkage between islet 1 locus on chromosome 5q and the ancestor for GLP-1, namely the proglucagon gene. Since islet 1 locus is a positive regulator of proglucagon gene transcription, this may influence GLP-1 elaboration and release to the circulation (Clement et al. 1999). Thus, a defect in this system may result in decreased plasma levels of GLP-1 with short postprandial satiety periods and short meal intervals, resulting in increased daily food intake. None of these studies of GLP-1 has examined any distinction between BED and non-binge-eating groups, and future research needs to integrate these approaches to allow more detailed evaluation of the importance of GLP-1.

Interactions between peripheral and central homeostatic controls of eating in normal and obese men

So far, the present review has examined peripheral and central homeostatic controls of appetite separately, but of course these signals ultimately are integrated in order to direct food intake. The classic model of central nervous regulation of food intake was the 'dual-centre model' (Stellar, 1954), which suggested that different neuronal nuclei in the hypothalamus governed food intake by modifying hunger and satiety. However, this model has since been shown to be simplistic, and neural structures beyond those described in the dual-centre model (the lateral and ventromedial hypothalamic nuclei) are now strongly implicated in the regulation of appetite. For example, both the arcuate nucleus and the periventricular nuclei are important centres in the control of food intake, while the nucleus of the solitary tract serves as a gateway of signals from the GI tract that are relayed to the hypothalamic level (Fig. 5). In addition, there is increasing recognition that non-homeostatic mechanisms, particularly reward processes, play an important role in short-term control of food intake, although these processes are beyond the scope of the present review.

The arcuate nucleus seems to be of central importance for the inflow of peripheral signals mediating information on metabolic storage and needs. In the arcuate nucleus there are two different populations of cells with an explicit role in food intake (Fig. 2). One of these cell populations contains NPY, which is a powerful stimulator of feeding behaviour in animals (Chamorro et al. 2002), whereas the other contains pro-opiomelanocortin, which is split into α -melanocyte stimulating hormone; this then inhibits eating by acting on melanocortin receptors (Wikberg et al. 2000). Peptide hormone receptors are located on these two cell populations, both of which are controlled by leptin and insulin; these inhibit the NPY-containing neurons and stimulate the pro-opiomelanocortin-containing neurons. The relevant postsynaptic NPY receptors are mainly of the Y₁-type, whereas presynaptic inhibitory Y₂-receptors are located directly on the NPY neurons (Fuxe et al.

1997; Broberger et al. 1999). Interestingly, PYY3-36 released from the GI tract after food intake is the natural ligand for these Y₂ receptors (Batterham et al. 2002). This implies that feedback signals from the GI tract not only act on the brain stem, but also on the hypothalamus. Within the arcuate nucleus there is also inhibitory crosstalk between the two neuropeptide systems, with NPY neurons exerting inhibitory input on α-melanocyte stimulating hormone neurons and agouti gene-related peptide, adding to the inhibition of the feeding-suppressive α -melanocyte stimulating hormone system (Broberger et al. 1998; Hahn et al. 1998). In cells considered to be targets for the innervation of NPY- and pro-opiomelanocortin-containing neurons in the lateral hypothalamus, another two orexigenic peptides, OXA and OXB (Sakurai et al. 1998), have been found.

The anorexigenic peptide GLP-1 is found in the nucleus tractus solitarius and in dorsal and ventral parts of the medullary reticular nucleus, corresponding to brain stem regions that receive vagal afferents from the gut (Jin et al. 1988). GLP-1 immunoreactive nerve fibres are also found in the hypothalamic paraventricular nucleus with fibres projecting to the lateral hypothalamus and periventricular strata (Shimizu et al. 1987; Jin et al. 1988; Kreymann et al. 1989). Interestingly, when OXA and GLP-1 are administered peripherally, they selectively activate c-fos expression in the lateral and baso-medial hypothalamus respectively (Levin, J Ma, Tong, PM Hëllstrom, AL Kirchgessner and E Näslund, unpublished results; Fig. 6), again indicating that GI signalling exerts a dual input both at a hypothalamic and brain stem level.

What then are the major central and peripheral signals associated with meal termination? A classic finding was that decerebrate rats are capable of terminating their eating upon distension of the stomach with activation of stretch receptors, but were incapable of increasing energy intake in response to energy dilution (Grill & Norgren, 1978). Structures in the brain stem appear to primarily control meal volume, energy content and duration, whereas the hypothalamus primarily senses and balances the metabolic status in the control of food intake. These feedback mechanisms involve a host of peptide hormones located at various points along the GI tract. In addition to their central

receptors, signals mediated from peripheral receptors are transferred via the vagus nerve to the nucleus tractus solitarius in the brain stem (Raybould, 1998). The peripheral and central vagus systems sense mechanical distension and chemical stimulation by different nutrients. These signals do not only affect the sensory nerve fibres directly, but are also released in the form of hormones to the circulation. Important examples of this are CCK and GLP-1. The effects of these peptides are likely to be mediated through the vagus nerve. In the vagus, the satiating effects of CCK are mediated by the CCKA receptor (Hewson et al. 1988; Reidelberger et al. 1991), while the effects of GLP-1 are mediated through its distinct receptor. Furthermore, sensory vagal ganglia in the rat, predominately those expressing the CCKA receptor, contain high levels of cocaine- and amphetamine-regulated transcript, which itself is associated with reduced food intake (Broberger et al. 1999). In general terms, the vagus nerve is activated by CCK and GLP-1 as the stomach is distended by food and nutrients are being delivered to the small intestine. Next, in the brain, vagal nerve fibres release cocaine- and amphetamine-regulated transcript in the nucleus tractus solitarius of the brain stem, where its satiety-promoting effect is greatest.

Pathways to the development of future anorexic drugs

Of the various mechanisms reviewed here, the melanocortin system has attracted greatest interest from a pharmaceutical perspective. Mutations in the melanocortin-4 receptor are not as uncommon as deleterious mutations in other systems regulating fat storage and may contribute to the early development of morbid obesity (Yeo et al. 1998). Melanocortin-4 receptor mutation was recently found to be much more common in BED than in non-BED obese subjects (Branson et al. 2003). The melanocortin-4 receptors involved have a limited distribution, which might reduce the risk of side effects. The neuropeptides acting within this area also seem to be extremely effective. For example, in the rat, central injection of agouti gene-related peptide causes long-standing satiety for up to 1 week after administration (Hagan et al. 2000). Research with GLP-1 and PYY also suggests clinical potential. Treatment with

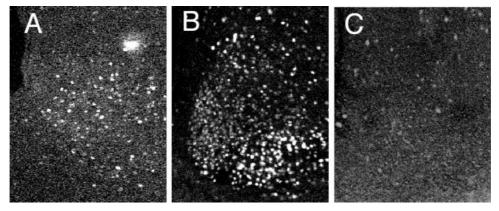


Fig. 6. Mean numbers of *c-fos*-positive neurons in the arcuate nucleus in control rats ((A), 30·2 (SEM 1·7)) and orexin A ((B), 114·0 (SEM 6·9)) and glucagon-like peptide 1 ((C), 25·0 (SEM 2·3)) -treated rats. (From Levin J Ma, Tong, PM Hëllstrom, AL Kirchgessner and E Näslund, unpublished results.)

GLP-1 in overweight subjects reduced their appetite and daily intake, leading to weight control (Näslund et al. 1999b; Näslund et al. 2004). Furthermore, infusion of physiologically appropriate of PYY3-36 decreased rated appetite and reduced daily food intake (Batterham, 2002) in normal and obese subjects (Batterham et al. 2003). This is in contrast to other satiety-stimulating peptide hormones such as CCK, where the latency period to the next meal was shortened and therefore the daily intake was unchanged. Although CCK today is less attractive as a target treatment, it may be beneficial when combined with a centrally acting 5-hydroxytryptamine-1A receptor agonist (Poeschla et al. 1993), since this receptor is considered pivotal for satiety feelings caused by peripheral peptide administration. Thus, in cases where single peptide treatments do not work, a dual treatment modality may be of choice for control of peripheral and central signalling in eating behaviour. It is also important to recognise that different treatment strategies will be needed to distinguish between binge-related and non-binge-related obesity.

Summary

The present review highlights the complexity of both the central and peripheral signalling systems underlying homeostatic controls of food ingestion and the complexity of obesity. Historically, there have been numerous breakthroughs, such as the initial discovery of CCK, the more recent discoveries of leptin, ghrelin and GI peptides, all of which have been heralded as likely to lead to major breakthroughs in the treatment of obesity. In practice, the multiplicity of controls greatly reduces the likelihood of treatments based on modification of any one signalling system. However, recent advances in the understanding of GLP-1, ghrelin and PYY and studies showing altered gastric capacity in obesity and BED pave the way for more novel single or combined pharmaceutical approaches to therapy, as well as greatly increasing our basic understanding of the underlying controls of appetite. Future research should be focused on the possible interactions between these systems, and how they relate to different forms of obesity and disordered eating.

References

- Adam JA, Menheere PP, van Dielen FM, Soeters PB, Buurman WA & Greve JW (2002) Decreased plasma orexin-A levels in obese individuals. *Int J Obes Relat Metab Disord* **26**, 274–276.
- Adami GF, Meneghelli A & Scopinaro N (1999) Night eating and binge-eating disorder in obese patients. *Int J Eat Disord* **25**, 335–338.
- Asakawa A, Inui A, Kaga T, Yuzuriha H, Nagata T, Fujimiya M, Katsuura G, Makino S, Fujino MA & Kasuga M (2001) A role of ghrelin in neuroendocrine and behavioral responses to stress in mice. *Neuroendocrinology* **74**, 143–147.
- Ballinger A, McLoughlin L, Medback S & Clark M (1995) Cholecystokinin is a satiety hormone in humans at physiological concentrations. *Clin Sci* **89**, 375–381.
- Batterham RL (2002) Gut hormone PYY3-36 physiologically inhibits food intake. *Nature* **418**, 650–654.

- Batterham RL, Cohen MA, Ellis SM, Le Roux CW, Withers DJ, Frost GS, Ghatei MA & Bloom SR (2003) Inhibition of food intake in obese subjects by peptide YY3-36. *N Engl J Med* **349**, 941–948.
- Batterham RL, Cowley MA, Small CJ, *et al.* (2002) Gut hormone PYY(3-36) physiologically inhibits food intake. *Nature* **418**, 650–654.
- Bell GI, Santerre RF & Mullenbach GT (1983) Hamster preproglucagon contains the sequence of glucagon and two related peptides. *Nature* **302**, 716–718.
- Berg AH, Combs TP & Scherer PE (2002) ACRP30/adiponectin: an adipokine regulating glucose and lipid metabolism. *Trends Endocrinol Metab* 13, 84–89.
- Bloom SR (1997) Glucagon-like peptide-1 and satiety. *Nature* **385**, 214.
- Branson R, Potoczna N, Kral JG, Lentes KU, Hoehe MR & Horber FF (2003) Binge eating as a major phenotype of melanocortin 4 receptor gene mutations. N Engl J Med 348, 1096–1103.
- Bray GA (2000) Afferent signals regulating food intake. *Proc Nutr Soc* **59**, 373–384.
- Broberger C, Holmberg K, Kuhar MJ & Hokfelt T (1999) Cocaine- and amphetamine-regulated transcript in the rat vagus nerve: a putative mediator of cholecystokinin-induced satiety. *Proc Natl Acad Sci USA* **96**, 13506–13511.
- Broberger C, Johansen J, Johansson C, Schalling M & Hokfelt T (1998) The neuropeptide Y/agouti gene-related protein (AGRP) brain circuitry in normal, anorectic, and monosodium glutamate-treated mice. *Proc Natl Acad Sci USA* **95**, 15043–15048.
- Brolin RE (1992) Critical analysis of results: weight loss and quality of data. *Am J Clin Nutr* **55**, 577S–581S.
- Buchan AM, Polak JM, Solcia E, Capella C, Hudson D & Pearse AG (1978) Electron immunohistochemical evidence for human intestinal I cells as the source of CCK. Gut 19, 403–407.
- Burdyga G, Lal S, Spiller D, *et al.* (2003) Localization of orexin-1 receptors to vagal afferent neurons in the rat and humans. *Gastroenterology* **124**, 129–139.
- Chamorro S, Della-Zuana O, Fauchere JL, Feletou M, Galizzi JP & Levens N (2002) Appetite suppression based on selective inhibition of NPY receptors. *Int J Obes Relat Metab Disord* **26**, 281–298.
- Clement K, Dina C, Basdevant A, Chastang N, Pelloux V, Lahlou N, Berlan M, Langin D, Guy-Grand B & Froguel P (1999) A sib-pair analysis study of 15 candidate genes in French families with morbid obesity: indication for linkage with islet 1 locus on chromosome 5q. *Diabetes* 48, 398–402.
- Cummings DE & Foster KE (2003) Ghrelin–leptin tango in bodyweight regulation. *Gastroenterology* **124**, 1532–1535.
- Cummings DE, Purnell JQ, Frayo RS, Schmidova K, Wisse BE & Weigle DS (2001) A preprandial rise in plasma ghrelin levels suggests a role in meal initiation in humans. *Diabetes* **50**, 1714–1719.
- De Zwaan M, Mitchell JE, Raymond NC & Spitzer RL (1994) Binge eating disorder: clinical features and treatment of a new diagnosis. *Harv Rev Psychiatry* 1, 310–325.
- Flegal KM, Carroll MD, Ogden CL & Johnson CL (2002) Prevalence and trends in obesity among US adults, 1999–2000. *J Am Med Assoc* **288**, 1723–1727.
- Flint A, Raben A, Astrup A & Holst JJ (1998) Glucagon-like peptide-1 promotes satiety and suppresses energy intake in humans. *J Clin Invest* **101**, 515–520.
- Flint A, Raben A, Rehfeld JF, Holst JJ & Astrup A (2000) The effect of glucagon-like peptide-1 on energy expenditure and substrate metabolism in humans. *Int J Obes* **24**, 288–298.
- French SJ, Murray B, Rumsey RD, Sepple CP & Read NW

- (1993) Is cholecystokinin a satiety hormone? Correlations of plasma cholecystokinin with hunger, satiety and gastric emptying in normal volunteers. *Appetite* **21**, 95–104.
- Fuxe K, Tinner B, Caberlotto L, Bunnemann B & Agnati L (1997) NPY Y1 receptor-like immunoreactivity exists in a subpopulation of β-endorphin immunoreactive nerve cells in the arcuate nucleus: a double immunolabelling analysis in the rat. Neurosci Lett 225, 49–52.
- Geliebter A (1988) Gastric distension and gastric capacity in relation to food intake in humans. *Physiol Behav* **44**, 665–668.
- Geliebter A (2002) Weight loss and plasma ghrelin levels. *N Engl J Med* **347**, 1379–1381.
- Geliebter A & Hashim SA (2001) Gastric capacity in normal, obese, and bulimic women. *Physiol Behav* **74**, 743–746.
- Geliebter A, Melton PM, McCray RS, Gallagher DR, Gage D & Hashim SA (1992) Gastric capacity, gastric emptying, and testmeal intake in normal and bulimic women. *Am J Clin Nutr* **56**, 656–661.
- Geliebter A, Westreich S, Pierson RN & Van Itallie TB (1986) Extra-abdominal pressure alters food intake, intragastric pressure, and gastric emptying rate. *Am J Physiol* **250**, R549–R552.
- Geliebter A, Yahav E, Gluck M & Hashim SA (2004) Gastric capacity, test meal intake, and appetitive hormones in binge-eating disorder. *Physiol Behav* (In Press).
- Geracioti TD & Liddle RA (1988) Impaired cholecystokinin secretion in bulimia nervosa. *N Engl J Med* **319**, 683–688.
- Grill HJ & Norgren R (1978) Chronically decerebrate rats demonstrate satiation but not bait-shyness. Science 201, 267–269.
- Gutzwiller JP, Drewe J, Göke B, Schmidt H, Rohrer B, Lareida J & Beglinger C (1999a) Glucagon-like peptide-1 promotes satiety and reduces food intake in patients with diabetes mellitus type 2. Am J Physiol 276, R1541–R1544.
- Gutzwiller JP, Göke B, Drewe J, Hildebrand P, Ketterer S, Handschin D, Winterhalder R, Conen D & Beglinger C (1999b) Glucagon-like peptide-1: a potent regulator of food intake in humans. *Gut* **44**, 81–86.
- Hagan MM (2002) PeptideYY: a key mediator of orexigenic behavior. *Peptides* **23**, 377–382.
- Hagan MM, Rushing PA, Pritchard LM, Schwartz MW, Strack AM, Van Der Ploeg LH, Woods SC & Seeley RJ (2000) Long-term orexigenic effects of AgRP-(831-32) involve mechanisms other than melanocortin receptor blockade. Am J Physiol Regul Integr Comp Physiol 279, R47-R52.
- Hahn TM, Breininger JF, Baskin DG & Schwartz MW (1998) Coexpression of Agrp and NPY in fasting-activated hypothalamic neurons. *Nat Neurosci* 1, 271–272.
- Hewson G, Leighton RG & Hughes J (1988) The cholecystokinin receptor antagonist L364,718 increases food intake in the rat by attenuation of the action of endogenous cholecystokinin. *Br J Pharmacol* **93**, 79–84.
- Heymsfield SB, Greenberg AS, Fujioka K, et al. (1999) Recombinant leptin for weight loss in obese and lean adults: a randomized, controlled, dose-escalation trial. J Am Med Assoc 282, 1568–1575.
- Hill JO & Peters JC (1998) Environmental contributions to the obesity epidemic. *Science* **280**, 1371–1374.
- Holst JJ (1997) Enteroglucagon. Annu Rev Physiol 59, 257–271.
 Hotta K, Funahashi T, Arita Y, et al. (2000) Plasma concentrations of a novel, adipose-specific protein, adiponectin, in type 2 diabetic patients. Arterioscler Thromb Vasc Biol 20, 1595–1599.
- Hu E, Liang P & Spiegelman BM (1996) AdipoQ is a novel adipose-specific gene dysregulated in obesity. *J Biol Chem* **271**, 10697–10703.

- Hug C & Lodish HF (2002) Diabetes, obesity, and Acrp30/adiponectin. *Biotechniques* 33, 654, 656, 658 passim.
- Imeryuz N, Yegen BC, Bozkurt A, Coskun T, Villanueva-Penacarrillo ML & Ulusoy NB (1997) Glucagon-like peptide-1 inhibits gastric emptying via vagal afferent-mediated central mechanisms. *Am J Physiol* **273**, G920–G927.
- Jin SL, Han VK, Simmons JG, Towle AC, Lauder JM & Lund PK (1988) Distribution of glucagonlike peptide I (GLP-I), glucagon, and glicentin in the rat brain: an immunocytochemical study. *J Comp Neurol* **271**, 519–532.
- Kersten S, Mandard S, Tan NS, Escher P, Metzger D, Chambon P, Gonzalez FJ, Desvergne B & Wahli W (2000) Characterization of the fasting-induced adipose factor FIAF, a novel peroxisome proliferator-activated receptor target gene. *J Biol Chem* **275**, 28488–28493.
- Kim I, Kim HG, Kim H, Kim HH, Park SK, Uhm CS, Lee ZH & Koh GY (2000) Hepatic expression, synthesis and secretion of a novel fibrinogen/angiopoietin-related protein that prevents endothelial-cell apoptosis. *Biochem J* **346**, 603–610.
- Kirchgessner AL (2002) Orexins in the brain–gut axis. *Endocrol Rev* 23, 1–15.
- Kirchgessner AL & Liu M (1999) Orexin synthesis and response in the gut. *Neuron* **24**, 941–951.
- Kissileff HR, Pi-Sunyer FX, Thornton J & Smith GP (1981) C-terminal octapeptide of cholecystokinin decreases food intake in man. Am J Clin Nutr 34, 154–160.
- Kojima M, Hosoda H, Date Y, Nakazato M, Matsuo H & Kangawa K (1999) Ghrelin is a growth-hormone-releasing acylated peptide from stomach. *Nature* **402**, 656–660.
- Komaki G, Matsumoto Y, Nishikata H, Kawai K, Nozaki T, Takii M, Sogawa H & Kubo C (2001) Orexin-A and leptin change inversely in fasting non-obese subjects. *Eur J Endocrinol* **144**, 645–651.
- Kreymann B, Ghatei MA, Burnet P, Williams G, Kanse S, Diani AR & Bloom SR (1989) Characterization of glucagon-like peptide-1-(73-6)amide in the hypothalamus. *Brain Res* **502**, 325–331.
- Lambert PD, Wilding PH, Ghatei MA & Bloom SR (1994) A role for GLP-1-(73-6)NH2 in the central control of feeding behaviour. *Digestion* **54**, 360–361.
- Lieverse RJ, Jansen JBMJ, Zwan A, Samson L, Masclee AAM & Lamers CBHW (1993) Satiety effects of a physiological dose of cholecystokinin in humans. *Gut* 36, 176–179.
- Londos C, Gruia-Gray J, Brasaemle DL, Rondinone CM, Takeda T, Dwyer NK, Barber T, Kimmel AR & Blanchette-Mackie EJ (1996) Perilipin: possible roles in structure and metabolism of intracellular neutral lipids in adipocytes and steroidogenic cells. *Int J Obes Relat Metab Disord* **20**, Suppl. 3, S97–S101.
- Long SJ, Sutton JA, Amaee WB, Giouvanoudi A, Spyrou NM, Rogers PJ & Morgan LV (1999) No effect of glucagon-like peptide-1 on short-term satiety and energy intake in man. *Br J Nutr* **81**, 273–279.
- Lovshin J & Drucker DJ (2000) New frontiers in the biology of GLP-2. *Regul Pept* **90**, 27–32.
- MacDougald OA & Mandrup S (2002) Adipogenesis: forces that tip the scales. *Trends Endocrinol Metab* **13**, 5–11.
- Matsubara M, Maruoka S & Katayose S (2002) Inverse relationship between plasma adiponectin and leptin concentrations in normal-weight and obese women. *Eur J Endocrinol* **147**, 173–180.
- Mercer JG & Lawrence CB (1992) Selectivity of cholecystokinin (CCK) receptor antagonists, MK-329 and L-365,260, for axonally-transported CCK binding sites on the rat vagus nerve. *Neurosci Lett* **137**, 229–231.
- Morley JE, Levine AS, Grace M & Kneip J (1985) Peptide YY (PYY), a potent orexigenic agent. *Brain Res* **341**, 200–203.
- Nakazato M, Murakami N, Date Y, Kojima M, Matsuo H,

P. M. Hellström et al.

Kangawa K & Matsukura S (2001) A role for ghrelin in the central regulation of feeding. *Nature* **409**, 194–198.

S56

- Näslund E, Barkeling B, King N, Gutniak M, Blundell JE, Holst JJ, Rössner S & Hellström PM (1999*a*) Energy intake and appetite are suppressed by glucagon-like peptide-1 (GLP-1) in obese men. *Int J Obes Relat Metab Disord* **23**, 304–311.
- Näslund E, Bogefors J, Skogar S, Grybäck P, Jacobsson H, Holst JJ & Hellström PM (1999b) GLP-1 slows solid gastric emptying and inhibits insulin, glucagon, and PYY release in humans. *Am J Physiol* **277**, R910–R916.
- Näslund E, Ehrström M, Ma J, Hellström PM & Kirchgessner AL (2002) Localization and effects of orexin on fasting motility in the rat duodenum. *Am J Physiol* **282**, G470–G479.
- Näslund E, Grybäck P, Backman L, Jacobsson H, Holst JJ, Theodorsson E & Hellström PM (1998a) Distal small bowel hormones: correlation with fasting antroduodenal motility and gastric emptying. *Dig Dis Sci* **43**, 945–952.
- Näslund E, Gutniak M, Skogar S, Rossner S & Hellström PM (1998b) Glucagon-like peptide 1 increases the period of post-prandial satiety and slows gastric emptying in obese men. *Am J Clin Nutr* **68**, 525–530.
- Näslund E, King N, Mansten S, Adner S, Holst JJ, Gutniak M & Hellström PM (2004) Prandial subcutaneous injuctions of glucagon-like peptide-1 cause weight loss in obsess human subjects. *Br J Nutr* **91**, 439–446.
- Nauck MA, Niedereichholz U, Ettler R, Holst JJ, Örskov C, Ritzel R & Schmiegel WH (1997) Glucagon-like peptide 1 inhibition of gastric emptying outweighs its insulinotropic effects in healthy humans. *Am J Physiol* **273**, E981–E988.
- Nowak KW, Mackowiak P, Switonska MM, Fabis M & Malendowicz LK (2000) Acute orexin effects on insulin secretion in the rat: in vivo and in vitro studies. *Life Sci* **66**, 449–454.
- O'Rahilly S (2003) Leptin: defining its role in humans by the clinical study of genetic disorders. *Nutr Rev* **60**, 30–34.
- Ørskov C (1992) Glucagon-like peptide-1, a new hormone of the enteroinsular axis. *Diabetologia* **35**, 701–711.
- Otto B, Cuntz U, Fruehauf E, Wawarta R, Folwaczny C, Riepl RL, Heiman ML, Lehnert P, Fichter M & Tschop M (2001) Weight gain decreases elevated plasma ghrelin concentrations of patients with anorexia nervosa. *Eur J Endocrinol* **145**, 669–673.
- Ouedraogo R, Näslund E & Kirchgessner AL (2003) Glucose regulates the release of orexin-a from the endocrine pancreas. *Diabetes* **52**, 111–117.
- Pajvani UB, Du X, Combs TP, Berg AH, Rajala MW, Schulthess T, Engel J, Brownlee M & Scherer PE (2003) Structure-function studies of the adipocyte-secreted hormone Acrp30/adiponectin. Implications for metabolic regulation and bioactivity. *J Biol Chem* 278, 9073–9085.
- Poeschla B, Gibbs J, Simansky KJ, Greenberg D & Smith GP (1993) Cholecystokinin-induced satiety depends on activation of 5-HT1C receptors. Am J Physiol 264, R62–R64.
- Porte D, Baskin DG & Schwartz MW (2002) Leptin and insulin action in the central nervous system. *Nutr Rev* **60**, S20–S29.
- Ranganath L, Norris F, Morgan L, Wright J & Marks V (1999) Inhibition of carbohydrate-mediated glucagon-like peptide-1 (73-6)amide secretion by circulating non-esterified fatty acids. *Clin Sci* **96**, 335–342.
- Ranganath LR, Beety JM, Morgan LM, Wright JW, Howland R & Marks V (1996) Attenuated GLP-1 secretion in obesity: cause or consequence. *Gut* 38, 916–919.
- Raybould HE (1998) Does your gut taste? Sensory transduction in the gastrointestinal tract. *News Physiol Sci* 13, 275–280.
- Read N, French SA & Cunningham K (1994) The role of the gut in regulating food intake in man. *Nutr Rev* **52**, 1–10.
- Reidelberger RD, Varga G & Solomon TE (1991) Effects of

- selective cholecystokinin antagonists L364,718 and L365,260 on food intake in rats. *Peptides* **12**, 1215–1221.
- Reubi JC, Waser B, Laderach U, Stettler C, Friess H, Halter F & Schmassmann A (1997) Localization of cholecystokinin A and cholecystokinin B-gastrin receptors in the human stomach. *Gastroenterology* **112**, 1197–1205.
- Ritter RC, Covasa M & Matson CA (1999) Cholecystokinin: proofs and prospects for involvement in control of food intake and body weight. *Neuropeptides* **33**, 387–399.
- Sakurai T, Amemiya A, Ishii M, *et al.* (1998) Orexins and orexin receptors: a family of hypothalamic neuropeptides and G protein-coupled receptors that regulate feeding behavior. *Cell* **92**, 573–585.
- Sanaka M, Koike Y, Yamamoto T, Mineshita S, Yamaoka S, Hirama S, Tanaka H, Kuyama Y & Yamanaka M (1997) A reliable and convenient parameter of the rate of paracetamol absorption to measure gastric emptying rate of liquids. *Int J Clin Pharmacol Ther* 35, 509–513.
- Sanaka M, Kuyama Y & Yamanaka M (1998) Guide for judicious use of the paracetamol absorption technique in a study of gastric emptying rate of liquids. *J Gastroenterol* **33**, 785–791.
- Scherer PE, Williams S, Fogliano M, Baldini G & Lodish HF (1995) A novel serum protein similar to C1q, produced exclusively in adipocytes. *J Biol Chem* **270**, 26746–26749.
- Schmidt PT, Näslund E, Grybäck P, Jacobsson H, Hartmann B, Holst JJ & Hellström PM (2003) Peripheral administration of GLP-2 to humans has no effect on gastric emptying or satiety. *Regul Pept* **116**, 21–25.
- Schwartz GJ, McHugh PR & Moran TH (1993) Gastric loads and cholecystokinin synergistically stimulate rat gastric vagal afferents. Am J Physiol 265, R872–R876.
- Shapiro L & Scherer PE (1998) The crystal structure of a complement-1q family protein suggests an evolutionary link to tumor necrosis factor. *Curr Biol* **8**, 335–338.
- Shiiya T, Nakazato M, Mizuta M, Date Y, Mondal MS, Tanaka M, Nozoe S, Hosoda H, Kangawa K & Matsukura S (2002) Plasma ghrelin levels in lean and obese humans and the effect of glucose on ghrelin secretion. *J Clin Endocrinol Metab* 87, 240–244.
- Shimizu I, Hirota M, Ohboshi C & Shima K (1987) Identification and localization of glucagon-like peptide-1 and its receptor in rat brain. *Endocrinology* 121, 1076–1082.
- Sobhani I, Bado A, Vissuzaine C, et al. (2000) Leptin secretion and leptin receptor in the human stomach. Gut 47, 178–183.
- Sobhani I, Buyse M, Goiot H, Weber N, Laigneau JP, Henin D, Soul JC & Bado A (2002) Vagal stimulation rapidly increases leptin secretion in human stomach. *Gastroenterology* **122**, 259–263.
- Sørensen LB, Flint A, Raben A, Hartmann B, Holst JJ & Astrup A (2003) No effect of physiological concentrations of glucagon-like peptide-2 on appetite and energy intake in normal weight subjects. *Int J Obes* 27, 450–456.
- Stellar E (1954) The physiology of motivation. *Psychol Rev* **61**, 5–22.
- Steppan CM, Bailey ST, Bhat S, Brown EJ, Banerjee RR, Wright CM, Patel HR, Ahima RS & Lazar MA (2001) The hormone resistin links obesity to diabetes. *Nature* 409, 307–312.
- Takaya K, Ariyasu H, Kanamoto N, et al. (2000) Ghrelin strongly stimulates growth hormone release in humans. J Clin Endocrinol Metab 85, 4908–4911.
- Tang-Christensen M, Larsen PJ, Göke R, Fink-Jensen A, Jessop DS, Moller M & Sheikh SP (1996) Central administration of GLP-1-(7-36) amide inhibits food and water intake in rats. *Am J Physiol* **271**, R848–R856.
- Tang-Christensen M, Larsen PJ, Thulesen J, Romer J & Vrang N (2000) The proglucagon-derived peptide, glucagon-like pep-

- tide-2, is a neurotransmitter involved in the regulation of food intake. *Nat Med* **6**, 802–807.
- Tansey JT, Sztalryd C, Gruia-Gray J, et al. (2001) Perilipin ablation results in a lean mouse with aberrant adipocyte lipolysis, enhanced leptin production, and resistance to diet-induced obesity. Proc Natl Acad Sci USA 98, 6494–6499.
- Toft-Nielsen MB, Madsbad S & Holst JJ (1999) Continuous subcutaneous infusion of glucagon-like peptide-1 lowers plasma glucose and reduces appetite in type 2 diabetic patients. *Diabetes Care* 22, 1137–1143.
- Tolessa T, Gutniak M, Holst JJ, Efendic S & Hellström PM (1998) Glucagon-like peptide-1 retards gastric emptying and small bowel transit in the rat: effect mediated through central or enteric nervous mechanisms. *Dig Dis Sci* **43**, 2284–2290.
- Trayhurn P & Beattie JH (2001) Physiological role of adipose tissue: white adipose tissue as an endocrine and secretory organ. *Proc Nutr Soc* **60**, 329–339.
- Tsai CH, Hill M & Drucker DJ (1997) Biological determinants of intestinotrophic properties of GLP-2. *Am J Physiol* **272**, G662–G668.
- Tsao TS, Lodish HF & Fruebis J (2002) ACRP30, a new hormone controlling fat and glucose metabolism. *Eur J Pharmacol* **440**, 213–221.
- Tschop M, Smiley DL & Heiman ML (2000) Ghrelin induces adiposity in rodents. *Nature* **407**, 908–913.
- Tschop M, Weyer C, Tataranni PA, Devanarayan V, Ravussin E & Heiman ML (2001) Circulating ghrelin levels are decreased in human obesity. *Diabetes* **50**, 707–709.
- Turton MD, O'Shea D, Gunn I, *et al.* (1996) A role for glucagon-like peptide-1 in the central regulation of feeding. *Nature* **379**, 69–72.
- Ukkola O & Santaniemi M (2002) Adiponectin: a link between excess adiposity and associated comorbidities? J Mol Med 80, 696–702.

- Wettergren A, Schjoldager B, Mortensen PE, Myhre J, Christiansen J & Holst JJ (1993) Truncated GLP-1 (proglucagon 781-07-amide) inhibits gastric and pancreatic functions in man. *Dig Dis Sci* **38**, 665–673.
- Wettergren A, Wøjdemann M & Holst JJ (1998) Glucagon-like peptide-1 inhibits gastropancreatic function by inhibiting central parasympathetic outflow. *Am J Physiol* **275**, G984–G992.
- Wettergren A, Wøjdemann M, Meisner S, Stadil F & Holst JJ (1997) The inhibitory effect of glucagon-like peptide-1 (GLP-1) 7-36 amide on gastric acid secretion in humans depends on an intact vagal innervation. *Gut* **40**, 597–601.
- Wikberg JE, Muceniece R, Mandrika I, Prusis P, Lindblom J, Post C & Skottner A (2000) New aspects on the melanocortins and their receptors. *Pharmacol Res* **42**, 393–420.
- Wøjdemann M, Wettergren A, Hartmann B & Holst JJ (1998) Glucagon-like peptide-2 inhibits centrally induced antral motility in pigs. *Scand J Gastroenterol* **33**, 828–832.
- Wren AM, Seal LJ, Cohen MA, Brynes AE, Frost GS, Murphy KG, Dhillo WS, Ghatei MA & Bloom SR (2001) Ghrelin enhances appetite and increases food intake in humans. *J Clin Endocrinol Metab* **86**, 5992.
- Yanovski SZ (1993) Binge eating disorder: current knowledge and future directions. *Obes Res* 1, 306–324.
- Yanovski SZ (1995) Biological correlates of binge-eating. *Addict Behav* **20**, 705–712.
- Yeo GS, Farooqi IS, Aminian S, Halsall DJ, Stanhope RG & O'Rahilly S (1998) A frameshift mutation in MC4R associated with dominantly inherited human obesity. *Nat Genet* **20**, 111–112.
- Zhang Y, Proenca R, Maffei M, Barone M, Leopold L & Friedman JM (1994) Positional cloning of the mouse obese gene and its human homologue. *Nature* **372**, 425–432.